

Hydrogen Rearrangements in the Biosynthesis of the Sesquiterpenoid, Alliacolide

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The occurrence of hydrogen rearrangements to C-1 and from C-5 to C-6 during the biosynthesis of alliacolide has been demonstrated by ²H n.m.r. experiments.

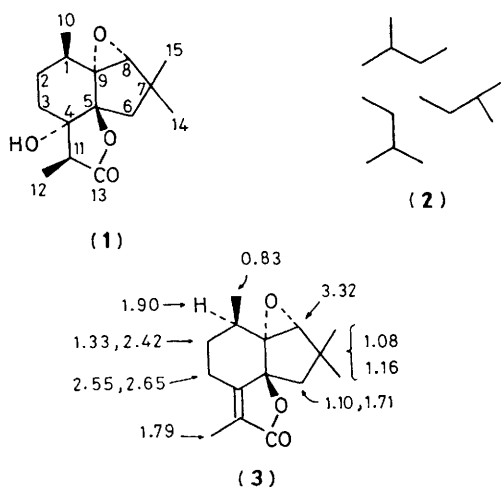
Alliacolide (1) is one of a group of biologically active metabolites of the Basidiomycete, *Marasmius alliaceus*.¹⁻³ Recently we have shown⁴ that the labelling pattern from [1-¹³C]- and [1,2-¹³C]-acetate is consistent with the formation of the sesquiterpenoid carbon skeleton from the three isoprene units as shown in (2). Examination of the mevalonoid hydrogen labelling pattern now reveals that a number of rearrangements are involved in this biosynthesis.

[4(*R*)-4-³H,2-¹⁴C]- and [5-³H₂,2-¹⁴C]-mevalonic acids (MVA's) were fed to *Marasmius alliaceus* after 14 days growth. The alliacolide (1) was isolated after a further 21 days and converted into dehydroalliacolide (3). The ³H:¹⁴C results are given in Table 1. Two of the three centres (C-4 and C-9) which would be expected to be labelled by [4(*R*)-4-³H]MVA are fully substituted and hence, in order to account for the incorporation of two labels, one *pro*-4(*R*)-

mevalonoid hydrogen atom must be involved in a rearrangement. The site of the 4-mevalonoid labels was determined using [4-²H₂]mevalonate.

Careful proton spin decoupling studies at 360 MHz enabled the proton resonances of dehydroalliacolide to be assigned [see (3)]. Signals in the ²H n.m.r. spectrum at δ 1.83 and 1.03, corresponding to 1-H and 6-H respectively, were found to be enriched in samples prepared biosynthetically from [4-²H₂]mevalonate. C-1 was thus the terminus of a hydrogen rearrangement involving a *pro*-4(*R*)-mevalonoid hydrogen atom.

One of the centres (C-5) which would be expected to be labelled by a [5-³H]mevalonate is fully substituted and another (C-8) bears only one hydrogen atom. Therefore in order to account for the incorporation of the fourth [5-³H]-mevalonoid label, it must also be involved in a rearrange-



^1H N.m.r. signals (from Me_2Si) of dehydroalliacolide were determined in CDCl_3 at 360 MHz.

Table 1. Incorporation of mevalonates into alliacolide (1) and dehydroalliacolide (3).

	[4(R)-4- ^3H ,2- ^{14}C]MVA	[5- $^3\text{H}_2$,2- ^{14}C]MVA
$^3\text{H}:^{14}\text{C}$ ratio in MVA	12.98:1	22.6:1
Atom ratio	3:3	6:3
Quantity fed ($\mu\text{Ci } ^{14}\text{C}$)	11.8	13.15
$^3\text{H}:^{14}\text{C}$ ratio in (1)	8.22:1	14.93:1
Atom ratio	1.90:3	3.96:3
% Incorporation (^{14}C)	0.99	1.59
$^3\text{H}:^{14}\text{C}$ ratio in (3)	8.14:1	15.09:1
Atom ratio	1.9:3	4:3

ment. Since there was no change in the $^3\text{H}:^{14}\text{C}$ atom ratio in the formation of dehydroalliacolide (3), the extra label is not at C-11 (*cf.* avocettin biosynthesis).⁵ The sites of labelling were located by feeding [$5\text{-}^3\text{H}_2$]mevalonate to *Marasmius*

alliaceus. Signals assigned to 3-H (α and β) (δ 2.62), 6-H (δ 1.71), and 8-H (δ 3.34) in the ^3H n.m.r. spectrum of the resultant dehydroalliacolide were enriched. Hence the rearrangement has led to a label at C-6. This rearrangement may involve a 1,2-shift (C-5 \rightarrow C-6) or a 1,3-shift (C-8 \rightarrow C-6). The former occurs within an isoprene unit and may therefore be distinguished from the latter by the use of [$5\text{-}^2\text{H}_2,4\text{-}^{13}\text{C}$]mevalonate. In the case of a 1,2-shift, a $^3\text{H}:^{13}\text{C}$ coupling will be generated whilst, provided there is a sufficient dilution with unlabelled material, a 1,3-shift will occur between labelled and unlabelled isoprene units and no such coupling will result. Since deuterium has a spin of 1 and carbon-13 a spin of $\frac{1}{2}$, these couplings are more easily observed in the deuterium spectrum. The dehydroalliacolide (3) derived from [$5\text{-}^2\text{H}_2,4\text{-}^{13}\text{C}$]mevalonate, showed a doublet, J 21 Hz, in place of a singlet at δ 1.71 in accordance with a 1,2-shift.

Alliacolide joins the increasing number of terpenoid substances in which a secondary methyl group has been shown to be generated by a hydrogen rearrangement.⁶ Furthermore many sesquiterpenoids derived from the Basidiomycetes contain a cyclopentane ring bearing a methylene group which is probably labelled by C-4 of mevalonate. The present results suggest these methylenes might also be generated by a hydrogen shift.

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