

Variation of $^1J(C1', H1')$ with Glycosidic Bond Conformation of Pyrimidine Nucleosides

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$^1J(C1', H1')$ magnitudes of a series of pyrimidine cyclonucleosides are found to vary quantitatively with glycosidic bond conformation. Measurements of $^1J(C1', H1')$ in pyrimidine nucleosides and nucleotides can be used to determine the range of allowed *syn* and *anti* conformations.

The glycosidic bond *syn* \rightleftharpoons *anti* conformer equilibrium of base with respect to sugar ring is of fundamental importance in determining the conformations of nucleic acids in solution. The majority of X-ray structure determinations of nucleosides, nucleotides, and oligo- and poly-nucleotides in the solid state show that the purine or pyrimidine ring exists in the *anti* conformation and that the *syn* conformation is promoted when the base ring contains a bulky substituent (at C8 for purine and C6 for pyrimidine).^{1,2} Many n.m.r. methods (chemical shift changes, lanthanide ion probe techniques, vicinal carbon-proton coupling constants, nuclear Overhauser enhancements, and proton spin-lattice relaxation times)³ have been used to investigate the glycosidic bond conforma-

tions in a qualitative manner. Although attempts have been made, especially with purine derivatives, to provide a quantitative assessment of the *syn* \rightleftharpoons *anti* equilibrium there are large discrepancies between results determined by different methods.⁴ A major limitation for comparison of such results is an independent assessment of the *syn* and *anti* conformations for molecules in solution. In this work the variation in $^1J(C1', H1')$ magnitudes of pyrimidine nucleosides is investigated and interpreted in terms of the allowed glycosidic bond conformations in equilibrium.

A number of cyclonucleosides (Figure 1) have been synthesised as model compounds in which the base ring exhibits different glycosidic bond conformations, charac-

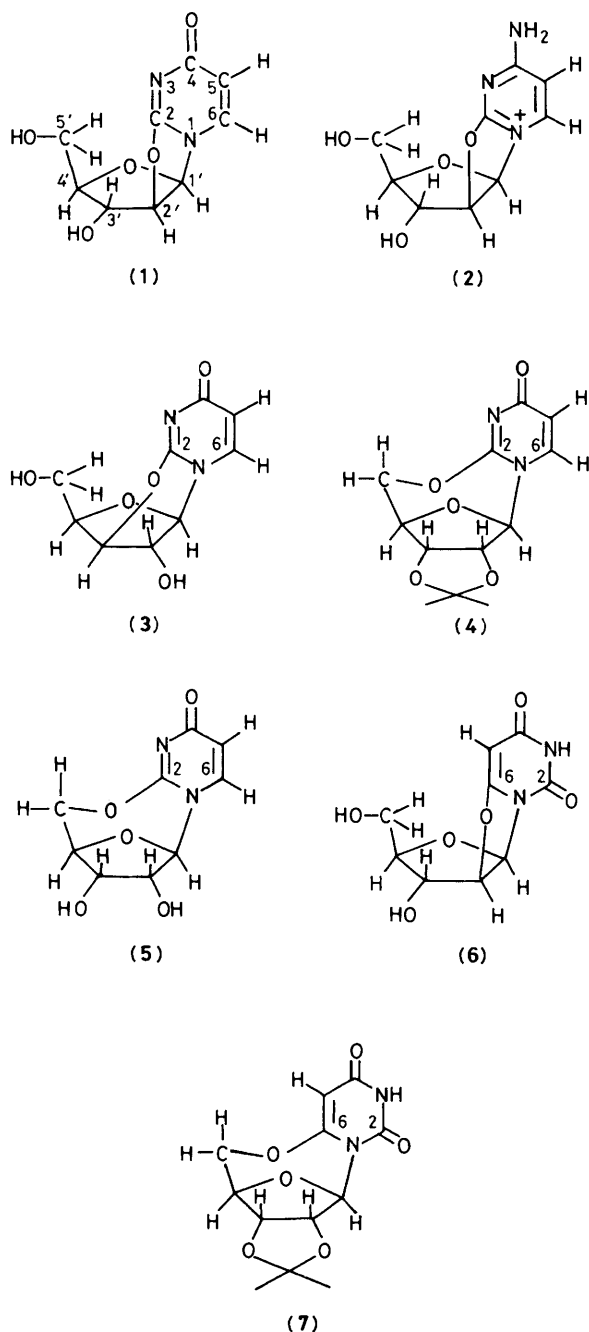
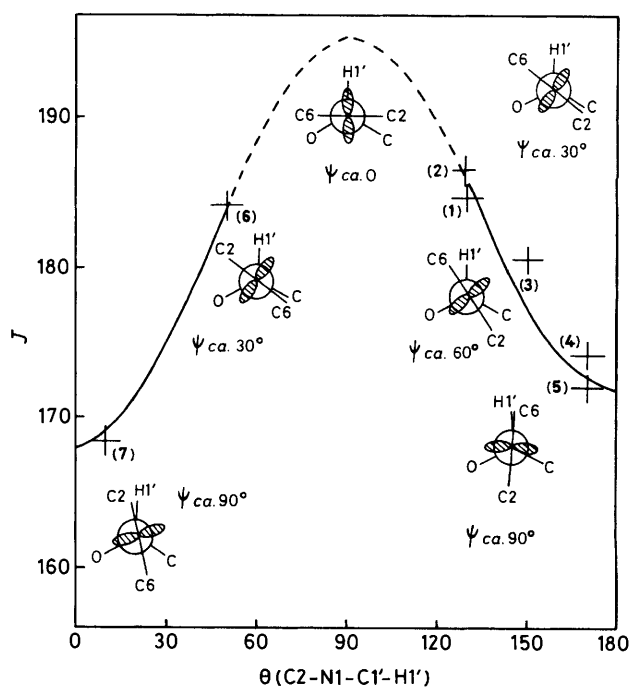


