

A Formal Synthesis of Aflatoxin B₂

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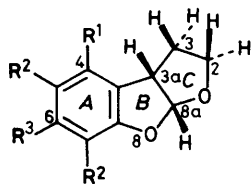
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A brief synthesis of a ring A differentiated tetrahydrofurobenzofuran intermediate previously converted into Aflatoxin B₂ is described.

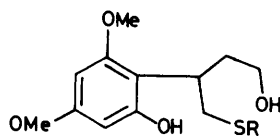
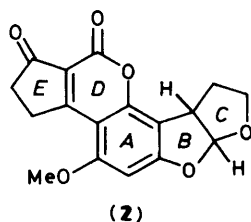
The aflatoxins are a well known group of acutely toxic and highly carcinogenic metabolites of various *Aspergillus* species. Long-standing problems associated with the synthesis of these mycotoxins have been identified and discussed.¹ A recent preparation² of the furo[2.3-*b*]benzofuran (**1**), constituting rings A, B, and C of Aflatoxin B₂ (**2**) with correctly differentiated oxygen substituents on the phloroglucinol (ring A) moiety, represents a formal synthesis of (**2**) and 'state of

the art' in aflatoxin synthesis. We now report a simple synthesis of (**1**) in *ca.* 4% overall yield from commercially available 3,5-dimethoxy phenol (**4**).

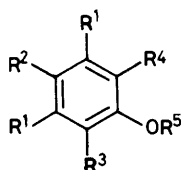
During our initial investigations, the dihydrobenzofuran (**6**) was prepared in two steps from *ortho*-iodophenol *via* the crotonate (**5**) by the intramolecular conjugate addition procedure we had recently developed.³ Reduction to the alcohol (**7**) and treatment of the latter with lead tetra-acetate (LTA),



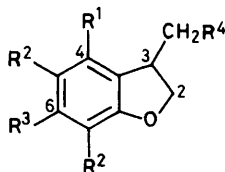
- (1) $R^1 = \text{OH}, R^2 = \text{H}, R^3 = \text{OMe}$
 (3) $R^1 = \text{OBn}, R^2 = \text{H}, R^3 = \text{OMe}$
 (8) $R^1 = R^2 = R^3 = \text{H}$
 (14) $R^1 = R^3 = \text{OMe}, R^2 = \text{I}$
 (21) $R^1 = \text{OBn}, R^2 = \text{I}, R^3 = \text{OMe}$
 (22) $R^1 = \text{OH}, R^2 = \text{I}, R^3 = \text{OMe}$
 Bn = PhCH_2 , benzyl



- (16) $R = \text{Et}$
 (17) $R = \text{CMe}_3$



- (4) $R^1 = \text{OMe}, R^2 = R^3 = R^4 = R^5 = \text{H}$
 (5) $R^1 = R^2 = R^3 = \text{H}, R^4 = \text{I}, R^5 = \text{CH}_2\text{CH}=\text{CHCO}_2\text{Et}$
 (9) $R^1 = \text{OMe}, R^2 = R^5 = \text{H}, R^3 = R^4 = \text{I}$
 (10) $R^1 = \text{OMe}, R^2 = \text{H}, R^3 = R^4 = \text{I}, R^5 = \text{CH}_2\text{CH}=\text{CHCO}_2\text{Et}$



- (6) $R^1 = R^2 = R^3 = \text{H}, R^4 = \text{CO}_2\text{Et}$
 (7) $R^1 = R^2 = R^3 = \text{H}, R^4 = \text{CH}_2\text{OH}$
 (11) $R^1 = R^3 = \text{OMe}, R^2 = \text{H}, R^4 = \text{CO}_2\text{Et}$
 (12) $R^1 = R^3 = \text{OMe}, R^2 = \text{H}, R^4 = \text{CH}_2\text{OH}$
 (13) $R^1 = R^3 = \text{OMe}, R^2 = \text{I}, R^4 = \text{CH}_2\text{OH}$
 (15) $R^1 = \text{OH}, R^2 = \text{H}, R^3 = \text{OMe}, R^4 = \text{CH}_2\text{OH}$
 (18) $R^1 = \text{OMe}, R^2 = \text{H}, R^3 = \text{OH}, R^4 = \text{CH}_2\text{OH}$
 (19) $R^1 = \text{OAc}, R^2 = \text{H}, R^3 = \text{OMe}, R^4 = \text{CH}_2\text{OAc}$
 (20) $R^1 = \text{OBn}, R^2 = \text{I}, R^3 = \text{OMe}, R^4 = \text{CH}_2\text{OH}$

