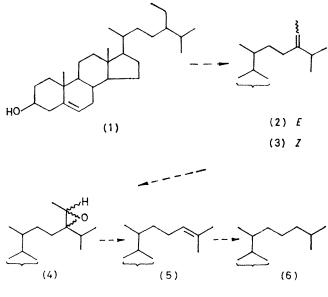
## Conversion of Isofucosterol-(24R,28S)-epoxide into Cholesterol in the Insect *Tenebrio molitor*

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Summary Tenebrio molitor larvae convert  $[7-^{3}H_{2}]-(24R, -28S)-24,28$ -epoxystigmast-5-en- $3\beta$ -ol, but not its (24S, -28R)-diastereoisomer, into cholesterol.

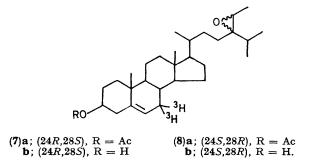
PHYTOPHAGOUS insects have developed a metabolic pathway to obtain cholesterol (6) from phytosterols, such as sitosterol (1), which they find in their diet.<sup>1</sup> The generally accepted scheme for this process<sup>2,3</sup> involves a 24(28)-double bond intermediate (2), which is transformed into cholesterol *via* epoxidation to (4), followed by elimina-



Scheme

tion of a C-2 unit to form desmosterol (5) and saturation of the 24(25)-double bond of this last compound (Scheme). In *Tenebrio molitor* this dealkylation process occurs with the formation of isofucosterol  $(3)^4$  as well as fucosterol. We now report the absolute stereochemistry of the epoxide involved in the transformation of isofucosterol into cholesterol in *Tenebrio molitor*.

Epoxidation<sup>5</sup> of  $[7^{-3}H_2]$ isofucosteryl acetate (synthesized according to the published procedure)<sup>6</sup> with *m*-chloroperbenzoic acid yielded a mixture of  $[7^{-3}H_2]$ -(24R, 28S)- and  $[7^{-3}H_2]$ -(24S, 28R)- $3\beta$ -acetoxy-24, 28-epoxystigmast-5-enes (7a) and (8a), which were separated by preparative t.l.c. (benzene-ethyl acetate 99:1, five elutions) and hydrolysed to the corresponding free sterols (7b) and (8b).<sup>5</sup> The



absolute configuration of (7a) and (8a) was determined from the corresponding 24-methoxy-28-alcohols obtained from each epoxide by acid catalysed methanolysis, using a g.l.c. modification<sup>6,7</sup> of Horeau's method.

A mixture of (7b)  $(1.23 \times 10^8 \text{ d.p.m.})$ , specific activity 24.8 mCi/mmol) and  $[4^{-14}\text{C}]$ sitosterol (Radiochemical Centre

TABLE Total radioactivity and <sup>3</sup>H/<sup>13</sup>C ratios of the administered precursors and of the isolated cholesterol

	Administered [7- <sup>3</sup> H.]epoxides	Administered [414C]sitosterol		Che	lesteryl acetate	
Erest			311 /140			911 /140
$\mathbf{Expt}$	dpm of <sup>3</sup> H	dpm of <sup>14</sup> C	3H/14C	арт ог чн	dpm of 14C	3H/14C
1	(7b) $1.23 \times 10^8$	$6.36 imes10^6$	193	$7.70 \times 10^{6}$	$2.18 \times 10^{5}$	35 3
2	$(8b)$ 1 33 $\times 10^{8}$	$9.14 imes10^6$	146	$7{\cdot}90 imes10^5$	$3 19 \times 10^{5}$	25

Amersham,  $6.36 \times 10^6$  d p m, sp act 53.6 mC1/mmol), deposited onto 320 mg of finely ground oatmeal, was fed to 225 young larvae (3 2 g) of Tenebrio molitor, which had been starved for 48 h (expt 1, see Table) After four days the larvae were sacrificed, macerated in ethanol, and submitted to mild alkaline hydrolysis (0.25% methanolic KOH) The unsaponifiable fraction was acetylated and separated by preparative tlc on silica gel (developed with benzeneethyl acetate 98:2) into two bands The less polar band corresponded to the acetates of sitosterol plus cholesterol, whereas the more polar band corresponded to the acetate of the unconverted epoxide The acetates of sitosterol and cholesterol were separated by preparative glc (25%)SE-30,  $T_c$  220 °C) Cholesteryl acetate was diluted with cold material and repeatedly crystallized to constant specific activity In expt 2, (8b)  $(1.33 \times 10^8 \text{ d p m})$ , sp act 24.8 mC1/mmol) and [4-14C](1) (9.14  $\times$  10<sup>6</sup> d p m ) deposited onto 300 mg of oatmeal were administered to 225

larvae (3 0 g) Cholesteryl acetate was isolated according to the procedure of expt 1 and crystallized to constant specific activity (see Table)

Comparison of the cholesteryl acetate <sup>3</sup>H/<sup>14</sup>C ratios in the two experiments clearly shows that the (24R, 28S)-epoxide (7b) is converted into (6) in ca ten times higher yield than its (24S, 28R)-isomer (8b), suggesting that (7b) is the actual precursor of cholesterol

It is interesting to note that in Bombyx mori fucosterol is epoxidized from the 24re,28si face,3 whereas in Tenebrio molitor epoxidation of isofucosterol occurs from the 24si,-28st face, this indicates a somewhat different steric approach of furosterol and isofucosterol molecules to the epoxidizing enzymatic system †

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† Added in proof Ikekawa and his colleagues have recently published a paper (Y Fujimoto, K Murakami, and N Ikekawa, J. Org Chem, 1980, 45, 566) in which the configuration previously assigned (ref 3) to the 24,28-fucosterol epoxides is reversed

<sup>1</sup> J A Svoboda and W E Robbins, Experientia, 1968, 24, 1131, Science, 1967, 156, 1637

<sup>2</sup> J A Svoboda and W E Robbins, Experientia, 1960, 24, 1101, 500, 1001, 1001, 1001, 1001, 1001, 210, 57, J P Allais and M Barbier, Experientia, 1971, 27, 506, M Morisaki H Ohtaka, M Okubayashi, N Ikekawa Y Horie, and S Nakasone, J Chem Soc, Chem Commun, 1972, 1275, J P Allais, A Alcaide, and M Barbier, Experientia, 1973, 29, 944 <sup>3</sup> S -M L Chen, K Nakanishi, N Awata, M Morisaki, N Ikekawa, and Y Shimizu, J Am Chem Soc, 1975, 97, 5297 <sup>4</sup> F Nicotra, F Ronchetti, and G Russo, Experientia, 1978, 34, 699 <sup>5</sup> F Nicotra, F Ronchetti, Chema Care Chem. 1101, 1080, 110, in the paper

<sup>6</sup> F Nicotra, F Ronchetti, G Russo, and L Toma, Gazz Chim Ital, 1980, 110, in the press <sup>6</sup> F F Knapp, L J Goad, and T W Goodwin, Phytochemistry, 1977, 16, 1683

<sup>7</sup>C J W Brooks and J D Gilbert, J Chem Soc, Chem Commun, 1973, 194.