Synthesis of Hepta- and Penta-saccharides, Part of the Complex-type Carbohydrate Portion of Glycoproteins

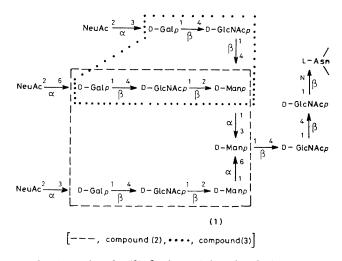
By Jan Arnarp and Jorgen Lonngren*

(Department of Organic Chemistry, Arrhenius Laboratory, University of Stockholm, S-106 91 Stockholm, Sweden)

Summary Silver trifluoromethanesulphonate-promoted condensation of 3,6-di-O-acetyl-4-O-(2,3,4,6-tetra-O-acetyl- β -D-galactopyranosyl)-2-deoxy-2-phthalimido- β -D-glucopyranosyl bromide with benzyl-3,6-di-O-benzyl- α -D-mannopyranoside and benzyl 2,4-di-O-benzyl-3,6-di-O-(3,4,6-tri-O-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside gave a protected pentasaccharide and a protected heptasaccharide in 56% and 43% yield, respectively, after deblocking the free penta- and heptasaccharides were obtained

The oligosaccharides that are N-glycosidically linked to L-asparagine residues in glycoproteins are of two major types, the 'high-mannose type' which contain D-mannosyl- and N-acetyl-D-glucosaminyl-residues and the 'complex type' which contain D-mannosyl-, D-galactosyl-, N-acetyl-D-glucosaminyl-, and sialyl-residues ¹ The 'complex type' has been found with different degrees of branching The N-glycosidically linked carbohydrate portion of fetuin (the predominant glycoprotein of fetal calf serum), which has the structure (1), ² is a representative example

These oligosaccharides are assumed to be involved in different biological functions¹ and the synthesis of oligosaccharides which form parts of these structures is a matter of some interest. We now report the synthesis of the reduc-

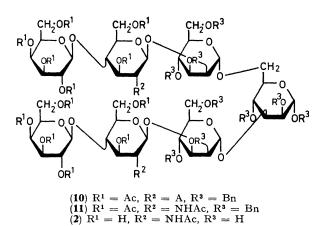


ing heptasaccharide (2) [indicated by the dashed line in structure (1)] and the reducing pentasaccharide (3) [indicated by the dotted line in structure (1)]

Hexa-O-acetyl-D-lactal³ (20 g) was subjected to azidonitration⁴ by treatment with ceric ammonium nitrate and sodium azide in acetonitrile to give, after work-up, a crystalline mixture (16 5 g) † According to the ¹H n m r spectrum

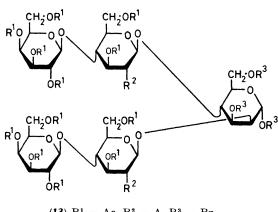
and to the sugar analysis⁵ of a sample that had been subjected to hydrogenation over palladium-charcoal, this mixture contained mainly compounds (4), (5), and (6) in the approximate proportions 1:4:8. The mixture was treated

sequentially with hydrogen over palladium-charcoal in ethyl acetate, phthalic anhydride (6 equiv.) in 90% aqueous ethanol adjusted to pH ca. 9 (r.t., 2 h), and acetic anhydridepyridine (r.t., 12 h; 100 °C, 1 h) to give, after silica gel chromatography and crystallization, (7); (m.p. 229-230 °C) and (8) (m.p. 263-265 °C). In pilot experiments compounds (7) and (8) were separately transformed into the same crystalline bromide (9) (m.p. 109-110 °C) by treatment with hydrogen bromide in methylene chloride (r.t., 4 h). For preparative purposes the crude mixture of (7) and (8) was used to give (9) [34% overall yield from a mixture of (4), (5) and (6)]. Sugar analysis of (9) showed D-galactose and Dglucosamine, but no D-mannosamine. The 1H n.m.r. spectrum of (9) showed, inter alia, a signal for the anomeric proton of the D-glucosaminyl residue at δ 6.39 (1 H, d, I 10 Hz) indicating that the β -bromide was obtained. Glycosidation with 3,4,6-tri-O-acetyl-2-deoxy-2-phthalimido-Dglucopyranosyl bromide6 in the presence of silver trifluoromethanesulphonate is known^{6,7} to give a high yield of β -glucoside and (9) could be expected to behave analogously. For the synthesis of compound (2), benzyl 2,4-di-Obenzyl-3,6-di-O-(3,4,6-tri-O-benzyl- α -D-mannopyranosyl)- α -D-mannopyranoside⁸ (0.22 mmol) was condensed with (9)



‡ Satisfactorily n m.r. data were obtained for all compounds.

