

Synthesis of 12-Oxophytodienoic Acid (12-oxoPDA), Metabolic Parent of OPC-Compounds and *epi*-Jasmonic Acid

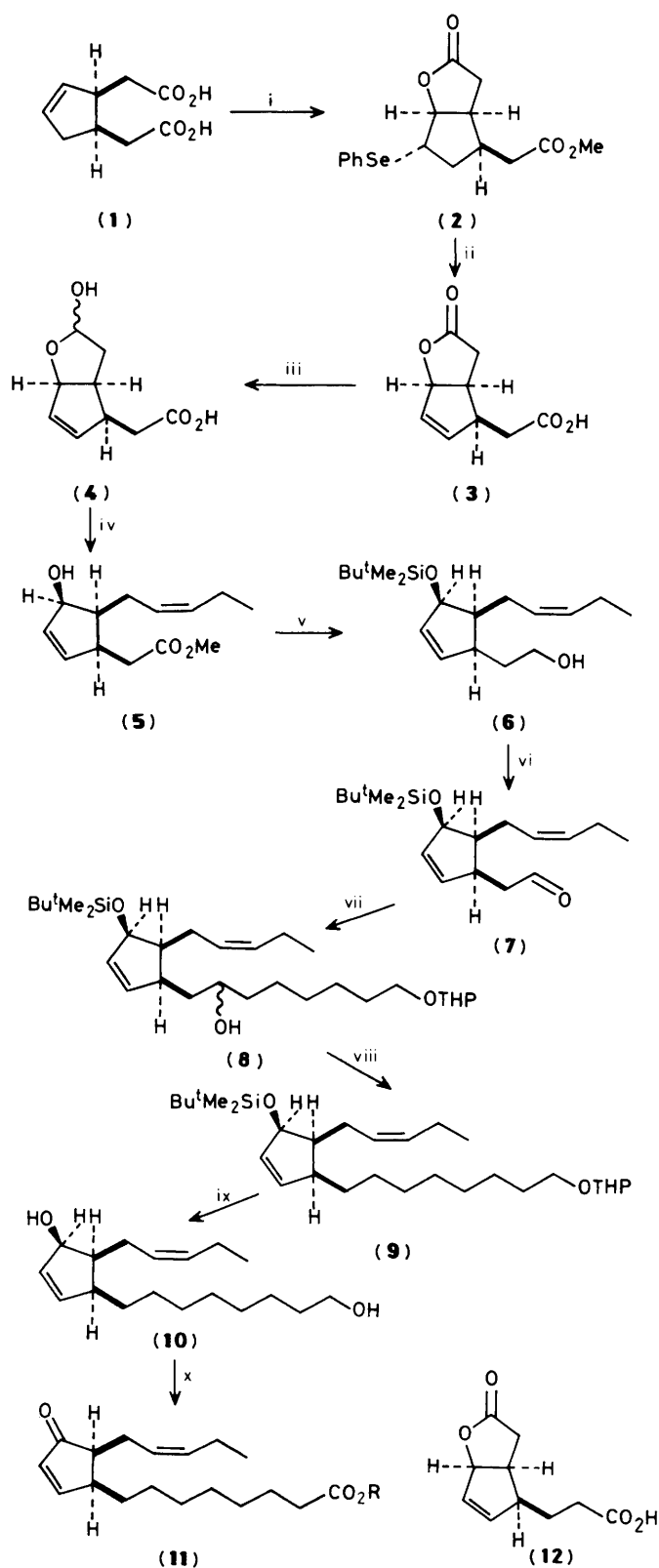
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The first synthesis of (\pm)-12-oxophytodienoic acid, a compound widely distributed in plants, and the metabolic parent of OPC compounds including *epi*-jasmonic acid, is described; whilst structurally resembling prostaglandin PGA_1 , it belongs to the *cis*-series not found in mammals.

