

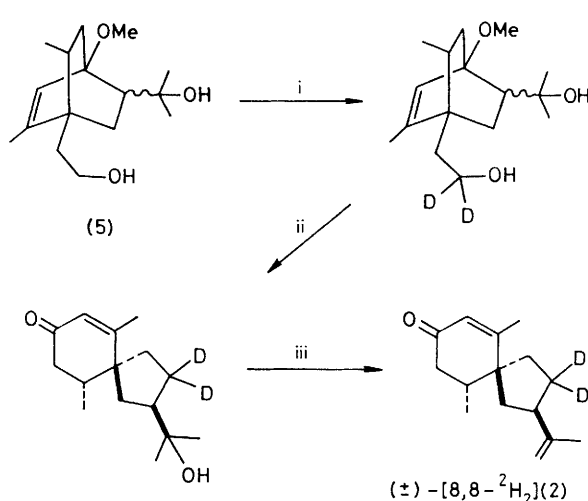
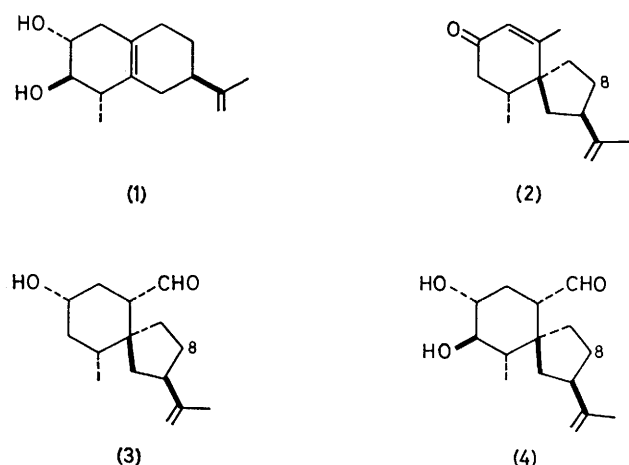
Biosynthesis from Solavetivone of the Phytoalexin Rishitin in Potato. Implicit Role of Solavetivone as an Activator

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The biosynthetic production of rishitin from solavetivone *via* lubimin and oxylubimin has been established on the basis of experiments with (\pm)-[8,8-²H₂]solavetivone.

Since the isolation of rishitin (1),¹ many sesquiterpenoid phytoalexins (stress compounds) have been isolated from infected potatoes, and attempts to elucidate the biosynthetic pathways leading to (1) have been made by several groups.²⁻⁴ Stoessel *et al.*^{3b} proposed that (1) and three major spirovetivane phytoalexins, solavetivone⁵ (2), lubimin⁶ (3), and oxylubimin⁶ (4), would be separately biosynthesized from farnesyl pyrophosphate. Kalan *et al.*^{4b} and we² reported almost the same biogenetic pathway, in which (2) would be transformed into (1) *via* (3) and (4) successively *in vivo*. While this hypothesis is based on our experiments⁷ with ¹⁴C-labelled (4), indicating that (4) is an intermediate on the main pathway to (1), the requisite studies with labelled (2) have not been performed yet. In this communication we present evidence confirming



Scheme 1. i, Jones oxidation, then CH₂N₂, then LiAlD₄, 99% yield, ii, MeSO₂Cl, -78 °C, then (CO₂H)₂, 33% aq. acetone, 85 °C, 4 h, 84% yield, iii, py-Al₂O₃, 220 °C, 8 min, 64% yield.

our proposed biogenesis on the basis of the experiments with (\pm)-[8,8-²H₂]solavetivone.

The starting material, (\pm)-[8,8-²H₂](2), was prepared from bicyclo-octenediol⁸ (5) as shown in Scheme 1 and showed spectra which differed markedly from those of natural (2) only in the following: *m/e* 220 (*M*⁺, 57%) and 218 (*ca.* 0%) ([8,8-²H₂]-content, *ca.* 100%); ν_{\max} 2200 and 2100 cm⁻¹. Thin slices of aged potato (Rishiri, *Solanum tuberosum* × *S. demissum*) were incubated with (\pm)-[8,8-²H₂](2) and related compounds, and extracted with methanol-chloroform (1:1). The chloroform extracts were fractionated sequentially by column and thin-layer chromatography, giving the deuteriated products shown in Table 1. These stress compounds were not detected on a thin-layer chromatogram of the corresponding chloroform extracts of the aged potato slices treated only with water-methanol (1:1) in the same manner. In view of the fact that substitution of the hydrogen atoms at C-8 by deuterium atoms affects the optical rotations of the respective compounds to some extent, the results summarized in Table 1 indicate that (i), ($-$)-[8,8-²H₂](2) and ($+$)-[8,8-²H₂](3) are incorporated efficiently into potato slices, (ii), natural ($-$)-[8,8-²H₂](2) is metabolized into ($+$)-[8,8-²H₂](4) *via* ($+$)-[8,8-²H₂](3), while synthetic ($+$)-[8,8-²H₂](2) does not undergo such enzymatic reactions, (iii) the [8,8-²H₂]-content of the stress metabolites decreases slightly (but definitely) (2-7%) on incubation of (\pm)-[8,8-²H₂](2) (run 1), but remains unchanged on that of ($+$)-[8,8-²H₂](3) (run 3). In summary, the present result, coupled with our previous one,⁷ establishes the main biosynthetic pathway leading to (1), the sequence ($-$)-(2) → ($+$)-(3) → ($+$)-(4) → ($-$)-(1), and also suggests that natural ($-$)-(2) might be an activator of the

Table 1. Transformation of (\pm)-[8,8-²H₂](2) and related compounds in potato tubers.^a

Run	Precursors	[8,8- ² H ₂]-Products ^b	Yield ^c	[α] _D ^d	[² H ₂]-content ^e
1	(\pm)-[8,8- ² H ₂](2) [α] _D ± 0, (<i>ca.</i> 100%) ^g	($+$)-(2)	74% ^f	+82 (-119)	<i>ca.</i> 100%
		($+$)-(3)	29% ^h	+31 (+36)	97.8%
		($+$)-(4) ⁱ	7% ^h	+17 (+22)	97.7%
		($-$)-(1)	10% ^h	-20 (-35)	93.5%
2	($+$)-[8,8- ² H ₂](2) [α] _D + 82, (<i>ca.</i> 100%) ^g	($+$)-(2)	71%	+71 (+119)	99.0%
		($+$)-(3)	9%	+42 (+36)	97.7%
3	($+$)-[8,8- ² H ₂](3) [α] _D + 31, (97.8%) ^g	($+$)-(3)	9%	+42 (+36)	97.7%
		($+$)-(4) ⁱ	11%	+12 (+22)	97.8%
		($-$)-(1)	10%	-30 (-35)	97.2%

^a All reactions were carried out by incubation of the precursors in water-methanol (1:1) to thin slices of aged potato tubers at 23-24 °C for 6 h. ^b The known phytoalexins other than the products described in Table 1 could not be detected on a thin-layer chromatogram. ^c Isolated yields. ^d Measured in ethanol. The figures in parentheses denote [α]_D of the corresponding unlabelled samples. ^e Determined by comparison of intensities of peaks corresponding to (*M*⁺) and (*M*⁺ - 2) in the respective mass spectrum. ^f Based on the ($+$)-enantiomer. ^g [8,8-²H₂]-content. ^h Based on the ($-$)-enantiomer. ⁱ Isolated as its diacetate.

enzyme system for formation of these stress metabolites from acetic acid.

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