Reaction of the Aquocopper(1) Ion with Hydrogen Peroxide: Evidence against Hydroxyl Free Radical Formation

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Kinetic evidence supports the formation of a copper(\mathfrak{m}) species, rather than the hydroxyl free radical, when the aquocopper(\mathfrak{l}) ion reacts with hydrogen peroxide in aqueous solution.

The reaction of the aquocopper(I) ion with H_2O_2 is involved in a variety of processes induced by O_2^- in vitro, including depolymerization of synovial fluid,¹ cytotoxicity,² and oxidative degeneration of DNA³ as well as in copper-catalysed epoxidation and hydroxylation processes.⁴ We present here evidence that has led us to question the validity of the assumption commonly made,³⁻⁶ that the OH free radical is formed by reaction (1). Our experimental findings are interpreted in terms of the formation of a Cu^{III} species by reaction of H_2O_2 with the Cu^I ion and some of the reactions of this Cu^{III} intermediate are discussed.

$$Cu^{I} + H_{2}O_{2} \rightarrow Cu^{II} + OH^{-} + OH^{-}$$
(1)

The method used to test if the OH radical is a product of reaction (1) involved investigation of the γ -ray induced oxidation of methanol to formaldehyde in aqueous solutions containing hydrogen peroxide and aquocopper(II) ions. Under the conditions used, where the oxidation occurs as a chain reaction involving reaction of Cu^I ions with H₂O₂ as a propagation step, it is possible to test if the reaction kinetics are consistent with the assumption that this propagation step produces the OH radical to act as chain carrier.

Aqueous solutions containing methanol $(0.1-2 \text{ mol dm}^{-3})$, copper(11) sulphate $(1-2 \times 10^{-3} \text{ mol dm}^{-3})$, and H₂O₂ $(0-5 \times 10^{-3} \text{ mol dm}^{-3})$, saturated with nitrous oxide gas (*ca.* $2 \times 10^{-2} \text{ mol dm}^{-3}$), were irradiated with γ -rays (Co-60, dose rate = 0.2-0.8 Gy s⁻¹). The HCHO yield was measured⁷ immediately after irradiation. The extent of a relatively slow thermal reaction yielding HCHO in the absence of radiation was minimised by introducing the H₂O₂ immediately before, and acidifying immediately after, exposure. Radiation yields given, which are initial values taken from linear yield-dose plots, have been corrected for the HCHO yield in the unirradiated control (<10% of the radiation yield).

In γ -irradiated, dilute aqueous solutions, reactions are initiated by the reactive species produced by the radiation-induced water decomposition process, equation (2).⁸ In our system, in the absence of H₂O₂, the reactions (3)—(7) occur subsequently.⁸

$$H_2O \longrightarrow OH^{\bullet}, H^{\bullet}, e_{ag}^{-}, H_2O_2, H_2, H^+, OH^-$$
 (2)

$$e_{aq}^{-} + N_2 O (+H^+) \rightarrow N_2 + OH \cdot$$
(3)

$$OH \cdot + CH_3OH \rightarrow CH_2OH + H_2O$$
 (4)

$$H \cdot + CH_3OH \rightarrow \cdot CH_2OH + H_2$$
 (5)

$$e_{aq}^{-} + Cu^{II} \rightarrow Cu^{I}$$
 (6)

$$\cdot CH_2OH + Cu^{II} \rightarrow HCHO + Cu^{I} + H^+$$
(7)

In the absence of added H_2O_2 , the observed yield of HCHO, $G(HCHO) = 0.64 \mu mol J^{-1}$ is consistent with the yield expected from the known radiation yields of the primary free radicals.⁸

With H_2O_2 present, the yield of HCHO was significantly increased to values that showed that a chain reaction occurred. The chain length is equal to $G(\text{HCHO})/G_i$, where $G_i = G(\text{OH} \cdot) + G(e_{aq}) + G(\text{H} \cdot) \approx 0.68 \,\mu\text{mol J}^{-1}$ is the yield of initiating radicals. The yield was markedly dependent on the pH $\{e.g., \text{ for } [CH_3OH] = 1.0, [H_2O_2] = 0.003, \text{ and } [Cu^{II}] =$ 0.001 mol dm⁻³, G(HCHO) decreased from 120 to 10 µmol J^{-1} over the range pH = 0.8-5.4}. At constant pH, G(HCHO) was directly proportional to [dose rate]^{-1/2} and to $[CH_3OH]^{1/2}$. The dependence of G(HCHO) on $[H_2O_2]$ was complex, increasing with increasing $[H_2O_2]$ at pH = 5.3 but decreasing with increasing $[H_2O_2]$ at pH <4. Over the range of conditions studied, G(HCHO) was independent of the Cu^{II} ion concentration. The maximum measured chain length was ca. 225. There was no evidence that significant concentrations of CuI products accumulated during the reaction. No chain production of HCHO was apparent when small concentrations of O_2 (ca. 10^{-4} mol dm⁻³) were present.

