

Direct Observation of Push Effect on the Heterolytic O–O Bond Cleavage Reaction of Acylperoxy-Iron(III) Porphyrin Adducts

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We report the first direct observation of the push effect of the sixth ligand on the heterolytic O–O bond cleavage step of acylperoxy-iron(III) porphyrin complexes to the corresponding oxo-ferryl porphyrin π -cation radicals by utilizing a series of imidazole derivatives.

The elucidation of the O–O bond cleavage process of hydroperoxy-iron(III) porphyrin complexes yielding active high-valent oxo species is very important to understand the nature of the oxygen activation mechanisms by haem enzymes such as cytochrome P-450, peroxidases and catalase.¹ It has been proposed that the proximal ligands of these haem enzymes such as thiolate, imidazolate and phenolate serve as strong internal electron donors to destabilize the O–O bond of hydroperoxy-iron(III) porphyrin (*push effect*),² and distal base residues are considered to favour the heterolytic O–O bond cleavage (*pull effect*).³ The push–pull mechanism has been examined by using synthetic haem model systems.^{4–6} On the other hand, we have reported that the reaction of perbenzoic acid and Fe^{III} porphyrins bearing a hydroxo ligand **1** [Fe(OH)] in dry CH₂Cl₂ at low temperature gives peracid-Fe^{III} adducts as an irreversible process followed by a heterolytic cleavage of the O–O bond of the adducts to form the corresponding oxo-ferryl porphyrin cation radicals (Scheme 1).^{6a–c,g,7} These stepwise reactions are directly observable by employing low-temperature UV–VIS measurement. Thus, the rate constants (k_{dec}) of the O–O bond cleavage of five coordinated acylper-

oxo-Fe^{III} porphyrins **2** in CH₂Cl₂ at low temperature have been reported.^{6c,g}

Here, we have examined the stoichiometric reaction of imidazole derivatives and **5** in a CH₂Cl₂ solution at low temperature in order to understand the push effect of the sixth ligand and found the immediate formation of peracid-Fe^{III}-imidazole ternary complexes **4** followed by the heterolytic O–O bond cleavage of **4** to yield oxo-ferryl porphyrin π -cation radicals.

The push effect of the sixth ligand on the heterolysis of the O–O bond: In a typical reaction, a methylene chloride solution of hydroxo-iron(III) TDMPP⁸ (**1**; 2.0×10^{-5} mol dm⁻³) was cooled to -80°C in a UV–VIS cuvette. Introduction of 1.2 equiv. of *m*-chloroperbenzoic acid (MCPBA) to the solution immediately produced acylperoxy-iron(III) porphyrin **2** followed by isosbestic spectral changes affording an oxo-ferryl porphyrin cation radical **3** according to heterolytic O–O bond cleavage as reported before (Fig. 1, inset).^{6c,g,9} In contrast, addition of 1 equiv. of 1-phenylimidazole (1-Ph-Im) after the formation of **2** was found to give a new intermediate which exhibits a UV–VIS spectrum typical of six coordinated ferric low-spin complex (**4** in Fig. 1).⁹

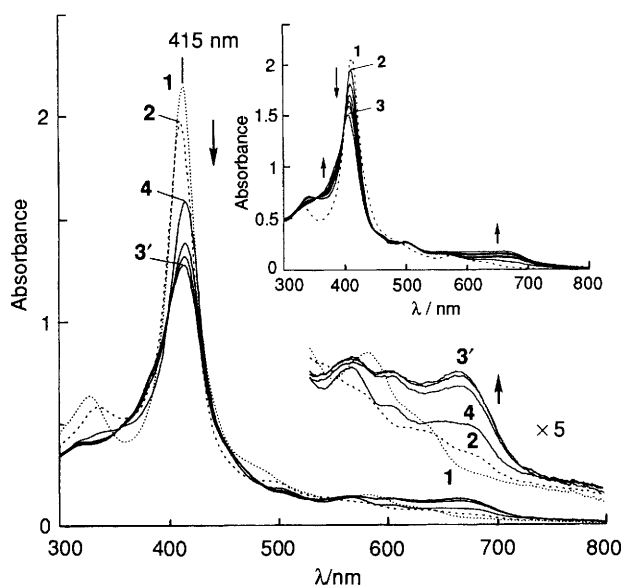
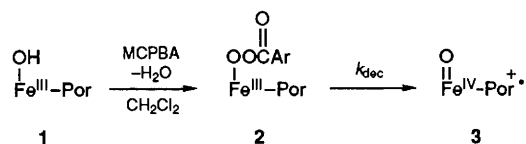


Fig. 1 Effect of 1-phenylimidazole (1 equiv.) on the reaction of 2.4×10^{-5} mol dm⁻³ of hydroxoiron(III) porphyrin (**1**; \cdots) and 1.2 equiv. of MCPBA in methylene chloride at -80°C . The spectrum of **2** ($---$) was recorded immediately after the addition of MCPBA. The spectrum of **4** ($—$) was recorded immediately after the addition of 1-phenylimidazole. The transformation of **4** was recorded every 5 min. Inset: time dependent spectral changes in the reaction of Fe^{III}TDMPP(OH) and MCPBA in methylene chloride at -80°C .

