The Three Dimensional Structure of all-*cis*-Diepoxy[15]annulenone and some Implications from the Structure

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An X-ray structural analysis of all-*cis*-diepoxy[15]annulenone (1) revealed that (1) exists in two different conformations, each of which possesses a saucer-like three-dimensional geometry, like that of the polyaromatic 14π hydrocarbon corannulene (5).

Diepoxy[15]annulenone (1),^{1a,b} a higher homologue of tropone, is of interest, since it produces mono-*trans* annulenylium ion (3) as the major product on protonation [*via* (2)], but (3) rapidly reverts to (1) on deprotonation [*via* (4)] (Figure 1). In order to examine the structural basis affording such easy sequential transformations, we have undertaken an X-ray structural analysis of (1) (recrystallized from MeCN). A saucer-like three-dimensional structure is expected for (1), because its perimeter is very similar to that of corannulene (5).² This expectation was confirmed by the structural determination (Figure 2).

The structures of each of the two independent molecules in the asymmetric unit show only slight conformational differences.[†] The *cis*-difuryl ethene moiety is nearly planar in both conformations, holding the furan oxygens 10.6 and 14.9°

[†] Crystal data: monoclinic. space group P2₁/n, a = 16.763(4), b = 11.998(2), c = 12.278(3) Å, $\beta = 106.83(2)^\circ$, Z = 8. 4813 Independent reflections were measured in the ω-2θ scan mode in a Rigaku AFC-5 diffractometer, using graphite-monochromated Mo- K_{α} radiation ($\lambda = 0.7107$ Å). 3978 reflections with $F_{o} \ge 2\sigma(F_{o})$ were used in the structure solution by direct methods and full-matrix least-squares refinement; R = 7.6%. Atomic co-ordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See Notice to Authors, Issue No. 1.



Figure 1. [15]Annulenone-hydroxy[15]annulenylium ion cycle.



upward from the corresponding mean planes of molecules A and B, respectively. The most prominent feature is that each C=O group is inclined at an angle of 57.9° to the mean square plane. This angle is very close to the dihedral angle of cyclo-octatetraene (55.7°) .³ As a consequence, all the bond angles and bond lengths of (1) show approximately normal values, and the C(1)–C(2) single and C(2)=C(3) double bond distances are also close to the corresponding values in cyclo-octatetraene (1.462 and 1.334 Å). The structural data for the difuryl ethene moiety shows good agreement with data for the recently prepared [20]annulene tetraoxide,⁴ which contains two *cis*-difuryl ethene moieties.

From the crystal data we have made the following observations. (i) The molecule consists of two segments, a planar looped segment encompassing atoms C(4)-C(13), and a non-planar flexible segment formed out of the five sp² carbon centres C(3)-C(2)-C(1)-C(15)-C(14) in which bond alternation is prominent. In these segments, the mobilities and planarities are very different, and this dual structural feature of (1) produces a sequential motion that makes (1) fluctuate in conformation.

(ii) The C(1)-C(15) single and C(2)=C(3) double bonds of (1) are nearly parallel to each other (Figure 2). On protonation, the bond angle C(15)-C(1)-C(2) is slightly widened to keep the above two bonds parallel. This induces a single rotational motion around the C(1)-C(2) single bond to cause isomerization (2) \rightarrow (3). Thus, ^{1b} the isomerization becomes an extremely fast process such that it can be detected only by using low-temperature n.m.r. spectroscopy. Conversely, on deprotonation, the mono-*trans*-[15]annulenone (4) should be destabilized by the presence of the inside hydrogen, and isomerization (4) \rightarrow (1) takes place. Consequently, the net result of these two isomerizations is to make a chemical cycle.

(iii) In the presence of H⁺, the flexible segment of (1) can undergo a co-operative H⁺-binding cycle. The conformational change (2) \rightarrow (3) takes place causing better H⁺-binding, since the annulenyl ion (3) can bind to H⁺ more tightly than (2), since (3) binds to H⁺ at the outside of the perimeter to adopt a planar geometry. The segmental motion provides two relationships {[(1)]/[(4)] > 1, pK_a of (2) < pK_a of (3)}⁵ which satisfy two requirements for causing co-operative H⁺-binding.⁶

(iv) The rigid half of (1) keeps both ends of the flexible segment at an almost constant distance, regardless of the cis/trans geometries and of the protolytic/deprotolytic states. Therefore, either 'bicycle pedal' motion or 'concerted twisting motion on the *n*-th sp² carbon atom' (CT-*n* mechanism)⁷ may become responsible for such rapid isomerizations. We suggest that isomerization $(1) \rightarrow (2) \rightarrow (3)$ may be likened to the isomerization sequence $\mathbf{M}_{412}(cis) \rightarrow \mathbf{H}^+ \cdot \mathbf{M}_{412}(cis) \rightarrow \mathbf{O}_{640}$ that occurs in the active site of bacteriorhodopsin,⁸ although these two chromophores are very different from each other (Figure 3). Indeed, the flexible segment of (1) can undergo formally analogous isomerizations (W to sickle transformation) that the specific five sp^2 atoms of retinylidene Schiff's base [C(12), C(13), C(14), C(15), and the N atom] can undergo in the protein (sickle to W transformation), as depicted in Figure 3 (boxed areas).



