

Enantiomerically Pure Cyclohexanols and Cyclohexane-1,2-Diol Derivatives; Chiral Auxiliaries and Substitutes for (–)-8-Phenylmenthol. A Facile Enzymatic Route

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A number of optically active cyclohexanol and cyclohexane-1,2-diol derivatives, chiral auxiliaries and substitutes for (–)- and (+)-8-phenylmenthol, have been prepared by enzymatic hydrolysis of their racemic acetates and chloroacetates in the presence of a highly selective ester hydrolase from *Pseudomonas sp.* (SAM-II).

Enantiomerically pure compounds with cyclohexanol substructures like (–)- and (+)-menthol [(–)/(+)-(1)] or (–)-8-phenylmenthol (–)-(2) are among the most widely used, classical chiral reagents in organic chemistry, both for analytical and synthetic applications.¹ Although (–)-(2) and (+)-(2) are among the most powerful auxiliaries in asymmetric synthesis,² synthetic routes starting from (+)-pulegone³ or (–)-pulegone,⁴ respectively, are less than satisfactory, often impractical on a synthetic scale. This is also reflected in the high price of these reagents, and so more readily accessible substitutes for (–)- and (+)-(2) are highly desirable. Closely related structures are phenyl- and benzyl-cyclohexanols (*R,S*)- and (*S,R*)-(3),(4), it was shown recently that (*R,S*)-(3) is as powerful as (–)-(2) for efficient absolute stereocontrol.⁵ Also structurally similar to (–)-(2) are the corresponding cyclohexanediol derivatives (*R,R*)- and (*S,S*)-(5),(6), while (*R,R*)- and (*S,S*)-(7),(8) could be of considerable interest as

chiral building blocks or ligands (*e.g.* for the synthesis of chiral crown ethers),⁶ respectively.

In view of the well documented excellent ability of many ester hydrolases for enantiomer differentiation, the enantioselective, enzymatic hydrolysis of racemic esters derived from (±)-(3)—(8) [*e.g.* the acetates (±)-(3a)—(7a),(8c)] seemed to be an obvious and facile approach to this whole class of molecules. Although porcine liver esterase (PLE) can be used for the resolution of (±)-(3a)⁷ and (±)-(8c),⁸ these reactions proved to be less than satisfactory on a large, preparative scale.† We report here successful experiments using a highly

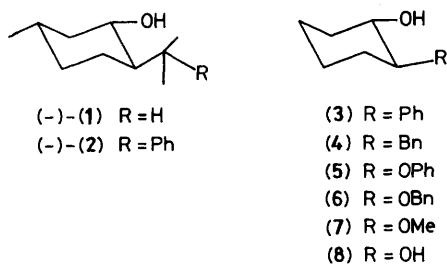
† In our hands, PLE catalysed hydrolysis of (±)-(3a)⁷ was extremely sluggish, incomplete, and suffered from severe product inhibition. The earlier described hydrolysis of (±)-(8c) was difficult to control and, disappointingly, always produced large quantities of racemic monoacetate (±)-(8a).⁸

Table 1. Enzymatic hydrolysis of acetates and chloroacetates derived from (\pm)-(3)—(8).

Entry	Substrate	% Conversion	t/h	Product	% Yield	% E. e. ^{a,b}	<i>E</i> ^c
1	(\pm)-(3a)	27	164	(<i>R,S</i>)-(3)	20	98 ^b	140
				(<i>S,R</i>)-(3a)	64	36	
2	(\pm)-(3b)	50	9 (!)	(<i>R,S</i>)-(3)	44	95 ^b	180
				(<i>S,R</i>)-(3b)	43	97	
3	(\pm)-(4a)	46	53	(<i>R,S</i>)-(4)	40	98 ^b	260
				(<i>S,R</i>)-(4a)	46	83	
4	(\pm)-(4b)	50.8	18	(<i>R,S</i>)-(4)	46	>95 ^a	>145
				(<i>S,R</i>)-(4b)	43	>95	
5	(\pm)-(5a)	48	40	(<i>R,R</i>)-(5)	42	>99 ^b	>790
				(<i>S,S</i>)-(5a)	45	96	
6	(\pm)-(6a)	51	29	(<i>R,R</i>)-(6)	47	>95 ^a	>145
				(<i>S,S</i>)-(6a)	45	>95	
7	(\pm)-(6b)	50	8	(<i>R,R</i>)-(6)	41	88 ^b	40
				(<i>S,S</i>)-(6b)	43	86	
8	(\pm)-(7a)	49	44	(<i>R,R</i>)-(7)	45	98 ^b	400
				(<i>S,S</i>)-(7a)	49	96	
9	(\pm)-(8c)	25.2	17	(<i>S,S</i>)-(8c)	53	79	120
				(<i>R,R</i>)-(8a)	36	96 ^b	
10	(\pm)-(8c)	27.8	22	(<i>S,S</i>)-(8c)	48	84	—
				(<i>R,R</i>)-(8a)	41	94 ^b	
				(<i>R,R</i>)-(8)	10	97	
11	(S,S)-(8c) from entry 10	9.3	19	(<i>S,S</i>)-(8c)	81	97	—
				(<i>R,R</i>)-(8a)	17	82 ^b	

