Structure and Stereochemistry of Amorphispironone, a Novel Cytotoxic Spironone Type Rotenoid from *Amorpha fruticosa*

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A novel cytotoxic spironone type rotenoid, amorphispironone **1** has been isolated from the leaves of *Amorpha fruticosa* and its structure and stereochemistry have been established from spectral data in conjunction with single-crystal X-ray analysis.

Crude extracts of *Amorpha fruticosa* have been shown to exhibit feeding deterrence along with insecticidal, antiparasitic, antimicrobial and hypotensive activities.' As a result of our continuing searches for novel cytotoxic antitumour compounds from plants, amorphispironone **1,** a novel rotenoid, has been isolated from the leaves of *Amorphafruticosa* as an active principle.[†] We report herein, the isolation and characterization of **1.**

Fig. 1 Structure and solid-state conformation of amorphispironone **1;** small circles represent hydrogen atoms

[†] Amorphispironone (1) showed significant (ED₅₀ $\leq 4.0 \text{ }\mu\text{g m}^{-1}$) selective cytotoxicity in KB (ED₅₀ = 0.58 μ g ml⁻¹) and RPMI (malignant melonoma) $(ED_{50} = 0.61 \,\mu g \,\text{m}^{-1})$ cells. Compound 1 was inactive against A-549 (lung), HCT-8 (colon) and TE671 (human medulloblastoma) tumour cells at $10 \mu g$ ml⁻¹. The cytotoxicity assay was carried out according to literature methods.^{6,7}

Amorphispironone $1\frac{1}{4}$ $\{C_{23}H_{22}O_7$, colourless crystals from 80% aqueous MeOH, m.p. 152-152.5 "C, **[aID** -61.6" (c0.162 in MeOH), λ_{max} nm (MeOH) (log ε) 269 (4.51) and 316 (4.05)) was isolated from the crude chloroform extract of *Amorpha fruticosa* by silica gel chromatography. Many rotenoids have been isolated from the fruit^{2,3} and the root bark4 of this same plant. The general features of the NMR spectra of **1** suggested that it also was a rotenoid. Comparison of the 1H NMR spectral data for **1** with those of the known rotenoid deguelin **25** revealed that the *C-D-E* ring protons were similar but the *A-B* ring protons were different. These data suggested that **1** was a rotenoid, like **2** but differing in the *AIB* ring region. The complete structure and stereochemistry of amorphispironone were established unequivocally by single-crystal X-ray analysis. *\$7* A view of the solid-state conformation is provided in Fig. 1. In general, bond lengths

\$ EI mass *mlz:* 410.1364 (M+). C23H2207 requires *mlz* 410.137. IR (KBr): vmax/cm-l 3010, 2970, 1665, 1640, 1625, 1575, 1393, 1270, 1110, 1070 and 820. NMR spectral assignments were made on the basis of ¹H-¹³C COSY and long range ¹H-¹³C COSY spectra. ¹H NMR (CDCl₃, 300 MHz, *J* in Hz) δ 5.02 (1H, s, 1-H), 5.52 (1H, s, 4-H), 4.50 (1H, dd, *J* 3 and 10, 6-H_a), 4.64 (1H, d, *J* 10, 6-H_b), 5.27 (1H, dd, *J* 3 and 4.5, 6a-H), 6.50 (1H, d, *J* 9, 10-H), 7.67 (1H, d, *J* 9, 11-H), 3.44 (lH, d, J4.5, 12a-H), 3.25 (3H, **s,** 2-OMe), 3.84 (3H, s, 3-OMe), 6.67 (1H, d, J 10, 4'-H), 5.64 (1H, d, J 10, 5'-H) and 1.44 (3H, **s,** 7'-H or 8'-H), 1.47 (3H, **s,** 7'-Hor 8'-H). 13C NMR (CDC13, 75 MHz) 6 105.8 (d, C-l), 84.1 **(s,** C-la), 148.3 (s, C-2), 166.2 (s, C-3), 99.8 (d, C-4), 198.8 (s, C-4a), 76.0 (t, C-6), 82.7 (d, C-6a), 160.1 (s, C-7a or C-9), 109.4 (s, C-8), 156.3 **(s,** C-7a or C-9), 111.7 (d, C-lo), 127.6 (d, C-ll), 114.5 **(s,** C-lla), 186.0 **(s,** C-12), 60.1 (d, C-l2a), 55.1 (4, C-2-OMe), 55.7 (q, C-3-OMe), 115.6 (d, C-4'), 129.2 (d, C-57, 77.7 (s, C-6'), 28.0 (q, C-7' or C-8') and 28.2 (q, C-7' or C-8').

