

p-[³H]-*m*-Azidophenyl Acetic Acid, a Useful Reagent for the Synthesis of Radioactive Photoaffinity Ligands. Synthesis of Photoaffinity Labelling Ecdysones

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The synthesis of [³H]-*p*-azidophenyl acetic acid and its use in the preparation of aryl azido photoaffinity labelled 20-hydroxyecdysone analogues is described.

20-Hydroxyecdysone **1** is the steroid moulting hormone of arthropods and, like other steroids, is a regulator of gene expression.¹⁻³ Although the molecular biology of its action is being studied in numerous laboratories,⁴⁻⁶ details of the ecdysone-receptor interaction are largely unknown. The synthesis of high affinity radiolabelled hormone analogues such as [¹²⁵I]-26-iodoponasterone A have facilitated detection of ecdysone receptors.^{7,8} We are now attempting to utilize radioactive photoaffinity labelling probes for receptor isolation and furthermore, to characterize the steroid binding site.

Radiolabelled photoaffinity ligands are light-activatable moieties that form reactive intermediates, *e.g.* carbenes and nitrenes, which insert into the amino acids of a receptor site and radioactively tag the receptor for isolation and structural studies. Several photolabile functionalities are widely utilized including diazoketones, diazoacetates, aziridines and aromatic azides.⁹ Of these moieties, the aromatic azides containing ³H or ¹²⁵I are synthetically the most readily incorporated in biologically active molecules where they form a reactive nitrene upon irradiation with 254 nm light. Although the specific activity of tritium is *ca.* 10–100 times lower than ¹²⁵I, the use of tritium as a radiolabel is often preferable to ¹²⁵I because of its longer half-life (12.3 years for tritium *vs.* 60 days for ¹²⁵I); also, loss of iodine and subsequent non-specific labelling with free iodine is often associated with the photolysis of iodinated molecules.¹⁰⁻¹²

We report the synthesis of two [³H]-photoaffinity labelling ecdysteroid analogues, 2-(*p*-azidophenylaceto)-20-hydroxy-

ecdysone (APA-20-hydroxyecdysone) **2b** and 26-(*p*-azido-phenylaceto)-inokosterone (APA-inokosterone) **3b** which competitively bind with the ecdysteroid receptor. We also describe a three step synthesis of *p*-azido-*m*-[³H]-phenyl acetic acid (APAA) **4b**¹³ from *p*-aminophenyl-acetic acid **5a** which can be coupled with nucleophiles (alcohols and amines), and its application to the photoaffinity labelling of ecdysone ligands. The *p*-azidophenyl acetic acid moiety was chosen