If reaction (1) occurs in this system, production of HCHO by a chain process, propagated by reactions (1), (4), and (7), would be expected. For this mechanism, the dependence of G(HCHO) on methanol concentration would imply chain termination by the OH \cdot free radical combination reaction. Taking reported values⁹ for the rate constants of this termination and of the propagation reaction (4), with dose rate = 0.8 Gy s⁻¹, [CH₃OH] = 1.0 mol dm⁻³, the expected chain length would be greater than the maximum observed by a factor of *ca.* 10⁵. Furthermore, this mechanism does not provide any explanation for the observed dependence of G(HCHO) on pH and on the H₂O₂ concentration.

As an alternative mechanism, we suggest the Cu^{I} ion reacts with hydrogen peroxide to give a species, formally Cu^{III} , that can react either with methanol to propagate, or with the Cu^{I} ion to terminate, the chain reaction.

$$Cu^{I} + H_2O_2 \rightarrow Cu^{III}$$
 (8)

$$Cu^{III} + CH_3OH \rightarrow CH_2OH + Cu^{II} + H^+$$
 (9)

$$Cu^{III} + Cu^{I} \rightarrow 2Cu^{II} \tag{10}$$

Assuming a mechanism for this chain reaction with initiation by reactions (2) to (6), propagation by reactions (7), (8), and (9) and termination by reaction (10), and making the steady-state approximation, equation (11) is obtained for G(HCHO) where D = dose rate.

$$G(\text{HCHO}) = 0.75G_{i} + \{k_{8}k_{9}[\text{CH}_{3}\text{OH}][\text{H}_{2}\text{O}_{2}]/2k_{10}D\}^{1/2}$$
(11)

This is consistent with the observed linear dependence of G(HCHO) on $[\text{CH}_3\text{OH}]^{1/2}$ and (dose rate)^{-1/2}. The mechanism is also compatible with our results in that it implies a low stationary concentration of Cu^I ions, a chain length independent of the Cu^{II} ion concentration, and effective termination by oxygen, which reacts rapidly with Cu^I species. The observed complex dependence of G(HCHO) on $[\text{H}_2\text{O}_2]$ is not explained by this mechanism alone but possibly indicates that, under certain conditions, the Cu^{III} intermediate can react with H_2O_2 to terminate the chain.

The formation of a Cu^{III} species by a reaction of hydrogen peroxide with Cu^I has previously been suggested.¹⁰ Furthermore, there is evidence that an aquo Cu^{III} species is produced by reaction of OH • radicals with Cu^{II} ions¹¹ and exists in forms linked by the protolytic equilibria in equation (12).^{8,12} There is considerable uncertainty associated with the values of the acid dissociation constants for these species¹² but it is probable that $pK_a < 3.5$ for Cu(OH)²⁺ and $pK_a = 4-6$ for Cu(OH)₂⁺. The dependence of chain length on pH in our system can be accounted for if it is assumed that the value of the rate constant ratio, k_9/k_{10} , depends on the extent of protonation of the Cu^{III} species.

$$\operatorname{Cu}_{\operatorname{aq}}^{3+} \rightleftharpoons \operatorname{Cu}(\operatorname{OH})^{2+} \rightleftharpoons \operatorname{Cu}(\operatorname{OH})_2^+ \rightleftharpoons \operatorname{Cu}(\operatorname{OH})_3$$
 (12)

In the presence of certain ligands capable of complexing copper ions, including amino acids, oxalate, and 1,10phenanthroline, the chain length in the irradiated Cu^{II}– hydrogen peroxide–methanol system was significantly decreased. This may be because of a decrease in the rate constant ratio, k_9/k_{10} , when the Cu ions are complexed, with a consequent decrease in the efficiency with which methanol is oxidised. Alternatively, the complexed Cu^{III} species may react by oxidation of the attached ligand, rather than by dehydrogenation of methanol. The latter appears to be the case, at least with glycine and α -alanine as ligands, where we have observed the formation of ammonia as a product in yields that indicate that ligand oxidation is a significant process.

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