

Stereochemistry of Pyrrolizidine Alkaloid Biosynthesis: Incorporation of Chiral [2-²H]Putrescines into Retrorsine

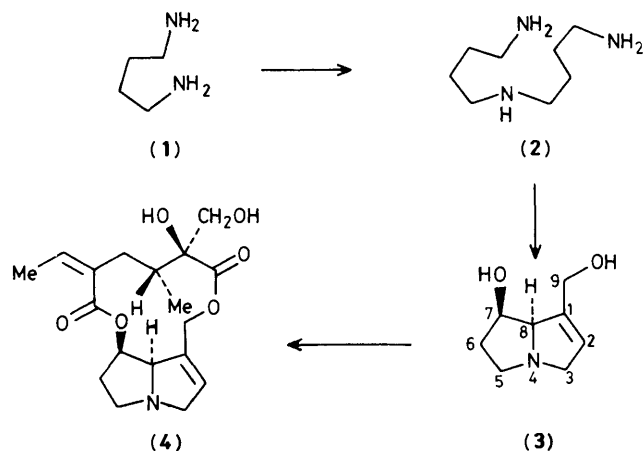
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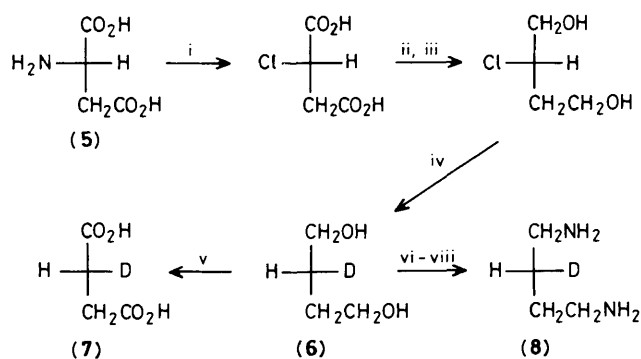
Feeding experiments with (2*R*)- and (2*S*)-[2-²H]putrescine on *Senecio isatideus* plants have shown that hydroxylation at C-7 of retronecine proceeds with retention of configuration, and formation of the 1,2-double bond involves removal of the *pro-S* hydrogen and retention of the *pro-R* hydrogen at C-2 of retronecine.

Retrorsine (**4**) is one of many pyrrolizidine alkaloids that contain retronecine (**3**) as the base portion.^{1,2} The C₈ skeleton of retronecine is derived biosynthetically from L-ornithine or L-arginine³ via putrescine (**1**).⁴⁻⁷ A later intermediate in the biosynthetic pathway is homospermidine (**2**), formed from two molecules of putrescine (Scheme 1).^{6,8} We have shown

that ²H n.m.r. spectroscopy can be used to establish complete labelling patterns in retrorsine derived biosynthetically from [1,4-²H₄]- and [2,3-²H₄]-putrescine in *S. isatideus* plants.⁹ With the latter precursor, ²H is retained at C-2, C-6 α , C-6 β , and C-7 α of retronecine (**3**). The stereochemical course of a number of enzymic processes involving removal of hydrogen



Scheme 1



Scheme 2. Reagents: i, HCl, HNO₃, urea; ii, SOCl₂, MeOH; iii, di-isobutylaluminium hydride; iv, LiAlD₄; v, chromic acid; vi, HBr; vii, NaN₃; viii, LiAlH₄.

from the 1- and 4-positions of putrescine has been established using chiral [1-²H]putrescine.^{10,11} We have now prepared samples of chiral [2-²H]putrescine and used them to obtain ²H-labelled retrorsine. The stereochemical course of further enzymic processes in retronecine biosynthesis has been established.

(2*R*)-[2-²H]Succinic acid (7) was prepared from (2*S*)-aspartic acid (5) as outlined by Arigoni and Eliel (Scheme 2).¹² An o.r.d. curve of this material (7) was in accord with literature values,^{12,13} and mass spectral data on the deuteriated succinic anhydride indicated ca. 98% ²H₁ species. The immediate precursor of the deuteriated succinic acid (7), (2*R*)-[2-²H]butane-1,4-diol (6) was converted into (2*R*)-[2-²H]putrescine (8) dihydrochloride (Scheme 2) [²H{¹H} n.m.r. (H₂O) δ 1.76 p.p.m. (s)]. Analogous treatment of (2*R*)-aspartic acid yielded samples of (2*S*)-[2-²H]succinic acid and (2*S*)-[2-²H]putrescine dihydrochloride. The latter sample contained an impurity of ca. 15% [1-²H]putrescine dihydrochloride [from ²H{¹H} n.m.r. (H₂O) δ 3.08 p.p.m. (s)]. This impurity may have arisen by lithium aluminium deuteride reduction of a small amount of incompletely reduced diester, left after the reduction with di-isobutylaluminium hydride (Scheme 2). Each precursor (200 mg) together with [1,4-¹⁴C]putrescine dihydrochloride (5 μCi) was pulse fed into the xylems of four *S. isatideus* plants over a period of two weeks. After a further two weeks, retrorsine (4) was isolated and recrystallised to constant specific radioactivity.