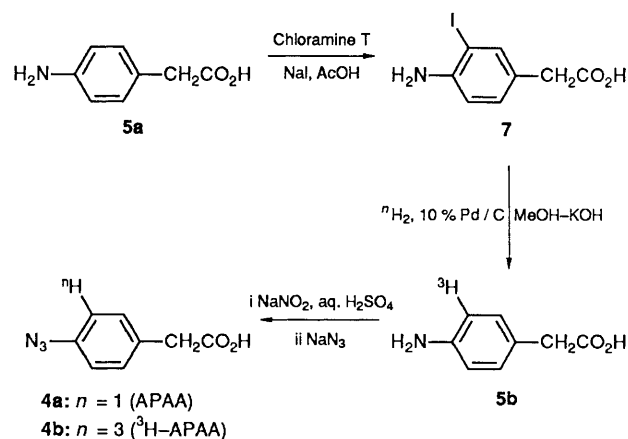


Fig. 1

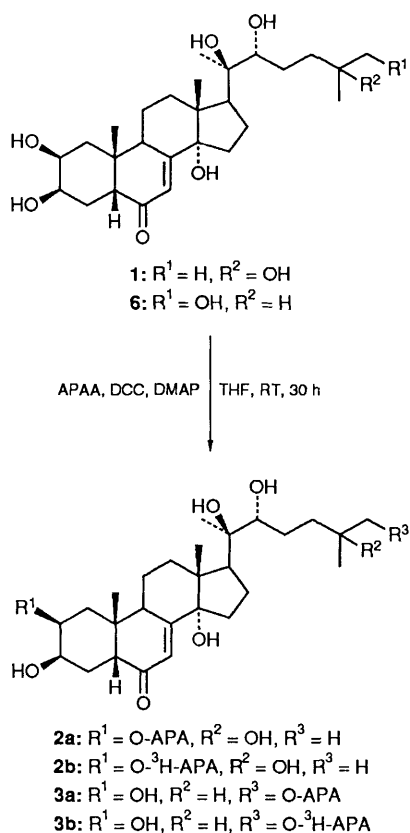


Fig. 2

because initial attempts to perform selective dicyclohexylcarbodiimide (DCC) coupling of *p*-azido benzoic acid with a hindered primary alcohol such as that found at C-26 of inokosterone **6**, resulted in no reaction, presumably because of the low reactivity of the benzoic acid. *p*-Azidophenyl acetic acid is more reactive than *p*-azidobenzoic acid, readily coupling with ecdysteroids and is proving to be useful in other systems containing free hydroxy's and/or amines (unpublished results).

p-Aminophenyl acetic acid **5a** was treated with chloramine T and NaI in acetic acid to give *p*-amino-*m*-iodophenyl acetic acid **7** in 41% yield (Fig. 1).^{14,15} Catalytic tritiation of *p*-amino-*m*-iodophenyl acetic acid in MeOH-KOH with 10% Pd/C and carrier-free ³H₂ resulted in *p*-amino-*m*-³H-phenyl acetic acid **5b** with specific activity of 19 Ci/mmol.† The phenyl amines **5a** and **5b** were oxidized with NaNO₂ in aqueous H₂SO₄, and treated with NaN₃ to give *p*-azido-*m*-ⁿH-phenyl acetic acids (APAA) **4a** (*n* = 1) and (³H-APAA) **4b** (*n* = 3) (89% yield, specific activity 19 Ci/mmol), UV (MeOH) λ_{max} = 252 nm, ε = 12,000.^{13,14} A mixture of 20-hydroxyecdysone **1** and inokosterone **6**‡^{16,17} (ratio = 1.3 : 1), extracted from the root of *Achyranthes fauriei*,^{18,19} was treated with 1.5 equiv. each of *p*-APAA **4a**, DCC and dimethylaminopyridine in dry THF for 30 h to give a 1:1 mixture of 2-(*p*-APA)-20-hydroxyecdysone **2a** and 26-(*p*-APA)-inokosterone **3a** in approximately 35% combined yield after HPLC¹⁶ purification (Fig. 2): HRFAB-MS 640.3578, calcd. for C₃₅H₅₀N₃O₈

† Catalytic tritiation was performed by Amersham using carrier-free tritium gas.

‡ Inokosterone **6** is a mixture of C-25 epimers.

640.3597; UV (MeOH) λ_{max} = 250 nm, ε = 18000. § Similarly, the radiolabelled ecdysone analogues were synthesized from ³H-APAA **4b** to give ecdysteroids **2b** and **3b** at specific activity > 6 Ci/mmol. The products coeluted with authentic samples of ecdysteroids **2a** and **3a**.

Preliminary biological tests were designed to test competition by the azidophenyl ecdysone analogues (in the absence of light) for specific [¹²⁵I]-26-ponasterone A binding in a Kc cell extract.⁷ At 1 μmol dm⁻³ concentration, both analogues compete poorly. However, after photolysis (254 nm), each reduced specific binding of the radioligand by 20–40%. Further photoaffinity studies are ongoing.

We are grateful to Dr. K. Nomoto, Suntory Institute for Biorganic Research, Osaka, for providing us with *Achyranthes fauriei*. The studies were supported by National Institutes of Health awards AI 10187 (K. N.) and GM 13552 (M. F. B.).

