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Photocyclisation of Haloacetyl Tryptophan Derivatives

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Irradiation of dichloroacetyl tryptophan methyl ester gives, after addition of a nucleophile in work-up, the 7-substituted pyrrolobenzazocines (7); similar cyclisation of the tryptophan derivative (11) gives the pyrrolobenzazocine (12), a compound related to serotobenine.

Of the naturally occurring indoles, those in which the indole 3-position is bridged to the 4-position ([c,d]-fused indoles) are among the most interesting in terms of biological activity. Examples include the well known ergot alkaloids, as well as clavicipitic acid (1),¹ serotobenine (2),² and indolactam V (3),³ where the indole is bridged by 6-, 7-, 8-, and 9-membered rings, respectively. Whilst in nature this class of compound can be synthesised from tryptophan, with few exceptions,⁴ the usual laboratory approach involves the linking together of existing functionality in both the 3- and 4-positions of the indole ring. However, we have adapted a photochemicallyinduced ring closure of simple tryptophan derivatives to the synthesis of [c,d]-fused indoles. The results described herein constitute a useful extension of classic tryptophan chemistry, which has long been used to prepare the well known [b]-fused indoles, to the much more difficult [c,d]-fused systems.

Our photochemical approach is based on an attempted photoreduction of chloroacetyl tryptophan,⁵ which resulted in an unusual cyclisation to the indole 4-position to give compound (4);⁶ such cyclisations are also relevant to the photochemistry of some tryptophan-containing peptides.⁷ However, in our hands the yield of the bridged indole (4) was low (ca. 25%), owing to competing formation of the [b]-fused isomer (5), and attempts to functionalise the 7-position in (4) for further elaboration (e.g. ring expansion) were unsuccessful. We have now overcome these limitations by using dichloroacetyl tryptophan derivatives (6) as the precursors, the additional halogen providing built-in functionality for the new ring. Thus irradiation of (-)-dichloroacetyl tryptophan methyl ester (6a) at 254 nm in acetonitrile gave, after chromatography, the 7-hydroxypyrrolo[4,3,2-f,g][3]benzazocine (7a) (Scheme 1) (58% yield), $[\alpha]_D - 96.4^\circ$ (c 0.44, MeOH), presumably by displacement of the initially formed 7-chloro derivative. Indeed, when the photolysis reaction was worked up in the presence of other nucleophiles such as methanol or hydroxylamine, the corresponding derivatives (7b) {m.p. 190–191 °C (decomp.), $[\alpha]_D$ -59.5° (c 0.37, MeOH)} and (7c) were isolated in 63 and 51% yield, respectively. In all cases the yields are better than that obtained from the monochloro derivative and no competing cyclisation to the indole 2-position was observed. The reaction also exhibits a remarkable degree of asymmetric induction, in



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that the 7-substituent is exclusively *trans* to the ester group, although internal displacement from the *cis* face occurs in the case of the acid (**6b**) and the alcohol (**6c**) to give the novel bridged lactone (**8a**) and ether (**8b**), respectively (Scheme 1). Irradiation of the trichloroacetyl tryptophan ester (**9**) gives, after aqueous work-up, the cyclic keto amide (**10**) (Scheme 2) [75% yield; m.p. 256–258 °C (decomp.)].

Thus these [c,d]-fused indoles can be prepared in two steps (acylation and photocyclisation with nucleophilic work-up) from tryptophan methyl ester. The cyclisation method is extended to the preparation of a compound related to serotobenine (2) via the chlorohydrocinnamoyl tryptophan derivative (11), the irradiation of which in acetonitrile gives the serotobenine analogue (12) (Scheme 3) (55% yield). The application of a dichloroacetyl tryptophan photocyclisation to the total synthesis of (-)-indolactam V (3) is reported in the following communication.

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References

- 1 G. S. King, E. S. Waight, P. G. Mantle, and C. A. Szczyrbak, J. Chem. Soc., Perkin Trans. 1, 1977, 2099.
- 2 H. Sato, H. Kawagishi, T. Nishimura, S. Yoneyama, Y. Yoshimoto, S. Sakamura, A. Furusaki, S. Katsuragi, and T. Matsumoto, *Agric. Biol. Chem.*, 1985, **49**, 2969.
- 3 K. Irie, M. Hirota, N. Hagiwara, K. Koshimizu, H. Hayashi, S. Murao, H. Tokuda, and Y. Ito, *Agric. Biol. Chem.*, 1984, **48**, 1269.
- 4 For example, see S. Nakatsuka, K. Yamada, and T. Goto, *Tetrahedron Lett.*, 1986, **27**, 4757.
- 5 O. Yonemitsu, P. Cerutti, and B. Witkop, J. Am. Chem. Soc., 1966, 88, 3941.
- 6 S. Naruto and O. Yonemitsu, Chem. Pharm. Bull., 1980, 28, 900.
- 7 J. Dillon, Photochem. Photobiol., 1981, 33, 137.