Absolute Stereochemistry of Proclavaminic Acid, the Monocyclic Biosynthetic Precursor of Clavulanic Acid

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Proclavaminic acid (1) has been synthesized by a route which indicated the absolute stereochemistry of the two chiral centres to be (2S, 3R).

In previous communications^{1,2} we have described the isolation of proclavaminic acid (1) and its involvement in the biosynthesis of clavulanic acid (2) in *Streptomyces clavuligerus* ATCC 27064. A synthesis of the bioactive enantiomer of proclavaminic acid which did not enable the absolute stereochemistry to be deduced was also reported.³ In order to shed further light on the absolute stereochemistry of proclavaminic acid we report here a further synthesis *via* a resolved β -hydroxyornithine and elaborating the β -lactam ring without altering the chiral centres.

Aldol condensation of 3-(benzyloxycarbonylamino)propionaldehyde⁴ (3) with the masked glycine ester⁵ (4) followed by acid hydrolysis⁶ of the intermediate imine yielded the N^5 -protected β -hydroxyornithine (5)† in 95% yield (diastereoisomer ratio ca. 1:1, erythro:threo) (Scheme 1). Separation of the diastereoisomers was accomplished via the differential solubilities of the hydrochloride salts of (5) or by column chromatography of the phenylacetyl derivatives (6). The less polar diastereoisomer of (6) was assigned the threo configuration by conversion to the deprotected amino acid and showing this material to be identical to threo-\beta-hydroxyornithine synthesized by an independent route.7 Mild base hydrolysis of threo-(6) followed by enzymic deacylation of the resulting acid (7) with immobilised Escherichia coli acylase (E.C. 3.5.1.11) yielded enantiomerically pure (chiral h.p.l.c.) threo-N⁵-benzyloxycarbonyl- β -hydroxyornithine (8) (31%). The known hydrolytic specificity⁸ of the E. coli acylase coupled with the threo relative stereochemistry indicated that the absolute stereochemistry of (8) was (2S, 3R).

The benzyl ester of (8), prepared by the method of Maclaran,9 was treated with an excess of acrylic acid in acetonitrile to yield the β -amino acid (10) [58% from (9)]. Ring closure using the Ohno procedure¹⁰ gave enantiomerically pure β -lactam (11) (51%). Model reactions on enantiomerically pure benzyl threoninates and benzyl N^5 -benzyloxycarbonyl-(S)-ornithinate demonstrated that this method of elaborating the β -lactam ring did not affect the stereochemistry of the chiral centres. Deprotection of (11) afforded proclavaminic acid (12) (85%). In the presence of partially purified clavaminic acid synthase,¹ Fe²⁺, O₂, and 2-ketoglutarate this material was converted to clavaminic acid (13).



⁺ Satisfactory analytical and/or spectroscopic data were obtained for all new compounds.

The synthesis of the racemic *erythro* analogue of proclavaminic acid from *erythro*-(5) was carried out using the route described above. The product was identical to the inactive diastereoisomer described previously³ and similarly was not converted to clavaminic acid (13) by the clavaminic acid synthase system.



Scheme 1. Reagents and conditions: $Z = PhCH_2OCO-$; i, (Me₃Si)₂NLi, tetrahydrofuran (THF), -70 °C; ii, 2 M HCl-ether, NaHCO₃; iii, PhCH₂CO₂H, Et-N=C=N(CH₂)₃NMe₂, HCl, THF; iv, separate diastereoisomers; v, 1 equiv. NaOH, THF-H₂O (2:1); vi, immobilised *E. coli* acylase, pH 7.5, 37 °C; vii, 1 equiv. KOH, MeCOCH₂CO₂Me, MeOH; viii, PhCH₂Br, dimethylformamide (DMF); ix, *p*-MeC₆H₄SO₃H, dioxane-ethyl acetate (3:1), NaHCO₃; x, 10 equiv. CH₂=CHCO₂H, MeCN; xi, di-2-pyridyl disulphide, Ph₃P, MeCN; xii, H₂, Pd-C (10%), EtOH.



On the basis of the above evidence we conclude that proclavaminic acid possesses the (2S,3R) absolute stereochemistry. Therefore the ring closure of the monocyclic proclavaminic acid (12) to the bicyclic clavaminic acid (13) which has the (3S,5S) stereochemistry by clavaminic acid synthase proceeds with retention of stereochemistry at the carbon bearing the carboxy function.

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References

- 1 S. W. Elson, K. H. Baggaley, J. Gillett, S. Holland, N. H. Nicholson, J. T. Sime, and S. R. Woroniecki, J. Chem. Soc., Chem. Commun., 1987, 1736.
- 2 S. W. Elson, K. H. Baggaley, J. Gillett, S. Holland, N. H. Nicholson, J. T. Sime, and S. R. Woroniecki, J. Chem. Soc., Chem. Commun., 1987, 1739.
- 3 K. H. Baggaley, J. T. Sime, N. H. Nicholson, S. W. Elson, J. Gillett, S. Holland, and S. W. Woroniecki, J. Chem. Soc., Chem. Commun., 1987, 1738.
- 4 T. Teshima, K. Konishi, and T. Shiba, Bull. Chem. Soc. Jpn., 1980, 53, 508.
- 5 M. J. O'Donnell and R. L. Polt, J. Org. Chem., 1982, 47, 2663.
- 6 P. Bey, J. P. Vevert, V. van Dorsseler, and M. Kolb, J. Org. Chem., 1979, 44, 2732.
- 7 Y. Ozaki, S. Maedan, M. Miyoshi, and K. Matsumoto, *Synthesis*, 1979, 216.
- 8 D. Rossi, A. Romeo, and G. Lucente, J. Org. Chem., 1978, 43, 2576, and references cited therein.
- 9 J. A. Maclaran, Aust. J. Chem., 1972, 25, 1293; A. J. Bose, M. S. Manhas, J. E. Vincent, K. Gala, and I. F. Fernandez, J. Org. Chem., 1982, 47, 4075.
- 10 S. Kobayashi, T. Imari, T. Iizawa, and M. Ohno, J. Am. Chem. Soc., 1981, 103, 2406.