12-Oxophytodienoic acid (12-oxoPDA) (**11**) is formed by an enzyme with cyclase activity which occurs in flax and other seeds and seedlings and whose substrate is (13*S*)-hydroperoxyoctadeca-(9*Z*),(11*E*),(15*Z*)-trienoic acid.¹ The latter, in its turn, is formed by the action of soya bean lipoxygenase on

linolenic acid. 12-OxoPDA is of considerable interest in connection with plant regulatory substances and its reduction and β -oxidation² explains the origins of the growth regulating³ and senescence inducing⁴ jasmonic acid. It is also likely to be associated with metabolic pathways leading to jasmone and



Scheme 1. Reagents and conditions: i, (a) PhSeCl, THF, $-78^{\circ}\text{C}\rightarrow 20^{\circ}\text{C}$, (b) CH_2N_2 ; ii, (a) H_2O_2 , THF, (b) KOH, MeOH; iii, DIBAL (3.3 mol), -45°C ; iv, (a) $\text{Ph}_3\text{P}=\text{CHEt}$, DMSO, 70°C , (b) CH_2N_2 ; v, (a) $\text{Bu}^t\text{Me}_2\text{SiCl}$, DMF, imidazole, (b) DIBAL, THF, -78°C ; vi, PDC, CH_2Cl_2 ; vii, $\text{BrMg}[\text{CH}_2]_6\text{OTHP}$, THF; viii, (a) MeSO_2Cl , pyridine, (b) LiAlH_4 , Et_2O , reflux; ix, (a), $\text{Bu}_4\text{N}^+\text{F}^-$, THF, 45°C , (b) PPTS, EtOH , 50°C ; x, for R = H: PDC, DMF, 20°C ; for R = Me: CH_2N_2 .

other jasmonoids, the pyrethrins ketols,⁵ and natural products from *Chromolaena* spp.⁶ 12-OxoPDA is widely distributed in higher plants⁷ and its structure (11) clearly resembles the mammalian prostaglandins (PGA_1), yet there are major differences. Thus, all of the latter have a *trans*-attachment of side chains, whilst 12-oxoPDA and its degradation cascade have *cis*-attachments. An even more marked contrast is the apparently differing modes of biosynthesis.^{8,9} We now report the first synthesis of (\pm)-12-oxoPDA.

In selecting a suitable starting point attention was turned to the *cis*-cyclopentenediacetic acid (1),¹⁰ which had earlier been employed in a synthesis of *trans*(*Z*)-jasnone.¹¹ Treatment with benzeneselenenyl chloride gave a selenolactonic acid (75% yield), m.p. $130\text{--}131^{\circ}\text{C}$, which was esterified to form the methyl ester (2), m.p. $68\text{--}69^{\circ}\text{C}$, in almost quantitative yield. Oxidative elimination followed by hydrolysis gave the unsaturated lactone acid (3), m.p. $114\text{--}115^{\circ}\text{C}$, in 92% yield overall. The lactone (3) was reduced to the lactol (4) by di-isobutylaluminium hydride (DIBAL) in tetrahydrofuran (THF), but the lactol was not purified but treated directly with excess of propyldimethylphenylphosphorane in dimethyl sulphoxide (DMSO) and then esterified with diazomethane to give (5). The Wittig reaction was not fully stereospecific, giving an 85:15 mixture of (*Z*):(*E*) alkenes (51% yield over three steps). After conversion into the *t*-butyldimethylsilyl derivative (99%), the (*Z*)/(*E*) mixture was separated by chromatography over 15% silver nitrate impregnated silica [eluant: ethyl acetate-benzene (1:10)]. The synthesis was then continued with the pure *t*-butyldimethylsilyl-(*Z*)-ester which was reduced to alcohol (6) (94%) by di-isobutylaluminium hydride.

