Biosynthesis of Vitamin B_{12} : Use of a Single ¹³C Label in the Macrocycle to Confirm C-11 Methylation in Precorrin-6x

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[11-¹³C]Uro'gen III is unambiguously synthesised for enzymic conversion into precorrin-6x which as its octamethyl ester gives a ¹³C NMR spectrum confirming the presence of a C-11 methyl group.

The direction of research on the biosynthesis of vitamin B_{12} has been sharply changed by the isolation of a new intermediate, precorrin- $6x^1$ whose octamethyl ester was assigned^{1,2} the structure **5b**. This leads to **5a**, used in this paper, as the structure of precorrin-6x itself but it is possible that the natural octa-acid has a tautomeric structure, *e.g.* with a 9,10 doublebond rather than 8,9, and that migration of the double-bond occurs during isolation and esterification. Precorrin-6x is biosynthesised from uro'gen III **3a** by many steps including six *C*-methylations. One of these introduced methyl groups was placed at C-11 and this was one of several unexpected features^{1,2} of structure **5a**: the C-11 methyl presumably migrates² to its final C-12 position at a later stage on the biosynthetic pathway to hydrogenobyrinic acid 6 and cobyrinic acid 7.

The structure **5b** was deduced^{1,2} from the results of many ³H, ¹⁴C and ¹³C labelling experiments using the early precursors 5-aminolaevulinic acid and *S*-adenosylmethionine (SAM). In particular, the positioning of a *C*-methyl group at C-11 depended on a set of three ¹³C labelling experiments² based on the biosynthetic incorporation into precorrin-6x of, respectively, [3-¹³C], [4-¹³C] and [5-¹³C]-5-aminolaevulinic acid together with [*methyl*-¹³C]SAM. The various NMR spectra of these samples of precorrin-6x ester showing one-bond ¹³C-¹³C couplings and 2-3 bond ¹H-¹³C couplings together with ¹H NOE-difference spectra gave an interlocking



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set of data supporting C-11 methylation. Nevertheless, when studying a molecule of this complexity and for a feature as unexpected as the C-11 methylation, it was important to provide confirmation by labelling only C-11 of the macrocycle. The results are now outlined.

The [5-13C]pyrrole³ 1 (90 atom % ¹³C) was built into [11-¹³C]uro'gen III **3b** via octamethyl [11-¹³C]uroporphyrin III 2 by an established synthesis⁴ as indicated in Scheme 1. Enzymic methylation of 3b using unlabelled SAM, SAM: uro'gen III methyltransferase5 and SAM : precorrin-2 methyltransferase⁶ afforded the trimethylated intermediate [11-¹³C]precorrin-3 4. The further biosynthetic steps through to precorrin-6x 5c were carried out by the protein preparation from Pseudomonas denitrificans including [methyl-13C]SAM but omitting the reducing cofactor NADPH; this omission¹ causes precorrin-6x 5c to accumulate.

It should be noted that by the foregoing procedure, the first three C-methyl groups introduced to form precorrin-3 4 are unlabelled and that two of these, at C-2 and C-7, carry forward into precorrin-6x 5c; that at C-20 is lost as acetic acid during the ring-contraction process.^{7,8} The next three C-methylations needed to reach precorrin-6x 5c insert ¹³C-labelled methyl groups.

The proton-decoupled ¹³C NMR spectrum of the resultant precorrin-6x octamethyl ester 5d showed two sharp singlets at δ 23.5 and 29.5 corresponding to the 17-methyl and 1-methyl groups respectively. Importantly, the third methyl signal was a doublet (J 37.9 Hz) at δ 21.5. This methyl group must be attached to C-11 since the macrocycle is only labelled at this site; the signal for C-11 itself was a doublet (J 38.2 Hz) at δ 71.1. All these chemical shifts and J values match exactly those

The C-11 methyl group of precorrin-6x 5a is thus confirmed and the structures of later intermediates9 which in part build on structure 5a also gain additional strength.

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References

- 1 D. Thibaut, L. Debussche and F. Blanche, Proc. Natl. Acad. Sci. USA, 1990, 87, 8795.
- 2 D. Thibaut, F. Blanche, L. Debussche, F. J. Leeper and A. R. Battersby, Proc. Natl. Acad. Sci. USA, 1990, 87, 8800.
- 3 A. R. Battersby, C. J. R. Fookes, M. J. Meegan, E. McDonald and H. K. W. Wurziger, J. Chem. Soc., Perkin Trans. 1, 1981, 2786.
- 4 A. R. Battersby, M. Ihara, E. McDonald, J. Saunders and R. J. Wells, J. Chem. Soc., Perkin Trans. 1, 1976, 283. 5 F. Blanche, L. Debussche, D. Thibaut, J. Crouzet and B.
- Cameron, J. Bacteriol., 1989, 171, 4222.
- 6 D. Thibaut, M. Couder, J. Crouzet, L. Debussche, B. Cameron and F. Blanche, J. Bacteriol., 1990, 172, 6245.
- 7 L. Mombelli, C. Nussbaumer, H. Weber, G. Müller and D. Arigoni, Proc. Natl. Acad. Sci. USA, 1981, 78, 11.
- A. R. Battersby, M. J. Bushell, C. Jones, N. G. Lewis and A. Pfenninger, Proc. Natl. Acad. Sci. USA, 1981, 78, 13.
- 9 E.g. D. Thibaut, F. Kiuchi, L. Debussche, F. J. Leeper, F. Blanche and A. R. Battersby, J. Chem. Soc., Chem. Commun., 1992, following communication.