Figure 2. Structure of $C_{15}H_{10}O_3$ [(1): molecule A]. Bond lengths: C(1)–O(1) 1.217(5), C(1)–C(2) 1.484(5), C(2)–C(3) 1.322(6), C(3)–C(4) 1.439(5), C(4)–O(2) 1.383(4), C(4)–C(5) 1.361(6), C(5)–C(6) 1.413(5), C(6)–C(7) 1.371(6), C(7)–O(2) 1.373(4), C(7)–C(8) 1.432(5), C(8)–C(9) 1.351(6), C(9)–C(10) 1.413(6), C(10)–O(3) 1.379(4), C(10)–C(11) 1.382(6), C(11)–C(12) 1.400(6), C(12)–C(13) 1.383(6), C(13)–O(3) 1.372(5), C(13)–C(14) 1.433(5), C(14)–C(15) 1.325(7), C(15)–C(1) 1.484(7) Å. Bond angles: O(1)–C(1)–C(2) 122.2(4), C(1)–C(2) 127.3(3), C(2)–C(3)–C(4) 128.1(3), C(3)–C(4)–O(2) 116.8(3), C(3)–C(4)–C(5) 133.1(3), O(2)–C(4)–C(5) 109.9(3), C(4)–C(5)–C(6) 107.0(4), C(5)–C(6)–C(7) 106.8(4), C(6)–C(7)–O(2) 109.8(3), C(6)–C(7)–C(8) 129.6(3), O(2)–C(7)–C(8) 120.6(3), C(7)–C(9) 134.0(4), C(8)–C(9)–C(10) 132.5(3), C(9)–C(10)–C(11) 129.6(3), O(3)–C(10)–C(11) 109.0(3), C(10)–C(12) 107.2(3), C(11)–C(12)–C(13) 107.3(4), C(12)–C(13)–O(3) 109.0(3), C(12)–C(13)–C(14) 133.1(4), O(3)–C(13)–C(14) 117.9(4), C(13)–C(14)–C(15) 127.7(5), C(14)–C(15)–C(1) 127.9(4), C(15)–C(1)–O(1) 122.8(4) C(15)–C(1)–C(2) 115.0(3)°.



Figure 3. The proton pumping cycle of bacteriorhodopsin (BR). LA and DA indicate light- and dark-adapted BR, respectively (according to Prof. K. Nakanishi, 1986, Komaba, Tokyo).⁸

In conclusion, although we have not yet found a rigorous answer to determine what types of motion are really responsible for the isomerization $(1) \rightarrow (3)$, we have now obtained a structural basis to understand why (1) can drive a co-operative H⁺-binding cycle with fast isomerization rates.

We thank Drs. Y. Wada and A. Kamiya (Takeda Pharmaceutical Co., Osaka, Japan) for the X-ray structural determination. We also acknowledge a Grant-in-Aid from the Ministry of Education, Japan (63570996).

Received, 11th July 1988; Com. 8/02758G

References

- 1 (a) H. Ogawa, T. Inoue, T. Imoto, I. Miyamoto, H. Kato, and Y. Taniguchi, Angew. Chem., Int. Ed. Engl., 1983, 22, 253; (b) H. Ogawa, T. Inoue, T. Imoto, I. Miyamoto, H. Kato, and Y. Taniguchi, Angew. Chem. Suppl., 1983, 243.
- 2 W. E. Barth and R. G. Lawton, J. Am. Chem. Soc., 1971, 93, 1930.

- 3 M. Traetteberg, Acta. Chem. Scand., 1966, 20, 1724; J. Bordner, R. G. Parker, and R. H. Stanford, Jr., Acta Cystallogr., Sect. B, 1972, 28, 1069.
- 4 E. Vogel, M. Sicken, P. Rohrig, H. Schmichler, J. Lex, and O. Ermer, Angew. Chem., Int. Ed. Engl., 1988, 27, 411.
 5 H. Ogawa, H. Morita, T. Imoto, I. Miyamoto, H. Kato, Y.
- 5 H. Ogawa, H. Morita, T. Imoto, I. Miyamoto, H. Kato, Y. Taniguchi, Y. Nogami, and T. Koga, unpublished results; manuscript is in preparation.
- 6 D. E. Metzler, 'Biochemistry,' International Edn., Academic Press, New York, 1977, p. 221.
- 7 R. S. H. Liu, D. Mead, and A. E. Asato, J. Am. Chem. Soc., 1985, 107, 6609, and references cited therein.
- 8 F. Derguini, D. Dunn, L. Eisenstein, K. Nakanishi, K. Odashima, V. J. Rao, L. Sastry, and J. Termini, *Pure Appl. Chem.*, 1986, 58, 719.