Figure 1. Structures of cyclonucleosides.

	χ°	$^1J(\text{C}1', \text{H}1')$ /Hz
(1) 2,2'-Anhydro-1-(β -D-arabinofuranosyl)-uracil	115	184.7
(2) 2,2'-Anhydro-1-(β -D-arabinofuranosyl)-cytosine	115	186.6
(3) 2,3'-Anhydro-1-(β -D-xylofuranosyl)-uracil	85	180.5
(4) 2,5'-Anhydro-2',3'-O-isopropylidene-uridine	65	174.2
(5) 2,5'-Anhydro-1-(β -D-ribofuranosyl)-uracil	65	172.0
(6) 2',6'-Anhydro-1-(β -D-arabinofuranosyl)-6-hydroxyuracil	290	184.3
(7) 5',6'-Anhydro-2',3'-O-isopropylidene-uridine	245	168.5

Figure 2. Variation of $^1J(\text{C}1', \text{H}1')$ of compounds (1)–(7) with dihedral angle θ ($\text{C}2\text{--N}1\text{--C}1'\text{--H}1'$). Inset structures show the approximate relation of the $\text{C}1'\text{--H}1'$ bond to the nitrogen atom p orbital (denoted by angle Ψ).

terised by angle χ .⁵ Two conformational regions are denoted as *syn* (χ , $0 \pm 90^\circ$) and *anti* (χ , $180 \pm 90^\circ$) conformations. Cyclisation from base to sugar ring produces relatively rigid molecules in which χ can be taken from crystal structures when available [(1) χ 114.5,^{6,7} (4) χ 66.4,⁶ 71.2⁸] or estimated from molecular models [(2) χ ca. 115, (3) χ ca. 85, (5) χ ca. 65, (6) χ ca. 290, (7) χ ca. 245°]. $^1J(\text{C}1', \text{H}1')$ magnitudes were determined for compounds (1)–(7) from proton-coupled ^{13}C n.m.r. spectra (observed under gated decoupling conditions to a data resolution of 0.3 Hz per point) and a variation from 168.5 Hz (7) to 184.7 Hz (1) is observed. It is known that magnitudes of $^1J(\text{C}, \text{H})$ vary markedly with s character of the carbon atom to give typical values for sp^3 , sp^2 , and sp carbon atoms which are modified by variations in substituent electronegativity, bond angle effects, etc.⁹ Also lone pair effects have been used to explain differences in the $^1J(\text{C}, \text{H})$ magnitudes of axial and equatorial C–H in carbohydrates,¹⁰ variations in $^1J(\alpha\text{C}, \text{H})$ of linear and cyclic peptides,¹¹ and the fact that the larger coupling constant occurs for a C–H *cis* to the nitrogen lone pair rather than *trans*.¹² Hence we might expect $^1J(\text{C}1', \text{H}1')$ to depend on both the glycosidic bond ($\text{N--C}1'$) and sugar ring ($\text{O}4'\text{--C}1'$) conformations.