^a Determined by 400 MHz ¹H n.m.r. using Eu(tfc)₃ [tfc = 3-(trifluoromethylhydroxymethylene)-(+)-camphorato] as chiral shift reagent.

^b By g.c. of the isopropylurethanes using a chiral column (ref. 11). ^c See text and ref. 10; *E* = k_R/k_S , calculated ratio of hydrolysis rates for the two enantiomers.



a; acetate derivative
b; chloroacetate derivative
c; diacetate derivative, (8c) only

selective lipase from *Pseudomonas sp.*,[‡] from which this whole class of molecules has become accessible, with excellent chemical and optical yields.

In a series of experiments 50 mmol of the racemic acetates (\pm)-(3a)—(7a) were hydrolysed in the previously described way⁹ using 40 g of 0.1 M phosphate buffer (pH 7, 20°C) and 500 mg (16 000 units, standard: tributyrin) of the lipase. Practically all the reactions came to a near standstill after ca. 50% conversion (*i.e.* hydrolysis of one enantiomer); this is to be expected for a highly selective enantiomer differentiation with values of *E* > 100¹⁰ and further documented by the high enantiomeric purities of all the products obtained (Table 1).

For (\pm)-(3a) a highly selective, but rather slow transformation was observed; this somewhat limits its synthetic usefulness. Clearly, for synthetic applications on a

practically useful scale higher rates of hydrolysis had to be achieved. This problem, encountered previously for other substrates,⁹ was successfully solved by the use of activated esters. As can be seen from Table 1 (compare *e.g.* entries 1 and 2, 3 and 4, 6 and 7) considerably higher rates of conversion were found for the corresponding chloroacetates (\pm)-(3b), (4b), (6b). While in the case of (\pm)-(3b), (4b) the enantiomeric purities obtained remained unchanged in comparison with the corresponding acetates (\pm)-(3a), (4a), a considerable decrease was observed going from (\pm)-(6a) to (\pm)-(6b) (compare Table 1, entries 6 and 7), (\pm)-(6a) in this case clearly being the preferred substrate.

The enantioselective hydrolysis of the diacetate (\pm)-(8c) [derived from (\pm)-(8)] [200 mmol, 40 g 0.1 M phosphate buffer, pH 7, T 20°C, 600 mg (19 800 units; standard: tributyrin) lipase SAM-II] was terminated after ca. 25% conversion (corresponding to the hydrolysis of one of the four esterfunctions in the racemate) leading to two major [(*R,R*)-(8c), (*S,S*)-(8a)] and one minor product (*R,R*)-(8) with the somewhat conversion-dependent enantiomeric purities listed in Table 1. While (*R,R*)-(8) both directly, and *via* the chemical hydrolysis (K₂CO₃, MeOH) of (*R,R*)-(8c), can be obtained nearly enantiomerically pure, changes in the conversion did not yield the monoacetate (*S,S*)-(8a) with high optical purity. It was finally obtained with 97% enantiomeric excess (*e.e.*) by enzymatic (low conversion) hydrolysis of enantiomerically enriched (*S,S*)-(8c) (Table 1, entry 11).