iodine, and solid calcium carbonate in refluxing benzene⁴ provided rapid access to the parent furobenzofuran (8) in four steps and 25% overall yield from *ortho*-iodophenol. In order to test the LTA-iodine cyclisation with a substrate possessing the activated phloroglucinol ring A of aflatoxin, the preparation of (12) was undertaken by the same method. Iodination⁵ of 3,5-dimethoxy phenol (4) with KI-KIO₃ produced the symmetrical (¹H n.m.r.) di-iodo derivative (9), which was converted into the crotonate (10) and cyclised with concomitant deiodination with two molar equivalents of *n*-butyllithium at -100 °C. Reduction of (11) with di-isobutyl(aluminium) hydride (DIBAL-H) produced the desired alcohol (12) in 23.2% overall yield. When the LTA-iodine reaction was attempted with (12), aromatic iodination rather than cyclisation resulted and when two molar equivalents of the reagent

were used, the di-iodo derivative (13) was isolated from among a mixture of products. However, when (13) was purified and resubjected to the cyclisation, the furobenzofuran (14) was formed and its presence indicated by the appearance of H-8a as a doublet at δ 6.35 (J 5.7 Hz) in the ¹H n.m.r. spectrum of the product.

The differentiation of the oxygen substituents on ring A was always a major difficulty that had been addressed in the initial stages of the early syntheses¹ of the aflatoxins. Our preference for a method of intramolecular discrimination between the methoxy groups in an intermediate like (11) or (12) was enhanced by a recent study⁶ of hydroxyl directed demethylation of various methoxylated aromatic substrates by the ethyl mercaptide anion. As it happened, the application of this method to (12) did result in retention of the C-6 methoxy group and demethylation at C-4 [(15) 8%], but a concurrent reaction at C-2 gave a second product identified as (16) (16%). Much better results ensued with *t*-butyl mercaptide [in dimethylformamide (DMF), 110 °C, 3 h]; the selectivity and yield were both vastly improved [(15) 61 and (17) 13%] and at no time was any of the 6-demethylated product† found. The two products were easily separated by column chromatography on silica gel. Iodination of (15) with KI-KIO₃, followed by benzylation (benzyl bromide-potassium carbonate), provided the 5,7-di-iodo-4-*O*-benzyl derivative (20), which was cyclized with LTA-I₂ to (21). Hydrogenolysis of this compound under a variety of conditions gave no worthwhile result; neither (1),(3) nor (22) could be obtained cleanly. However, deiodination of (21) was effected (Bu^nLi , -100 °C; H₂O) and the product (3) characterized‡ by 500 MHz ¹H n.m.r. As before,² transfer hydrogenolysis (cyclohexa-1,4-diene, Pd-charcoal) provided (1) with melting point and ¹H n.m.r. data identical with literature values, thus completing a formal synthesis of Aflatoxin B₂. We are making progress towards the synthesis of (22) which we expect to employ as an ABC intermediate for the attachment of rings D and E.

We thank the Natural Sciences and Engineering Research Council of Canada for support of this work.

Received, 21st January 1988; Com. 8/00178B

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† The identity of (15) was provisionally inferred by comparison of the ¹H n.m.r. spectra of (15) and its diacetate (19). The *meta* coupled (J 2.1 Hz) aromatic protons of (15) at δ 5.99 and 6.02 were shifted to δ 6.16 and 6.28 (J 2.1 Hz) in (19). If (18) had been formed by demethylation at C-6, both aromatic protons H-5 and H-7 should have been equally affected by acetylation.⁷

‡ Spectral data for (3): ¹H n.m.r. (500 MHz, CDCl₃), δ 2.10–2.15 (1H, m, H-3 β), 2.205 and 2.23 (1H, dd, H-3 α), 3.63–3.68 (1H, dq, H-2 α), 3.74 (3H, s, OMe), 4.01 and 3.99 (1H, dd, H-3 α), 4.05 (1H, t, H, 2 β), 4.30 (2H, s, OCH₂Ph), 6.07, 6.09 (1H each, d each, H-5 and -7), 6.30 (1H, d, H-8a), 7.40 (5H, m, OCH₂Ph). Coupling constants $J(\text{Hz})$: 2 α ,2 β 8.6; 2 α ,3 α 5.0; 2 α ,3 β 12.0; 2 β ,3 α ~0; 2 β ,3 β 8.1; 3 α ,3 β 12.2; 3 α ,3 α ~0; 3 β ,3 α 8.1; 3 α ,8a 5.7; and H-5,H-7 2.0.