

The Fate of Valine-Oxygen during Penicillin Biosynthesis

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[4-²H₆, ¹⁸O₂]valine has been fed to intact cells of *Cephalosporium acremonium*; isolation of the biosynthesised α-aminoadipoylcysteinyl valine (ACV) tripeptide has demonstrated that an intracellular exchange of *one* and *both* valine oxygens occurs.

Penicillin biosynthesis from L-valine, L-cysteine, and L-α-aminoadipic acid is known¹ to proceed *via* the tripeptide L-α-aminoadipoyl-L-cysteinyl-D-valine, LLD-ACV (**1a**). During the course of formation of the tripeptide (**1a**) the stereochemistry of the valinyl residue is inverted. Exogenous feeding experiments of L-[¹⁸O₂]valine to *Penicillium chrysogenum* performed by Thomas² suggested the elimination of *one* of the *two* carboxy-oxygen atoms during the overall conversion of L-valine to penicillin V (**2a**). From this it was concluded that 'the elimination of a valine-oxygen atom could take place either in the course of biosynthesis of this tripeptide [ACV (**1**)] or its subsequent oxidative cyclisation to the penam nucleus.' Subsequently we showed³ that *no* oxygen atoms were exchanged during the conversion of the tripeptide (**1a**) to isopenicillin N (**2b**) [the biosynthetic precursor of (**2a**)]. We now report that valine is transformed by intact cells of the β-lactam negative mutant, N-2, of *Cephalosporium acremonium*⁴ A.T.C.C. 14553 into ACV (**1**) with both partial and complete loss of both oxygen atoms.

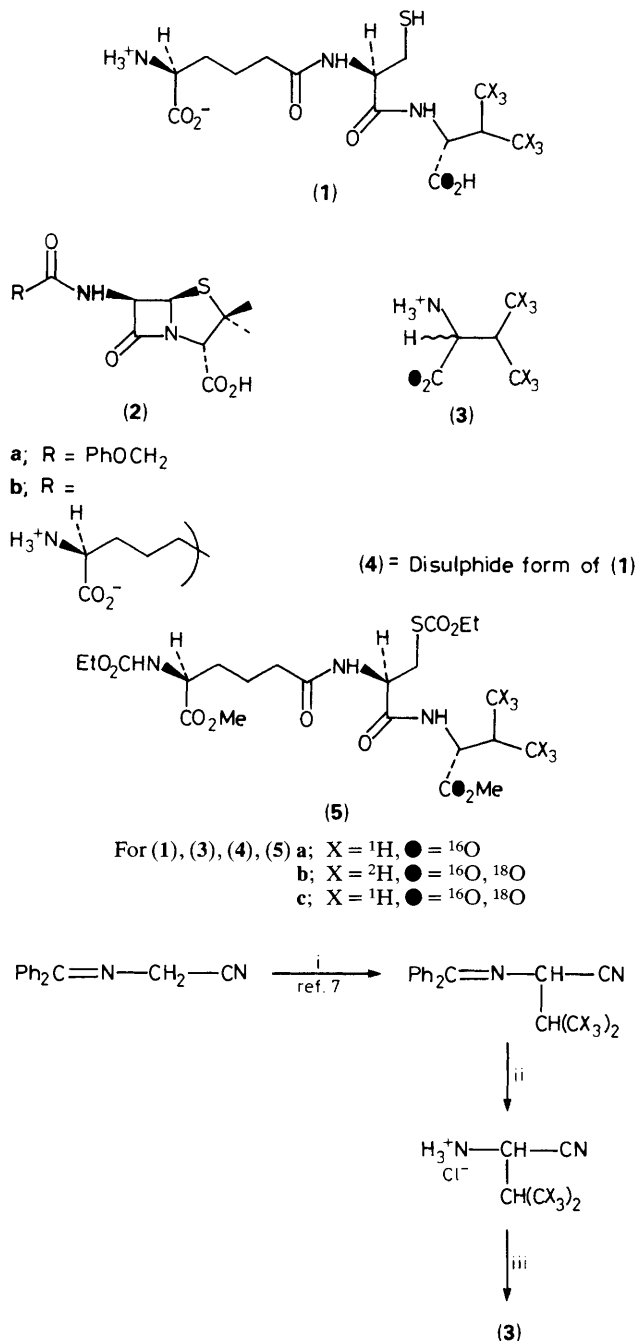
A possible problem of the previous experiment² is that if *both* oxygen atoms of the exogenously labelled valine were exchanged prior, during, or subsequent to assembly of tripeptide (**1**), then such exchange would be masked by the endogenous production of ACV (**1**). In order to avoid such problems [4-²H₆, ¹⁸O₂]valine (**3b**) was synthesised (*ca.* 70% from labelled 2-bromopropane; Scheme 1) so that the fate of the exogenous precursor could be followed discretely. In addition the use of a β-lactam negative Takeda N-2 mutant of