For the cyclonucleosides studied here, only small variations in sugar ring conformations [*viz.* $\text{C}4'\text{--O}4'\text{--C}1'\text{--H}1'$ has magnitudes ca. 130 (1), (2), (6), ca. 150 (4), (5), (7), and ca. 160° (3)] are estimated in comparison to the glycosidic bond conformation [*viz.* $\text{C}2\text{--N}1\text{--C}1'\text{--H}1'$ has magnitudes ca. 10 (7), ca. 50 (6), ca. 130 (1), (2), ca. 150 (3), and ca. 170° (4), (5)] and so, to a first approximation, the variation in $^1J(\text{C}1', \text{H}1')$ magnitudes with conformation has been fitted to torsion angle θ ($\text{C}2\text{--H}1'$) in a generalised equation of the form of equation (1) with the magnitudes of the constants being $A = -26$, $B = -2$, and $C = 196$ Hz.

$$^1J(\text{C}1', \text{H}1') = A \cos^2\theta + B \cos\theta + C \quad (1)$$

As shown in Figure 2, the calculated curve fits the observed 1J magnitudes and predicts a minimum in $^1J(C1', H1')$ for conformations corresponding to $\theta = 0^\circ$ (1J ca. 168 Hz, C2–N1 eclipsed with C1'–H1') and $\theta = 180^\circ$, (1J ca. 172 Hz, C6–N1 eclipsed with C1'–H1') which is verified by compounds (4), (5), and (7). The curve also predicts a maximum of $^1J(C1', H1')$ of ca. 196 Hz for $\theta = 90^\circ$ where both C2–N1 and C6–N1 are perpendicular to C1'–H1'; in this conformation the p orbital of the trigonal nitrogen atom (involved in weak π bonding with adjacent C2 and C6) is coplanar with the C1'–H1' bond and may be responsible for the increase in $^1J(C1', H1')$.

In principle the variation in $^1J(C1', H1')$ with θ may be used to determine the glycosidic bond conformational angle χ , though in practice observation of one coupling constant gives rise to four possible χ values (two *syn* and two *anti*). Magnitudes of $^1J(C1', H1')$ of a number of pyrimidine nucleosides and nucleotides in solution¹³ are found to vary between 168 and 172 Hz which limits the glycosidic bond to a narrow range of *syn* ($\chi 60 \pm 10^\circ$ corresponding to $\theta 180 \pm 10^\circ$) and *anti* ($\chi 240 \pm 20^\circ$ corresponding to $\theta 0 \pm 20^\circ$) conformational regions, assuming that variations in sugar ring conformation have only minor effects on $^1J(C1', H1')$ magnitudes.† Although the *syn* and *anti* conformations determined from $^1J(C1', H1')$ magnitudes are consistent with those observed in the solid state by X-ray crystallography,^{1,2} determination of the position of the *syn* \rightleftharpoons *anti* equilibrium requires additional information such as vicinal proton–carbon coupling con-

stants.^{13,14} It is also expected that $^1J(C1', H1')$ magnitudes of purine nucleosides will vary with conformation in a manner analogous to that found for pyrimidine nucleosides, providing a new and general method for determination of glycosidic bond conformations of nucleosides and nucleotides in solution.

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References

- 1 M. Sundaralingam, in 'Conformations of Biological Molecules and Polymers,' eds. E. D. Bergmann and B. Pullman, Academic Press, New York, 1973, pp. 417–455.
- 2 W. Saenger, *Angew. Chem., Int. Ed. Engl.*, 1973, **12**, 135.
- 3 D. B. Davies, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1979, **12**, 135.
- 4 R. Stolarski, L. Dudycz, and D. Shugar, *Eur. J. Biochem.*, 1980, **108**, 111.
- 5 Abbreviations and symbols for the description of conformations of polynucleotide chains: *Eur. J. Biochem.*, 1983, **105**, 9.
- 6 L. T. J. Delbaere, M. N. G. James, and R. U. Lemieux, *J. Am. Chem. Soc.*, 1973, **95**, 7866.
- 7 D. Suck and W. Saenger, *Acta Crystallogr., Sect. B*, 1973, **29**, 1323.
- 8 P. C. Manor, W. Saenger, D. B. Davies, K. Jankowski, and A. Rabczenko, *Biochim. Biophys. Acta*, 1974, **340**, 472.
- 9 P. E. Hansen, *Prog. Nucl. Magn. Reson. Spectrosc.*, 1981, **14**, 175.
- 10 B. Coxon, *Dev. Food Carbohydr.*, 1980, **2**, 351.
- 11 H. Egli, U. Vögeli, and W. von Philipsborn, in 'Nuclear Magnetic Resonance Spectroscopy in Biology,' ed. B. Pullman, D. Reidel, Dordrecht, 1978.
- 12 G. van Binst and D. Tourwe, *Tetrahedron Lett.*, 1974, 257.
- 13 D. B. Davies, *Stud. Biophys.*, 1976, **55**, 29.
- 14 D. B. Davies, P. Rajani, H. Sadikot, M. MacCoss, and S. S. Danyluk, in preparation.

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