

¹⁷O N.M.R. Studies of Enol and Phenol Compounds with Intramolecular Hydrogen Bonds

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The natural abundance 40.67 MHz ¹⁷O n.m.r. spectra of enol and phenol (naphthol) compounds with intramolecular hydrogen bonds are reported and the chemical shifts of the chelated hydroxy groups are assigned.

The fast 1,5-sigmatropic $A \rightleftharpoons B$ tautomerism shown by six-membered enol chelates with intramolecular hydrogen bonds is one of the most common types of tautomeric interconversion.^{1,2} These equilibria should be amenable to study by ¹⁷O n.m.r. spectroscopy, which provides a reasonably wide range of chemical shifts.³ However, the available ¹⁷O n.m.r. data for enol compounds are controversial and do not allow the range of shifts of enol oxygen to be established. In the 1960's, the spectra of a number of highly enolised β -dicarbonyl compounds were reported,⁴ but only the averaged position of the oxygen signals was obtained, since the fast $A \rightleftharpoons B$ equilibrium made it impossible to obtain hydroxylic oxygen shifts. This information may be obtained from the spectra of β -ketoesters, which show no enolisation of ester groups and, consequently, no fast exchange. Lambert and Wharry have recently reported the ¹⁷O n.m.r. spectra of ethyl acetoacetate (1) analogues.⁵ They did not, however, take into account the presence of two tautomers in the mixture ($K \rightleftharpoons E$), which makes their assignment of signals doubtful.

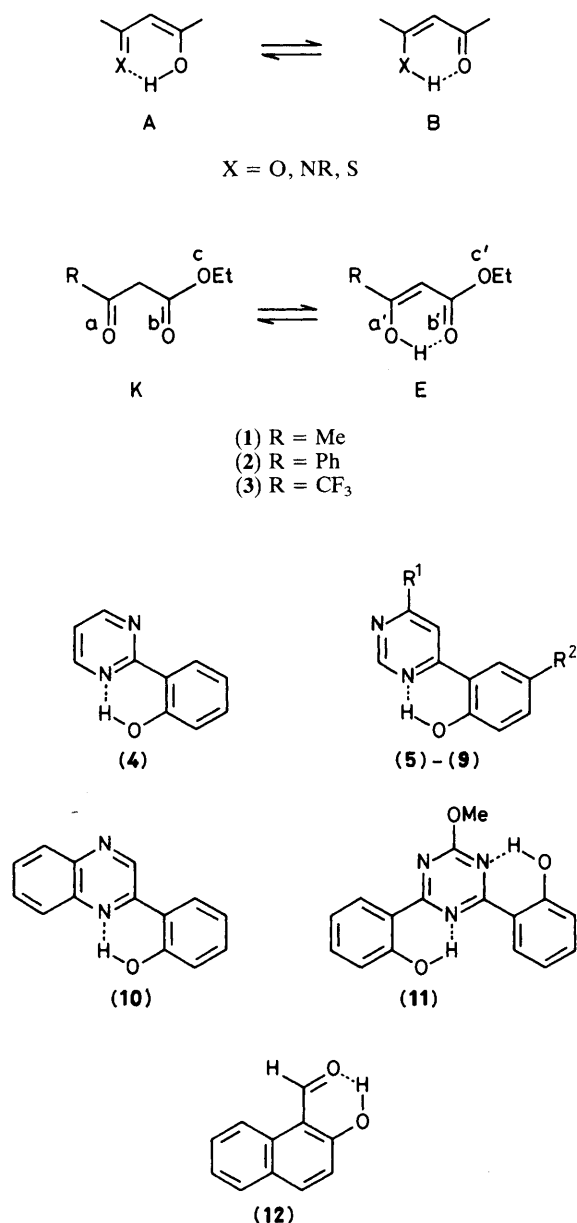
We report here the ¹⁷O n.m.r. signals for the chelated hydroxy groups of the enol and phenol (naphthol) compounds (1)—(12).

In the ¹⁷O n.m.r. spectra of ethyl acetoacetate (1) (Figure

1), the signals of the keto and enol forms may be easily differentiated by their intensities; as shown by ¹H n.m.r. spectroscopy, the content of the keto and enol tautomers is 92 and 8%, respectively. Further assignments have been made in accordance with ref. 6 and are in good agreement (see Figure 1) with the data for ethyl benzoylacetate (2) (17% of enol) and the trifluoroacetoacetate (3) (>90% of enol).

