

Mode of Enzymic Oxygenation at Primary Carbon Atoms: Stereochemistry of Hydroxylation of C-1 Chiral Octanes by *Pseudomonas oleovorans*†

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Summary It was shown, using (1*R*)- and (1*S*)- [1-³H,²H,-¹H;¹⁴C]octanes, that C-1 hydroxylation by *P. oleovorans* strain TF4-1L proceeds with retention of configuration.

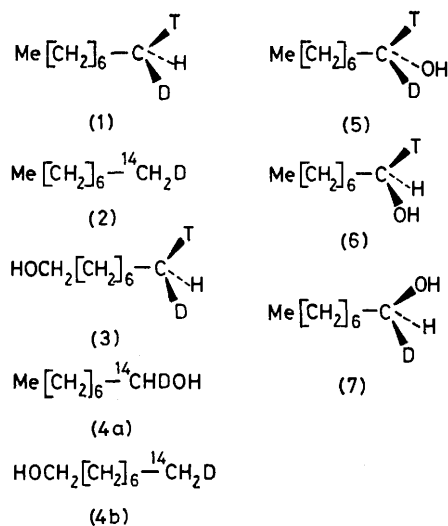
In contrast to numerous reports on the stereochemistry of enzymic hydroxylation of unactivated secondary¹ and tertiary² carbon atoms, no published information is available on the stereochemistry of hydroxylation of primary carbon atoms.³ The starting materials (1*R*)- and (1*S*)-[1-

³H,¹H;¹⁴C]octan-1-ols, were synthesized essentially as previously described.⁴ The (1*R*)-alcohol was at least 95–96% pure, since on oxidation with horse liver alcohol dehydrogenase [HLAD, EC.1.1.1.1] and NAD it lost 95–96% of tritium. By implication the parent⁴ (1*S*)-octanol must contain at least 95–96% of (1*S*)-alcohol. The (1*R*)- and (1*S*)-octanols were mesylated, then hydrogenolysed (lithium triethylborodeuteride in diglyme) to yield (1*S*)-[1-³H,²H,-¹H;¹⁴C]octane {spec. act. *ca.* 45 μCi ³H μl⁻¹ (7.3 mCi ³H

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mmol⁻¹), [³H:¹⁴C] ratio 8.47:1} and (1*R*)-[1-³H,²H,¹H;¹⁴C]-octane {spec. act. *ca.* 45 μCi ³H μl⁻¹ (7.3 mCi ³H mmol⁻¹), [³H:¹⁴C] ratio 8.96:1}, respectively.†

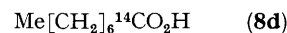
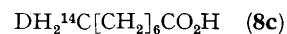
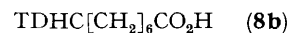
We found that the method of determining the stereochemical purity of (1*S*)-octanol by oxidation with horse liver, yeast, or *P. oleovorans* strain TF4-1L alcohol dehydrogenase and measuring the [³H:¹⁴C] ratio of the resulting aldehyde is not applicable. Apparently the [1-³H]octanal produced (RC³HO) is oxidized enzymically to octanoic acid⁸ at a considerably *slower rate* than is R¹⁴CHO. Consequently, the [³H:¹⁴C] ratio of the aldehydes isolated at a given point in time (from 7 min to 24 h) was higher (up to 40%) than that of the substrate (1*S*)-alcohol.



We then investigated the Günther, Simon *et al.*⁸ equilibration method, which is presumed to exchange the (1*R*)-hydrogen atom exclusively.^{8,7} In our hands, both the (1*R*)- and (1*S*)-hydrogen atoms of [1-³H]octanols were exchanged, although the (1*S*)-hydrogen was exchanged at a considerably slower rate. Under the conditions employed by us the (1*R*)-octanol lost all the tritium within 24 h, while the (1*S*)-octanol consistently and reproducibly lost 30–40% of tritium, and (1*RS*)-[1-³H;¹⁴C]octanol lost 65–70% of tritium. Based on these observations, determination of the overall chirality of the C-1 alcohols obtained by enzymic hydroxylation of C-1 chiral octanes was possible.

