## Structure and Stereochemistry of Amorphispironone, a Novel Cytotoxic Spironone Type Rotenoid from Amorpha fruticosa

Leping Li,<sup>a</sup> Hui-Kang Wang,<sup>a</sup> Toshihiro Fujioka,<sup>a</sup> Jer-Jang Chang,<sup>b</sup> Mutsuo Kozuka,<sup>c</sup> Takao Konoshima,<sup>c</sup> James A. Estes, <sup>d</sup> Donald R. McPhail, <sup>e</sup> Andrew T. McPhail\* <sup>e</sup> and Kuo-Hsiung Lee\* <sup>a</sup>

<sup>a</sup> Natural Products Laboratory, Division of Medicinal Chemistry and Natural Products, School of Pharmacy, University of North Carolina, Chapel Hill, North Carolina 27599, USA

<sup>b</sup> Laboratory of Animal Medicine, School of Medicine, University of North Carolina, Chapel Hill, North Carolina 27599, USA

c Kyoto Pharmaceutical University, Misasagi, Yamashina-Ku, Kyoto 607, Japan <sup>d</sup> Department of Botany, The University of Oklahoma at Norman, Norman, Oklahoma 73019, USA

e Department of Chemistry, Paul M. Gross Chemical Laboratory, Duke University, Durham, North Carolina 27706,

USA

A novel cytotoxic spironone type rotenoid, amorphispironone 1 has been isolated from the leaves of Amorpha fruticosa and its structure and stereochemistry have been established from spectral data in conjunction with single-crystal X-ray analysis.

Crude extracts of Amorpha fruticosa have been shown to exhibit feeding deterrence along with insecticidal, antiparasitic, antimicrobial and hypotensive activities.1 As a result of our continuing searches for novel cytotoxic antitumour compounds from plants, amorphispironone 1, a novel rotenoid, has been isolated from the leaves of Amorpha fruticosa as an active principle.<sup>†</sup> We report herein, the isolation and characterization of 1.



Fig. 1 Structure and solid-state conformation of amorphispironone 1; small circles represent hydrogen atoms

<sup>†</sup> Amorphispironone (1) showed significant (ED<sub>50</sub>  $\leq$ 4.0 µg ml<sup>-1</sup>) selective cytotoxicity in KB (ED<sub>50</sub> = 0.58  $\mu$ g ml<sup>-1</sup>) and RPMI (malignant melonoma) (ED<sub>50</sub> =  $0.61 \,\mu g \,ml^{-1}$ ) cells. Compound 1 was inactive against A-549 (lung), HCT-8 (colon) and TE671 (human medulloblastoma) tumour cells at 10 µg ml<sup>-1</sup>. The cytotoxicity assay was carried out according to literature methods.6,7

Amorphispironone  $1\ddagger \{C_{23}H_{22}O_7, \text{ colourless crystals from } \}$ 80% aqueous MeOH, m.p. 152–152.5 °C, [α]<sub>D</sub> –61.6° (c 0.162 in MeOH),  $\lambda_{max}$  nm (MeOH) (log  $\epsilon$ ) 269 (4.51) and 316 (4.05)} was isolated from the crude chloroform extract of Amorpha fruticosa by silica gel chromatography. Many rotenoids have been isolated from the fruit<sup>2,3</sup> and the root bark<sup>4</sup> of this same plant. The general features of the NMR spectra of 1 suggested that it also was a rotenoid. Comparison of the <sup>1</sup>H NMR spectral data for 1 with those of the known rotenoid deguelin  $2^5$  revealed that the C-D-E ring protons were similar but the A-B ring protons were different. These data suggested that 1 was a rotenoid, like 2 but differing in the A/B ring region. The complete structure and stereochemistry of amorphispironone were established unequivocally by single-crystal X-ray analysis.§¶ A view of the solid-state conformation is provided in Fig. 1. In general, bond lengths