(0.7 mmol) in methylene chloride, using silver trifluoromethanesulphonate-s-collidine (0.7 mmol) as a promotor? $(-50-+20 \, ^{\circ}\text{C} \, \text{for } 16 \, \text{h})$. Compound (10) $\{[\alpha]_{589} + 10^{\circ}$ (CHCl₃)}, isolated as a syrup in 43% yield after silica gel chromatography, was treated subsequently with sodium methoxide in methanol (r.t., 16 h), hydrazine hydrate⁷ (10 equiv. in boiling ethanol, 16 h), and acetic anhydridepyridine (r.t., 16 h) to give, after silica gel chromatography, compound (11) $\{ [\alpha]_{589} + 11^{\circ} (CHCl_{3}) \}$ in 55% yield. Compound (11) was finally subjected to de-O-acetylation and de-O-benzylation by catalytic hydrogenation (Pd-C catalyst) to give, after gel-filtration (Sephadex G-25), compound (2) $\{[\alpha]_{589} + 14^{\circ} (H_2O)\}$ in 46% yield. The ¹H n.m.r. spectrum (200 MHz, D₂O, 85 °C) of (2) showed, inter alia, signals for anomeric protons at δ 5.14 (1.7 H, J small; α -D-mannoseresidue and 1.2-linked α-D-mannosyl-residue), 4.89 (1.3 H. J small; β -D-mannose-residue and 1,2-linked α -D-mannosylresidue), 4.60 br (2 H, d; N-acetyl-β-D-glucosaminyl-residues), and 4.46 (2 H, d, J 7.5 Hz; β -D-galactosyl-residues). Methylation analysis of the alditol of (2) showed 2,3,4,6tetra-O-methyl-D-galactose, 3,4,6-tri-O-methyl-D-mannose, 3,6-di-O-methyl-N-methyl-N-acetyl-D-glucosamine, 1,2,4,5-tetra-O-methyl-D-mannitol.



Benzyl 3,6-di-O-benzyl- α -D-mannopyranoside (12) {[α]₅₈₉ + 42° (CHCl₃)} was prepared in 40% yield from benzyl α-D-mannopyranoside by tributylstannylation¹⁰ and benzylation. Compound (12) (0.38 mmol) and (9) (1.1 mmol) were condensed, as described above, to give (13) $\{[\alpha]_{589} + 12^{\circ}\}$ (CHCl₃) in 56% yield after silica gel chromatography. Compound (13) was then transformed into (14) $\{ [\alpha]_{589} - 1^{\circ} \}$ (CHCl₃) in 74% yield, as described for the corresponding transformation (10) to (11). Removal of the O-acetyl- and O-benzyl- groups of (14) gave (3) $\{ [\alpha]_{589} - 13^{\circ} (H_2O) \}$ in 61% yield. The ¹H n.m.r. spectrum (200 MHz, D₂O, 85 °C) of (3) showed, inter alia, signals for anomeric protons at $\delta 5.16$ (0.8 H, 4d, J 1.5 Hz; α -D-mannose-residue), 4.90 (0.2 H, 4d, J < 1 Hz; β -D-mannose-residue), 4.58 br (2 H, d; Nacetyl-β-D-glucosaminyl-residues), and 4.47 (2 H, d, J 7.5 Hz; β -D-galactosyl-residues). Methylation analysis of the alditol of (3) showed 2,3,4,6-tetra-O-methyl-D-galactose.

3,6-di-O-methyl-N-methyl-N-acetyl-D-glucosamine, and 1,3,5,6-tetra-O-methyl-D-mannitol

The ¹H n.m r. data given above for (2) and (3) are in good agreement with those reported for related natural oligosaccharides 11

We thank Professors Per J Garegg and Bengt Lindberg for their interest and the Swedish Natural Science Research Council for financial support

(Received, 21st July 1980, Com. 788)

- ¹ R G Spiro, Adv Protein Chem, 1973, 27, 349

- ¹ R G Spiro, Adv Protein Chem, 1973, 27, 349
 ² B Nilsson, N E Nordén, and S Svensson, J Biol Chem, 1979, 254, 4545
 ³ W N Haworth, E L. Hirst, M M T Plant, and R J W Reinolds, J Chem Soc, 1930, 2647
 ⁴ R U Lemieux and R M Ratcliffe, Can J Chem, 1979, 57, 1244
 ⁵ J S Sawardeker, J H Sloneker, and A R Jeanes, Anal Chem, 1965, 37, 1602
 ⁶ R U Lemieux, T Takeda, and B Y Chung, Am Chem Soc Symposium Series, 1976, 39, 90
 ⁷ D R Bundle and S Josephson, J Chem Soc, Perkin Trans 1, 1979, 2736
 ⁸ J Arnarp and J Lonngren, Acta Chem Scand, Ser B, 1978, 32, 696
 ⁹ P E Jansson, L Kenne, H Liedgren, B Lindberg, and J Lonngren, Chem Commun, Univ Stockholm, 1976, 8
 ¹⁰ T Ogawa and M Matsui, Carbohydr Res, 1978, 62 Cl
 ¹¹ L Dorland, J Haverkamp, J F G Vliegenthart, G Strecker, J-C Michalski, B Fournet, G Spik, and J Montreuil, Eur J Biochem, 1978, 87, 323