All products were isolated by the extraction of the crude reaction mixtures with Et₂O, followed by column chromatography on silica gel (petrol, ether). Acetates and chloroacetates were converted into the corresponding alcohols (K₂CO₃, MeOH), the enantiomeric purities were determined either by (i) g.c. separation of the isopropyl urethanes on a chiral g.c. column¹¹ or (ii) 400 MHz ¹H n.m.r. studies using Eu(tfc)₃ as

[‡] Lipase SAM-II from Amano Pharmaceutical Co., supplied by Fluka AG, CH-9470 Buchs, Switzerland (Cat. No. 62312) and Mitsubishi Int. GmbH, D-4000 Düsseldorf, Germany.

chiral shift reagent, observing the signal of the methyl group in the acetate functions (see Table 1).

Absolute configurations were either known [(*R,S*)-, (*S,R*)-(**3**)]⁵ or correlated by (i) oxidation of (1*R,2S*)-(**4**) to the known (*S*)-2-benzylcyclohexanone,§ (ii) chemical degradation (ether cleavage) of (*R,R*-, (*S,S*)-(**5**)-(7) to the known (*R,R*)- and (*S,S*)-(**8**) whose absolute configurations were confirmed earlier.⁸

We feel that the results in this paper further demonstrate the synthetic usefulness of ester hydrolases, allowing the preparation of a whole class of compounds with high enantiomeric purity and in synthetically useful quantities.

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§ Oxidation of (1*R,2S*)-(**4**) (pyridinium dichromate, CH₂Cl₂, room temp., 77%) produced the known¹² (*S*)-2-benzylcyclohexanone with the highest optical rotation recorded {[α]_D²⁵ -47°, *c* 3.8 MeOH; lit.,¹² [α]_D²⁵ -41.4°, *c* 4.9 MeOH}.

References

- 1 H. Kipphardt and D. Enders, *Kontakte (Merck)*, 1985 (2), 37; J. Sauer and J. Kredel, *Angew. Chem.*, 1965, **77**, 1037; *Tetrahedron Lett.*, 1966, 6359; W. Oppolzer, M. Kurth, D. Reichlin, C. Chapuis, M. Mohnhaupt, and F. Moffat, *Helv. Chim. Acta*, 1981, **64**, 2802; G. Quinkert and H. Stark, *Angew. Chem.*, 1983, **95**, 651.
- 2 J. K. Whitesell, D. James, and J. F. Carpenter, *J. Chem. Soc., Chem. Commun.*, 1985, 1449 and other papers in that series.
- 3 J. K. Whitesell, C. L. Liu, C. B. Buchanan, H. H. Chen, and M. A. Minton, *J. Org. Chem.*, 1986, **51**, 551.
- 4 E. J. Corey and H. E. Ensley, *J. Am. Chem. Soc.*, 1975, **97**, 6908; E. J. Corey, H. E. Ensley, and J. W. Suggs, *J. Org. Chem.*, 1976, **41**, 380; H. E. Ensley, C. A. Parnell, and E. J. Corey, *ibid.*, 1978, **43**, 1610.
- 5 J. K. Whitesell, H. H. Chen, and R. M. Lawrence, *J. Org. Chem.*, 1985, **50**, 4663.
- 6 R. C. Hayward, C. H. Overton, and G. H. Whitham, *J. Chem. Soc., Perkin Trans. 1*, 1976, 2413.
- 7 J. K. Whitesell and R. M. Lawrence, *Chimia*, 1986, **40**, 318.
- 8 D. H. G. Crout, V. S. B. Gaudet, K. Laumen, and M. P. Schneider, *J. Chem. Soc., Chem. Commun.*, 1986, 808.
- 9 K. Laumen and M. P. Schneider, *J. Chem. Soc., Chem. Commun.*, 1988, 598.
- 10 C. S. Chen, Y. Fujimoto, G. Girdaukas, and C. J. Sih, *J. Am. Chem. Soc.*, 1982, **104**, 7294.
- 11 XE-60-L-Valine-(*S*)- α -phenylethylamide, commercially available from: Chrompack International, Middelburg, The Netherlands; W. A. König, W. Franke, and J. Benecke, *J. Chromatogr.*, 1982, **239**, 227.
- 12 A. I. Meyers, D. R. Williams, and M. Druelinger, *J. Am. Chem. Soc.*, 1976, **98**, 3032.