‡ EI mass m/z: 410.1364 (M<sup>+</sup>). C<sub>23</sub>H<sub>22</sub>O<sub>7</sub> requires m/z 410.137. IR (KBr):  $v_{max}/cm^{-1}$  3010, 2970, 1665, 1640, 1625, 1575, 1393, 1270, 1110, 1070 and 820. NMR spectral assignments were made on the basis of <sup>1</sup>H-<sup>13</sup>C COSY and long range <sup>1</sup>H-<sup>13</sup>C COSY spectra. <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz, J in Hz) δ 5.02 (1H, s, 1-H), 5.52 (1H, s, 4-H), 4.50 (1H, dd, J 3 and 10, 6-H<sub>a</sub>), 4.64 (1H, d, J 10, 6-H<sub>b</sub>), 5.27 (1H, dd, J 3 and 4.5, 6a-H), 6.50 (1H, d, J 9, 10-H), 7.67 (1H, d, J 9, 11-H), 3.44 (1H, d, J 4.5, 12a-H), 3.25 (3H, s, 2-OMe), 3.84 (3H, s, 3-OMe), 6.67 (1H, d, J 10, 4'-H), 5.64 (1H, d, J 10, 5'-H) and 1.44 (3H, s, 7'-H or 8'-H), 1.47 (3H, s, 7'-H or 8'-H). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75 MHz) & 105.8 (d, C-1), 84.1 (s, C-1a), 148.3 (s, C-2), 166.2 (s, C-3), 99.8 (d, C-4), 198.8 (s, C-4a), 76.0 (t, C-6), 82.7 (d, C-6a), 160.1 (s, C-7a or C-9), 109.4 (s, C-8), 156.3 (s, C-7a or C-9), 111.7 (d, C-10), 127.6 (d, C-11), 114.5 (s, C-11a), 186.0 (s, C-12), 60.1 (d, C-12a), 55.1 (q, C-2-OMe), 55.7 (q, C-3-OMe), 115.6 (d, C-4'), 129.2 (d, C-5'), 77.7 (s, C-6'), 28.0 (q, C-7' or C-8') and 28.2 (q, C-7' or C-8').

§ Crystal data for 1:  $C_{23}H_{22}O_7$ , M = 410.43, monoclinic, space group 1038.6(4) Å<sup>3</sup>, Z = 2,  $D_c = 1.312$  g cm<sup>-3</sup>,  $\mu$ (Cu-K $\alpha$  radiation,  $\lambda =$  $1.5418 \text{ Å}) = 7.7 \text{ cm}^{-1}$ . Intensity data  $[\pm h, +k, +l; \theta_{\text{max}} = 75^{\circ}, \text{scan}]$ width  $(0.75 + 0.14 \tan \theta)^{\circ}$ ; 2279 non-equivalent reflections] were recorded on an Enraf-Nonius CAD-4 diffractometer (Cu-Kα radiation, graphite monochromator). The crystal structure was solved by direct methods (MULTAN11/82). Full-matrix least-squares refinement of atomic parameters (anisotropic C, O; isotropic H) converged at R = 0.032 ( $R_w = 0.047$ ) over 1975 reflections with  $I > 3.0\sigma(I)$ . Crystallographic calculations were performed by use of the Enraf-Nonius SDP package. Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See Notice to Authors, Issue No. 1.

¶ The absolute stereochemistry represented by structure 1 could not be established from the X-ray data. It follows, however, by assuming identical configurations at common asymmetric centres in 1 and another rotenoid derivative with which it co-occurs and for which the absolute stereochemistry has been established in our laboratory by an X-ray crystallographic analysis.



Scheme 1

agree well with expected values. Bond strain, however, is evident in the elongated C(1a)–C(12a) [1.572(3) Å] and C(1a)–C(4a) [1.548(4) Å] bonds. Ring A is fairly flat but puckered slightly towards a shallow envelope form with C(1a) as the out-of-plane atom, || rings B and C have envelope conformations with C(6a) as the out-of-plane atom in each case, ring D is planar, while ring E approximates to a 1,3-diplanar form.

The unusual spiro A-B ring system in 1 is probably biogenetically derived from 2 in the manner indicated in Scheme 1.

This investigation was supported by a grant from the US National Cancer Institute (K. H. L.).

Received, 15th July 1991; Com. 1/03590H

## References

- 1 Z. Rozsa, J. Hohmann, K. Szendrei, J. Reisch and I. Mester, Heterocycles, 1982, 19, 1793.
- 2 T. Somleva and U. Ognyanov, Planta Med., 1985, 51, 219.
- 3 L. A. Mitscher, A. Al-Shamma, T. Haas, P. B. Hudson and Y. H.
- Park, *Heterocycles*, 1979, 12, 1033.
  J. Hohmann, Z. Rozsa, J. Reisch, and K. Szendrei, *Herba Hung.*, 1982, 21, 179.
- 5 L. Crombie and J. W. Lown, J. Chem. Soc., 1962, 775; D. G. Carlson, D. Weisleder and W. H. Tallent, *Tetrahedron*, 1973, 29, 2731.
- 6 K. H. Lee, Y. M. Lin, T. S. Wu, D. C. Zhang, T. Yamagishi, T. Hayashi, I. H. Hall, J. J. Chang, R. Y. Wu and T. H. Yang, *Planta Med.*, 1988, 54, 308.
- 7 R. I. Geran, N. H. Greenberg, M. M. MacDonald, A. M. Schumacher and B. J. Abbott, *Cancer Chemother. Rep.*, 1972, 3, 1.