§ Crystal data for 1: $C_{23}H_{22}O_7$, $M = 410.43$, monoclinic, space group $P2_1$, $a = 12.596(2)$, $b = 8.635(1)$, $c = 9.763(1)$ Å, $\beta = 102.03(1)$ ^o, $U =$ 1038.6(4) Å³, $Z = 2$, $D_c = 1.312$ g cm⁻³, μ (Cu-K α radiation, $\lambda =$ 1.5418 \hat{A}) = 7.7 cm⁻¹. Intensity data $[\pm h, + k, + l; \theta_{\text{max}} = 75^{\circ}$, scan width $(0.75 + 0.14\tan\theta)$ °; 2279 non-equivalent reflections] were recorded on an Enraf-Nonius CAD-4 diffractometer (Cu-Ka radiation, graphite monochromator). The crystal structure was solved by direct methods (MULTAN11/82). Full-matrix least-squares refinement of atomic parameters (anisotropic C, 0; isotropic H) converged at $R = 0.032$ $(R_w = 0.047)$ over 1975 reflections with $I > 3.0\sigma(I)$. Crystallographic calculations were performed by use of the Enraf-Nonius SDP package. Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See Notice to Authors, Issue No. 1.

'T[The absolute stereochemistry represented by structure **1** could not be established from the X-ray data. It follows, however, by assuming identical configurations at common asymmetric centres in **1** and another rotenoid derivative with which it co-occurs and for which the absolute stereochemistry has been established in our laboratory by an X-ray crystallographic analysis.

Scheme 1

agree well with expected values. Bond strain, however, is evident in the elongated $C(1a) - C(12a)$ [1.572(3) Å] and C(1a)-C(4a) [1.548(4) Å] bonds. Ring A is fairly flat but puckered slightly towards a shallow envelope form with $C(1a)$ as the out-of-plane atom, \parallel rings B and C have envelope conformations with C(6a) as the out-of-plane atom in each case, ring D is planar, while ring E approximates to a 1,3-diplanar form.

The unusual spiro $A-B$ ring system in 1 is probably biogenetically derived from 2 in the manner indicated in Scheme 1.

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References

- 1 Z. Rozsa, J. Hohmann, K. Szendrei, J. Reisch and I. Mester, Heterocycles, 1982, 19, 1793.
- 2 T. Somleva and U. Ognyanov, Planta Med., 1985, 51, 219.
- 3 L. A. Mitscher, A. Al-Shamma, T. Haas, P. B. Hudson and Y. H.
- Park, Heterocycles, 1979, 12, 1033. 4 J. Hohmann, Z. Rozsa, J. Reisch, and K. Szendrei, Herba Hung., 1982, 21, 179.
- 5 L. Crombie and J. W. Lown, J. Chem. Soc., 1962, 775; D. G. Carlson, D. Weisleder and W. H. Tallent, Tetrahedron, 1973, 29, 2731
- 6 K. H. Lee, Y. M. Lin, T. S. Wu, D. C. Zhang, T. Yamagishi, T. Hayashi, I. H. Hall, J. J. Chang, R. Y. Wu and T. H. Yang, Planta Med., 1988, 54, 308.
- 7 R. I. Geran, N. H. Greenberg, M. M. MacDonald, A. M. Schumacher and B. J. Abbott, Cancer Chemother. Rep., 1972, 3, 1.

[|] Endocyclic torsion angles (ω_{ij} , $\sigma \pm 0.2-0.4^{\circ}$) about the bonds between atoms i and j follow: $\omega_{1a,1}$ 12.8, $\omega_{1,2}$ – 3.3, $\omega_{2,3}$ – 6.4, $\omega_{3,4}$ 4.9, 04, 4a 5.6, $\omega_{4,1a}$ - 13.8° in ring A; ω_{1a} , 0.1, $\omega_{5,6}$ - 24.1, $\omega_{6,6a}$ 38.4,
 $\omega_{6a,12a}$ - 37.2, $\omega_{12a,1a}$ 23.5° in ring B; $\omega_{6a,7}$ 45.4, $\omega_{7,7a}$ - 19.0, $\omega_{7a,11a}$

- 6.3, $\omega_{11a,12}$ 2.8, $\$ ring E .