Chemical Communications

Number 22 1985

Synthesis of a Naturally Occurring Inhibitor of Glutamine Synthetase, Tabtoxinine-β-lactam

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 (\pm) -Tabtoxinine- β -lactam, a potent inhibitor of glutamine synthetase, has been synthesised by a route involving a nitroso Diels-Alder reaction.

Tabtoxin (1) is a dipeptide exotoxin produced by Pseudomonas tabaci, the organism responsible for Wildfire disease in tobacco plants. When hydrolysed by peptidases, in vivo, this exotoxin releases tabtoxinine-β-lactam (2), which inhibits the glutamine synthetase of the photorespiratory nitrogen cycle, causing chlorosis and death of the tobacco plant. It seems likely that this inhibition is the result of tight binding of tabtoxinine-β-lactam (2) to the enzyme as an analogue of the postulated tetrahedral intermediate (3) involved in the enzymatic reaction. 1a In 1983 we achieved a stereospecific synthesis of the dipeptide tabtoxin³ but found that this approach was not amenable to the synthesis of the actual toxin, tabtoxinine- β -lactam (2), nor was the hydrolysis of (1), under acidic^{1e} or enzymatic2b conditions, a satisfactory source of (2). Consequently, we have developed a new route to the toxin (2) which is based on an improved Diels-Alder strategy (Scheme 1, X = CN) and offers direct access to (\pm) -(2), with the correct regioand stereo-chemistry.

Thus, 2-chloroacrylonitrile was treated with butadiene (sealed tube, 90-100 °C, 24 h) to give the cyclohexene (4) (85%),4 which on heating at reflux in pyridine gave the diene (5)† (72%). Diene (5) was treated with benzyl nitrosoformate (generated in situ from benzyl N-hydroxycarbamate and tetraethylammonium periodate in CH₂Cl₂⁵) affording the adduct (6) as a single regioisomer [73%; δ_H(CDCl₃) 4.89 (1H, m, allylic), 6.56 (1H, dd, J 8.5 and 2Hz, vinylic), and 6.70 (1H, dd, J 8.5 and 6.3 Hz, vinylic)]. The nitrile function of (6) was smoothly reduced with NaBH₃(OCOCF₃)⁶ to give the amine (7) (58%), identical to that synthesised previously by a more complex procedure,³ confirming the desired regiochemistry of the Diels-Alder step. After protection of the amine (7) with a t-butoxycarbonyl group [BOC-ON (2-t-butoxycarbonyloxyimino-2-phenylacetonitrile), 7 CH₂Cl₂ to give (8) (83%; m.p. 115.5 °C), oxidative cleavage was accomplished by the method of Starks (KMnO₄, Bu₄N⁺HSO₄⁻, C₆H₆-H₂O, 25 °C)8 to give the diacid (9) [98%; $\delta_{H}(CD_{3}COCD_{3})$ 3.4—3.8 (2H, ABX, CH₂NH) and 4.63 (1H, m, allylic)]. The difficult step of differentiation of the two carboxy groups of (9) was successfully achieved by decarboxylative esterification [benzyl chloroformate (1.5 equiv.), pyridine, CH_2Cl_2 , 25 °C] to give the monoester (10) [57%; $\delta_H(CDCl_3)$ 5.08 (2H, br, benzylic), 5.15 (2H, br, benzylic), and 7.0—7.5 (10H, m, 2 × Ph)] along with a small amount of the diester (11) [4.8%; $\delta_H(CDCl_3)$, 5.05—5.38 (7H, m, 6 benzylic H and NH), and 7.25—7.45

Scheme 1. $X = CO_2Et$, CN.

 $[\]dagger$ 1H N.m.r. (300 MHz), i.r., and mass spectra were entirely consistent with the assigned structures for all new compounds and satisfactory combustion analyses were obtained.

(4) (5) (6)
$$R = CN$$
(7) $R = CH_2NH_2$

$$R^{1} O_{2}C$$

$$R^{2}O_{2}C$$

$$H$$

$$CO_{2}CH_{2}Ph$$

$$PhCH_{2}O_{2}C$$

$$H$$

$$(3)$$

$$R^{1} = R^{2} = H, \quad X = CO_{2}Bu^{t}$$

$$(13)$$

(8) $R = CH_2NHCO_2Bu^t$

- (10) $R^1 = H$, $R^2 = CH_2Ph$, $X = CO_2Bu^t$
- (11) $R^1 = R^2 = CH_2Ph$, $X = CO_2Bu^t$
- (12) $R^1 = H$, $R^2 = CH_2Ph$, X = H

 $(15H, m, 3 \times Ph)$]. Removal of the primary amino protection of (10) with 98% formic acid (25 °C, 3 h)9 gave the penultimate precursor, the β-amino acid (12) (99%). The β-lactam closure was achieved by Ohno's procedure [Ph₃P-di-2-pyridyl disulphide-MeCN, 80 °C, 2 hlio to yield the spiro β-lactam (13) [63%; $ν_{max}$.(CHCl₃) 1780, 1740, and 1710 cm⁻¹; $δ_H$ (CDCl₃) 3.33 (1H, d, J 5 Hz, β-lactam) and 3.56 (1H, d, J 5 Hz, β-lactam)]. Hydrogenation of (13) (10% Pd-C, EtOH) resulted in complete deprotection and concomitant reductive cleavage of the N-O bond to provide (±)-tabtoxinine-βlactam (2) [quantitative, v_{max} (D₂O) 1736 cm⁻¹; δ_H (D₂O) 1.62-2.00 (4H, m, CH₂CH₂), 3.15 (1H, d, J 6 Hz, β -lactam), 3.28 (1H, d, J 6 Hz, β -lactam), and 3.60 (1H, m, CHNH₂)] which was identical (500 MHz n.m.r.) to the sample isolated from P. tabaci and was biologically active in vitro and in vivo.

We are grateful to Dr. J. G. Turner, University of East Anglia, for a sample of natural tabtoxinine-β-lactam and for biological tests, to Professor R. D. Durbin, University of Wisconsin, for the ¹H n.m.r. spectrum of natural tabtoxinineβ-lactam and Professors M. Ohno and S. Kobayashi, University of Tokyo, for helpful advice on the β-lactam formation procedure.

Received, 25th June 1985; Com. 898

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