A number of attempts were made to extend the length of the saturated side chain† and the method eventually selected was oxidation of alcohol (6) to aldehyde (7) (78%) using pyridinium dichromate (PDC) and treatment of (7) with a refluxing solution of $\text{BrMg}[\text{CH}_2]_6\text{OTHP}$ (THP = tetrahydropyran-2-yl) in THF for 1 h. This gave, in 92% yield, a pair of (\pm)-diastereoisomers (8) which were chromatographically separable. Each diastereoisomer was mesylated in almost theoretical yield, and then reductively demesyated by refluxing with lithium aluminium hydride in ether. Both gave the same product (9), one in 75%, the other in almost 100% yield. The cyclopentene (9) was stripped of its protecting groups, first removing the silyl protection with tetrabutylammonium fluoride ($\sim 100\%$), and then the tetrahydropyran-2-yl group (56% yield) by pyridinium toluene-*p*-sulphonate (PPTS). Oxidation of both the secondary and primary alcohol functions in (10), using pyridinium dichromate in DMF, followed by esterification, gave (\pm)-12-oxoPDA as its methyl ester (20% yield). The compound was identical (^1H n.m.r. and t.l.c. comparisons), with a specimen of authentic 12-oxoPDA made in our laboratory⁹ enzymically using flax enzyme.¹ It was epimerised at C-13 by acid to give the *trans*-isomer which was spectroscopically and chromatographically (g.l.c.) distinct from the natural *cis*-isomer.

During the synthesis checks on *cis*-stereochemistry were made using nuclear Overhauser effects on the set of *cis*-disposed ring methine protons: the validity of this correlation was checked by an X-ray structure determination of (12).

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† For example, Kolbe anodic synthesis at an earlier stage employing (3), and successfully used in the cyclopentanone OPC series, gave low yields. Similarly, employment of an Arndt-Eistert homologated precursor (12) gave unsatisfactory yields.

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References

- 1 D. C. Zimmerman and P. Feng, *Lipids*, 1978, **13**, 313.
 - 2 B. A. Vick and D. C. Zimmerman, *Plant Physiol.*, 1986, **80**, 202.
 - 3 *Inter alia*: J. Ueda and J. Kato, *Physiol. Plant.*, 1982, **54**, 248; *Plant. Physiol.*, 1980, **66**, 246; H. Yamane, H. Takagi, H. Abe, T. Yakota, and N. Takahashi, *Plant Cell Physiol.*, 1981, **22**, 689; H. Fukui, K. Koshimizu, Y. Yamazaki, and S. Usuda, *Agric. Biol. Chem.*, 1977, **41**, 189.
 - 4 W. Dathe, H. Ronsch, A. Preiss, W. Schade, G. Sembdner, and K. Schreiber, *Planta*, 1981, **153**, 530; H. Yamane, J. Sugawara, Y. Suzuki, E. Shimamura, and N. Takahashi, *Agric. Biol. Chem.*, 1980, **44**, 2857.
 - 5 L. Crombie and S. J. Holloway, *J. Chem. Soc., Perkin Trans. 1*, 1985, 1393.
 - 6 F. Bohlmann, R. K. Gupta, R. M. King, and H. Robinson, *Phytochemistry*, 1981, **20**, 1417; F. Bohlmann, P. Singh, J. Jakupovic, R. M. King, and H. Robinson, *ibid.*, 1982, **21**, 125; F. Bohlmann, N. Borthakur, R. M. King, and H. Robinson, *ibid.*, 1982, **21**, 371. One of the natural compounds reported in the latter paper is given (without stereochemistry) the same structure as 12-oxoPDA (**1**).
 - 7 B. A. Vick and D. C. Zimmerman, *Plant Physiol.*, 1979, **64**, 203.
 - 8 B. A. Vick, F. Feng, and D. C. Zimmerman, *Lipids*, 1980, **15**, 468.
 - 9 L. Crombie and D. O. Morgan, *J. Chem. Soc., Chem. Commun.*, 1988, in the press.
 - 10 L. Crombie and K. M. Mistry, *J. Chem. Soc., Chem. Commun.*, 1988, following communication.
 - 11 R. V. Stevens and N. Hrib, *Tetrahedron Lett.*, 1981, **22**, 4791.
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