C. acremonium,⁴ an organism which excretes ACV (**1**) and has no active penicillin-forming enzyme available, allowed direct examination of the incorporation of valine into the tripeptide (**1**), rather than in the downstream penicillin (**2**).²

Thus racemic† [4-²H₆, ¹⁸O₂]valine (**3b**) was fed at a level of 12.5 mg ml⁻¹ to a culture of Takeda N-2 mutant of *C. acremonium*⁴ as soon as the stationary phase was established. Fermentation was continued for 3 days at 250 rev min⁻¹. The culture was sonicated, centrifuged, and the supernatant filtered to provide a crude lysate from which ACV disulphide (**4**) and recovered valine (**3**) were isolated by reverse phase h.p.l.c. [octadecylsilane, (i) MeOH: 25 mM NH₄HCO₃ (1:4); (ii) 25 mM NH₄HCO₃ as eluants]. The ACV disulphide (**4**) was then reduced [dithiothreitol (5 equiv.); water, 5 min] and converted to its *N,S*-diethoxycarbonyldimethyl ester derivative (**5**).³ The recovered valine (**3**) and ACV derivative (**5**) were then examined by mass spectrometry (Table 1, entries 1,2,3). This data indicates substantial loss of both *one* and *two* oxygen atoms from valine (**3**) to ACV (**1**).

Control experiments were also performed. Thus substitution of valine (**3b**) by [¹⁸O₂]ACV disulphide (**4c**) demonstrated that there was no extensive extracellular exchange of

† Only L-valine is thought to be converted to ACV (**1**) in broken cell experiments (*ex. C. acremonium* C-91^{5a} or C-10^{5b}) but it is known that both L- and D-forms contribute to the intracellular pool of L-valine when added to intact cells of *P. chrysogenum*⁶ (although incorporation of D-valine into the mycelium is much slower).



Scheme 1. Reagents: i, (CX₃)₂CHBr, PhCH₂NEt₃Cl⁻, 50% aq. NaOH, 0°C → room temp. over 3 h, then room temp., 12 h; ii, 1 M HCl, 25°C, 12 h; iii, H₂¹⁸O, HCl(g) (satd.), reflux, 12 h.

valinyl oxygens from the tripeptide (Table 2, entries 1, 2) and a culture fluid only experiment (which had been separated from mycelia grown to the stationary phase) with [¹⁸O₂] valine gave no labelled ACV (1), demonstrating that the conversion of valine (3) to ACV (1) was intracellular.

We have demonstrated that an intracellular exchange of one or both valinyl oxygen atoms occurs prior to ACV (1) excretion from intact cells of the N-2 mutant of *C. acremonium*. Whether such exchange could result from the operation of processes not related to penicillin biosynthesis, during tripeptide assembly, at the tripeptide (1) level, or a combination of such events, is uncertain.

Table 1. NH₃ desorption chemical ionisation results: labelled valine (3b) experiment.

Entry	Valine (3b)	m/z	123	124	125	126	127	128	129
1	[1- ¹⁶ O ₂ , 4- ² H ₆]	(%)obs.	5	9	7	53	13	100	10
	Synthetic		●	—	¹⁶ O ₂	—	¹⁸ O ¹⁶ O	—	¹⁸ O ₂
Tripeptide derivative ^a		m/z	541	542	543	544	545	546	547
2	(5b) [from (3b)]	(%)obs.	5	77	32	100	35	19	5
			●	—	¹⁶ O ₂	—	¹⁸ O ¹⁶ O	—	¹⁸ O ₂
Recovered valine (3b)		m/z	123	124	125	126	127	128	129
3	[1- ¹⁶ O ₂ , 4- ² H ₆]	(%)obs.	1	14	5	61	11	100	6

^a Spectrometry also revealed endogenous ACV (1a) production (ca. 33% of exogenous derived production), (5a) m/z obs. 535(1%), 536(100), 537(28), 538(12), 539(4); calcd. for C₂₂H₃₈N₃O₁₀S [MH]⁺ 535(0), 536(100), 537(28), 538(10).

Table 2. NH₃ desorption chemical ionisation results: extracellular exchange control experiment.

Entry	m/z	536	537	538	539	540	541	542	543
1	Derivative (5c) from tripeptide disulphide (4c) before incubation	(%)obs.	1	1	29	12	100	30	4
			●	¹⁶ O ₂	—	¹⁸ O ¹⁶ O	—	¹⁸ O ₂	—
		m/z	536	537	538	539	540	541	542
2	Derivative (5c) from recovered tripeptide (4c) after incubation	(%)obs.	45 ^a	14 ^a	38 ^a	13	100	29	12
			●	¹⁶ O ₂	—	¹⁸ O ¹⁶ O	—	¹⁸ O ₂	—

^a These values are effected by endogenous ACV (1a) production.

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