The ²H{¹H} n.m.r. spectrum⁹ of retrorsine derived from (2*R*)-[2-²H]putrescine (8) was obtained in chloroform at 60 °C

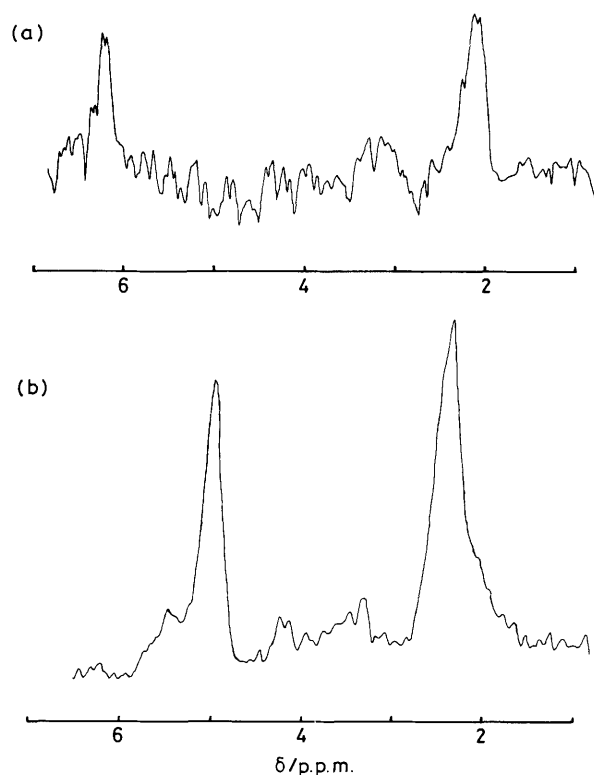


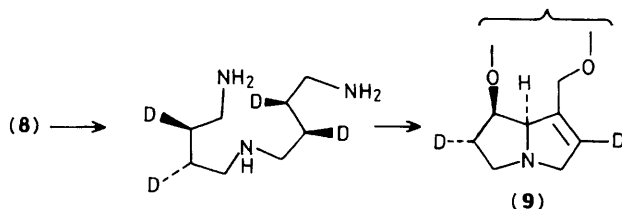
Figure 1. 30.72 MHz ²H{¹H} N.m.r. spectra of retrorsine in chloroform at 60 °C: (a) sample of (9) derived from (2*R*)-[2-²H]putrescine (8); (b) sample of (11) derived from (2*S*)-[2-²H]putrescine (10). Natural abundance ²H in CHCl₃ was the internal reference at δ 7.15.

[Figure 1(a)]. This showed two major signals at δ 2.15 and 6.20 p.p.m. corresponding to retrorsine (9) labelled with ²H at C-6α and C-2, respectively (Scheme 3). The enrichment factors† for the two labelled sites are nearly equal at ca. 0.18% ²H. The specific incorporation of ²H per C₄ unit is therefore 0.18% [0.18/2 × 2/98 × 100%, where 98/2 atom % ²H is the average enrichment of ²H at each labelled site of putrescine (8)]. This value is about half the ¹⁴C specific incorporation of 0.30% per C₄ unit. Removal of ²H from the carbon atoms destined to become C-1 and C-7 in retronecine accounts for this loss of about half of the ²H from (2*R*)-[2-²H]putrescine (8) on its conversion into retrorsine (9).‡

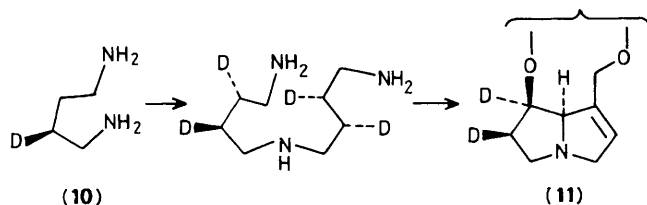
Using (2*S*)-[2-²H]putrescine (10) as a precursor, a sample of retrorsine was obtained which showed two major signals in the ²H{¹H} n.m.r. spectrum, obtained in chloroform at 60 °C, at δ 2.38 and 5.00 p.p.m. [Figure 1(b)]. This corresponds to retrorsine (11) labelled with ²H at C-6β and C-7α, respectively (Scheme 4). Enrichment factors† of 0.34% ²H for C-6β and 0.20% for C-7α were estimated, leading to a specific incorporation of ²H of 0.65% (0.54/2 × 2/83 × 100%) for the C₄ unit in which most of the ²H appears. This value is similar to the ¹⁴C specific incorporation of 0.73% per C₄ unit.‡ About half of the

† The enrichment factor for a labelled site in retrorsine is calculated from [(integral of labelled site in retrorsine) - (natural abundance integral/concentration of retrorsine)] / [(natural abundance integral of ²H in chloroform at δ 7.25 p.p.m./concentration of chloroform) × 0.0156%].

‡ Differences between the ²H and ¹⁴C specific incorporations, after taking into account the loss of ²H from some carbon atoms in retronecine, may be due to the operation of ²H isotope effects in the biosynthetic pathway.



Scheme 3§



Scheme 4§

^2H is lost from (2*S*)-[2- ^2H]putrescine (10) as it is transformed into retrorsine (11). Removal of ^2H from the carbon atoms which become C-1 and C-2 of retronecine leads to the appearance of most of the ^2H in one C₄ unit derived from putrescine.

The hydroxylation at C-7 of retronecine thus occurs with retention of configuration, in agreement with the stereospecificity generally observed for direct hydroxylation at sp³ carbon atoms.¹⁴ Formation of the 1,2-double bond in retronecine

§ No molecules of putrescine can contain more than one ^2H atom. The structures in Schemes 3 and 4 are therefore representations of all the ^2H -labelled species present.

takes place with retention of the *pro-R* hydrogen and loss of the *pro-S* hydrogen at the carbon atom which becomes C-2 of retronecine.

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