Received, 30th August 1990; Com. 0/03927F

References

- 1 *Fundamentals of Insect Physiology*, ed. M. S. Blum, Wiley, New York, 1985.
- 2 M. Raabe, *Recent Developments in Insect Neurohormones*, Plenum Press, New York, 1989.
- 3 V. B. Wigglesworth, *Insect Physiology*, 8th edn., Chapman & Hall, New York, 1984.
- 4 L. Cherbas, H. Benes, M. Bourouis, K. Burtis, A. Chao, P. Cherbas, M. Crosby, M. Garfinkel, G. Guild, D. Hogness, J. Jami, C. W. Jones, M. Koehler, J.-A. Lepesant, C. Martin, F. Maschat, P. Mathers, E. Meyerowitz, R. Moss, R. Pictet, J. Rebers, G. Richards, J. Roux, R. Schulz, W. Segraves, C. Thummel and K. Vijayraghavan, *Insect Biochem.*, 1986, **16**, 241.
- 5 H. J. Bidmon and T. J. Sliter, *Invertebr. Reprod. and Devel.*, 1990, **18**, 13.
- 6 W. A. Segraves and G. Richards, *Invertebr. Reprod. and Devel.*, 1990, **18**, 67.
- 7 P. Cherbas, L. Cherbas, S.-s. Lee and K. Nakanishi, *Proc. Natl. Acad. Sci. USA*, 1988, **85**, 2096.
- 8 S.-S. Lee, K. Nakanishi and P. Cherbas, *J. Chem. Soc., Chem. Commun.*, preceding Communication.
- 9 H. Bayley, *Photogenerated Reagents in Biochemistry and Molecular Biology*, Elsevier, New York, 1983.
- 10 D. S. Watt, K. Kawada, E. Leyva and M. S. Platz, *Tetrahedron Lett.*, 1989, **30**, 899.
- 11 U. Henriksen and O. Buchard, *Tetrahedron Lett.*, 1990, **31**, 2443.
- 12 P. Cherbas, M. F. Boehm and K. Nakanishi, unpublished results.
- 13 Myers and Utter report the synthesis of *p*-azidophenyl acetic acid for use in the enzymatic synthesis of photoaffinity analogues of benzoyl-coenzyme A, however, radiolabelled *p*-azidophenyl acetic acid was not synthesized. D. E. Myers and M. F. Utter, *Anal. Biochem.*, 1981, **112**, 23.
- 14 K. Kawada, E. K. Dolence, H. Morita, T. Kometani, D. S. Watt, A. Balapure, T. A. Fitz, D. J. Orlicky and L. E. Gerschenson, *J. Med. Chem.*, 1989, **32**, 256.
- 15 T. Kometani, D. S. Watt and J. Tae, *Tetrahedron. Lett.*, 1985, **26**, 2043.
- 16 S. Ogawa, A. Yoshida and K. Reiko, *Chem. Pharm. Bull.*, 1977, **25**, 904.
- 17 H. Hikino, M. Mohri, y. Hikino, S. Arihara, T. Takemoto, H. Mori and K. Shibata, *Tetrahedron*, 1976, 3015.
- 18 T. Takemoto, S. Ogawa and N. Nishimoto, *Yakugaku, Zasshi*, 1967, **87**, 1463.
- 19 *Advances in Natural Products Chemistry*, eds. S. Natori, N. Ikekawa and M. Suzuki, Wiley, New York, 1981.

§ Selected spectroscopic data for: **2a** ¹H NMR ([²H₆]MeOH) δ 0.79 (s, 13-CH₃), 0.89 (s, 10-CH₃), 1.09 [s, 25-(CH₃)₂], 1.10 (s, 20-CH₃), 2.31 (dd, *J* 9.1 and 4.4 Hz, 5-CH), 3.20 (m, 9-CH), 3.58 (s, Ph-CH₂), 4.01 (m, 3-CH), 4.89 (dt, *J* 14.8, 3.8 and 3.8 Hz, 2-CH), 5.72 (d, *J* 2 Hz, 7-CH), 6.93 (d, *J* 8.5 Hz, Ph-H), 7.24 (d, *J* 8.5 Hz, Ph-H).

3a ¹H NMR ([²H₆]MeOH) δ 0.79 (s, 13-CH₃), 0.82 (d, *J* 6.9 Hz, 25-CH₃), 0.87 (s, 10-CH₃), 1.07 (s, 20-CH₃), 2.27 (dd, *J* 11.7 and 5.7 Hz, 5-CH), 3.06 (m, 9-CH), 3.55 (s, Ph-CH₂), 3.73 (dt, *J* 12.1, 3.7 and 4.1 Hz, 2-CH), 3.85 (m, 3-CH + 26-CH₂), 5.71 (d, *J* 2.0 Hz, 7-CH), 6.93 (d, *J* 8.4 Hz, Ph-H), 7.22 (d, *J* 8.4 Hz, Ph-H).