Assuming that hydroxylation involves a significant isotope effect $k_H > k_D > k_T$ and proceeds with retention of configuration, then (1*R*)-octane (1) should yield mainly (3) and lesser amounts of products of oxygenation of the chiral methyl [(5) > (6) > (7)]. For determination of the stereochemistry of the hydroxylation reaction, only the chirality of the major component of the mixture of C-1 tritiated alcohols (5) and (6) need be considered. Should hydroxylation of (1*R*)-octane proceed with inversion of configuration, then (1*R*)-octanol will be the major product. Similarly, hydroxylation of (1*S*)-octane with retention of configuration will yield mainly (1*R*)-octanol, whereas hydroxylation with inversion will yield mainly (1*S*)-octanol.

Chiral [1-³H,²H,¹H]octanes containing the [1-¹⁴C]octane (2) were incubated with homogenates of *P. oleovorans* strain TF4-1L,⁸ and the octanols from each incubation were recovered and purified to yield *ca.* 0.75 μCi of [³H]octanol per incubation (*ca.* 1.7% ³H recovered as octanol) admixed with [1-¹⁴C]octanols (4). The extent of hydroxylation of the chiral methyls was determined by oxidizing aliquots of the biosynthesized alcohols with Jones' reagent and counting the obtained acids (8) as their *p*-toluidides.



The Günther, Simon *et al.* exchange reactions were carried out in parallel using octanols derived enzymically from (1*R*)- and (1*S*)-octanes and synthetic (1*RS*)-octanol. The tritium lost in the equilibrations of the biosynthesized alcohols was corrected for the accompanying loss of (1*S*) tritium.§ From the difference between the tritium content at C-1 before equilibration and the *corrected* amount of tritium abstracted during equilibration, the fraction of (1*R*)-octanol in the biosynthetic mixtures was calculated.

The results are summarized in the Table. It is evident that hydroxylation of the octanes occurred mainly (70–80%) at the achiral methyl (column 3, Table). This indicates that enzymic hydroxylation involves a normal hydrogen isotope effect.

The hydroxylations of (1*R*)-octane gave mixtures of C-1 tritiated octanols. Equilibration of these mixtures of alcohols proceeded with the (corrected) loss of 37–38% of the tritium at C-1. It follows that 37–38% of the octanol in the mixtures was (1*R*)-octanol and, hence, the major

TABLE. Hydroxylation of (1*R*)- and (1*S*)-[1-³H,²H,¹H;¹⁴C]octanes by homogenates of *Pseudomonas oleovorans* strain TF4-1L.

Experiment	Chirality of octane	% ³ H at C-1 of octanols	% of (1 <i>R</i>)-octanol ^a	C-1 chirality of the major octanol	Stereochemistry of hydroxylation	
1	{	1 <i>R</i>	27	38	1 <i>S</i>	Retention
		1 <i>S</i>	21	82	1 <i>R</i>	Retention
2	{	1 <i>R</i>	29	37	1 <i>S</i>	Retention
		1 <i>S</i>	29	74	1 <i>R</i>	Retention

^a The average deviation in the [³H:¹⁴C] ratios from successive crystallizations of the acid and alcohol derivatives is ±0.05. The resulting relative error in the calculated (ref. 9) % of (1*R*)-octanol determination is ±7% (*e.g.* 38 ± 2.7%).

† The assignment of the configuration of the chiral octanes rests on the known inversion of configuration in the hydrogenolysis of methanesulphonyl esters.

§ The correction was computed on the basis of the amount of tritium lost from (1*S*)-octanol in the control equilibration of (1*RS*)-octanol.

product of hydroxylation of the chiral methyl terminus of (1*R*)-octane is (1*S*)-octanol. In contrast, hydroxylations of (1*S*)-octane gave octanols which on equilibration lost most (74—82%) of the tritium present at C-1. Therefore, the major product of hydroxylation of the chiral terminus of (1*S*)-octane is (1*R*)-octanol. These results are consistent with the view that the hydroxylation proceeds with retention of configuration in which mainly a hydrogen atom and, to a lesser extent, a deuterium atom is displaced.

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