As seen in the case of the enols (1)—(3), the signal position of hydroxylic oxygen is sensitive to the neighbouring group R. By contrast, in the (2-hydroxyphenyl)-di- and tri-azines the δ_{OH} value is almost independent of the nature of the chelate's basic moiety. For these chelates the δ_{OH} value remains practically constant (90—97 p.p.m.), in spite of considerable variations in the basicity of the azine fragment. Very similar δ_{OH} values (85—90 p.p.m.) are also shown by phenol chelates with a carbonyl group as the basic fragment.⁷ Therefore, in spite of considerable variations in the structure of the six-membered chelate [(2-hydroxyphenyl)-di- and tri-azines; *o*-hydroxyacetophenones⁷], the δ_{OH} range of phenolic oxygen is comparatively narrow (85—97 p.p.m.). In related chelates of the enol type the δ_{OH} range is wider (95—125 p.p.m.), however.

Reuben and Samuel⁸ have assigned the signal at 315 p.p.m.



¹⁷O N.m.r. chemical shifts in p.p.m.

Compound	δ_{OH}
(4)	96
(5) R ¹ =H, R ² =H	94
(6) R ¹ =H, R ² =Br	95
(7) R ¹ =H, R ² =OMe	90
(8) R ¹ =Cl, R ² =H	95
(9) R ¹ =NEt ₂ , R ² =H	97
(10)	95
(11)	97
(12)	95

to the hydroxy group of 1-phenylazo-2-naphthol (see also ref. 9). However such a strong shift of the signal could not be induced by hydrogen bonding or annelation, as illustrated by our ¹⁷O n.m.r. studies of the naphthol (12); the shift of the hydroxy group is almost unaffected by annelation. The signal at 315 p.p.m.⁸ seems therefore to be that of the hydroxylic and keto oxygens of the hydroxy and oxo tautomers of 1-phenylazo-2-naphthol averaged by fast exchange. Compari-

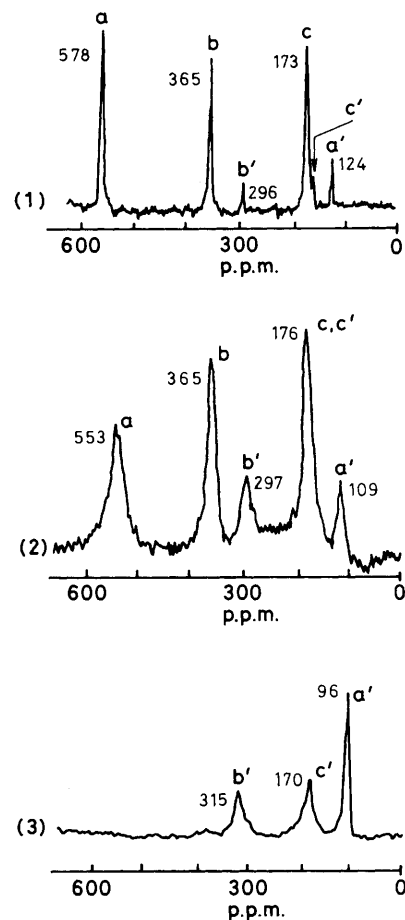


Figure 1. ¹⁷O N.m.r. spectra of ketoesters (1)–(3) in CHCl₃ at ca. 300 K (Bruker CXP-300 spectrometer 10⁴–10⁶ scans, chemical shifts in p.p.m. downfield from external H₂O).

son of our data (95–125 p.p.m. range) with the enol chemical shifts (–40 to +40 p.p.m.) adopted by Gorodetsky⁴ suggests that his results (see also ref. 10) on the enol–enol tautomerism of β -diketones need to be revised. The disagreement between our signal assignment for ethyl acetoacetate and that by Lambert and Wharry⁵ might be due to the lower sensitivity of their instrument.

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References

- Y. Karlstrom, B. Jonsson, B. Roos, and H. Wennerstrom, *J. Am. Chem. Soc.*, 1976, **98**, 6851.
- R. S. Brown, *J. Am. Chem. Soc.*, 1977, **99**, 5497.
- W. G. Klemperer, *Angew. Chem., Int. Ed. Engl.*, 1979, **17**, 246.
- M. Gorodetsky, L. Luz, and Y. Mazur, *J. Am. Chem. Soc.*, 1967, **89**, 1183.
- J. B. Lambert and S. M. Wharry, *J. Am. Chem. Soc.*, 1982, **104**, 5857.
- J. W. Emsley, J. Feeney, and L. H. Sutcliffe, 'High Resolution N.M.R. Spectroscopy,' Pergamon Press, Oxford, 1966.
- T. Amour, M. Burgar, B. Valentine, and D. Fiat, *J. Am. Chem. Soc.*, 1981, **103**, 1128.
- J. Reuben and D. Samuel, *Isr. J. Chem.*, 1963, **1**, 279.
- J.-P. Kintzinger, in 'NMR Basic Principles and Progress,' eds. P. Diehl, E. Fluck, and R. Kosfeld, Springer Verlag, Berlin, 1982, vol. 17, p. 1.
- A. I. Koltsov and G. M. Kheifets, *Usp. Khim.*, 1971, **40**, 1646.