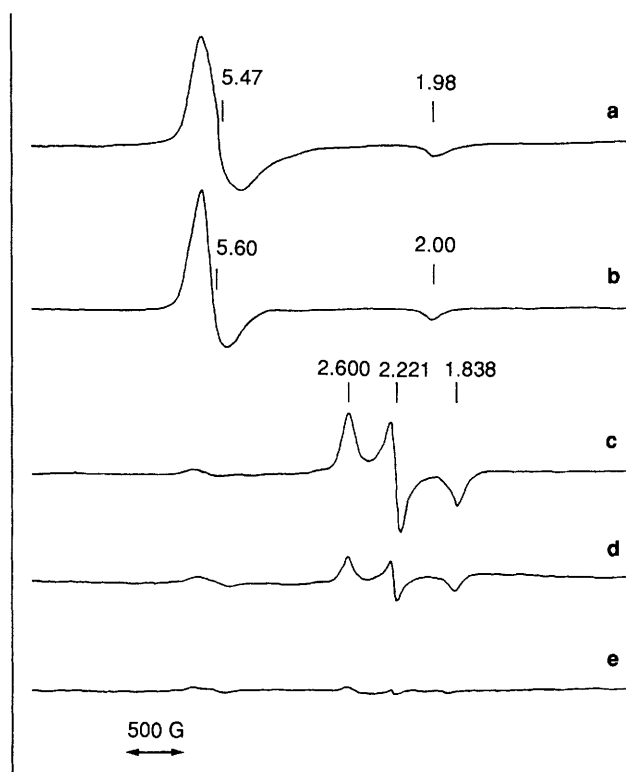


Fig. 2 Time-dependent EPR spectral changes in the stoichiometric reaction of **1** and MCPBA (8.0×10^{-3} mol dm⁻³) in methylene chloride at -78°C . Each sample was frozen (77 K) for measurement, **a**: a methylene chloride solution of **1**, **b**: sample immediately after the addition of MCPBA to the solution of **a**, **c**: sample immediately after the introduction of 1 equiv. of 1-phenylimidazole to the solution of **b**, **d** and **e**: time-dependent EPR spectral changes of **c**.

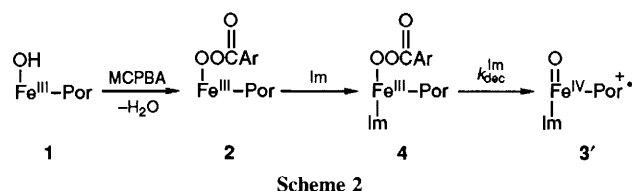


Table 1 Effect of the sixth ligand on the first-order rate constants for the O–O bond cleavage of **2** and **4**

Imidazole	$k_{\text{hetero}}^{(\text{Im})}/10^{-3}\text{s}^{-1}$
Without Im	0.3 ± 0.2
1-Me-5-Cl-Im	0.6 ± 0.2
1-Ph-Im	2.0 ± 0.1
1-Me-Im	16.6 ± 0.6

^a 1.2 equiv. of *m*-chloroperbenzoic acid was used in CH₂Cl₂ at –80 °C.

Further, conversion of **4** into the oxo-ferryl cation radical **3'** is evident from isosbestic changes of UV–VIS spectra. On the basis of stoichiometric formation, spectral features and reactivity of **4**, the structure of **4** is assigned to be a six coordinated low-spin acylperoxo-Fe^{III}TDMPP(1-Ph-Im). Further evidence for the stoichiometric formation of the acylperoxo-iron(III)-imidazole adduct was obtained from the electron paramagnetic resonance (EPR) spectrum of Fe^{III}TDMPP(MCPBA)-(1-PhIm) at 77 K. Upon the introduction of MCPBA at –78 °C, a small EPR spectral change due to the replacement of the fifth ligand was observed (Fig. 2, **a** → **b**). Addition of 1-Ph-Im to the resulting solution gave a EPR spectrum characteristic of six coordinated ferric low-spin complexes (**b** → **c**).¹⁰ Finally, EPR signals disappeared according to the formation of **3'** [(**c** → **d** → **e**).¹¹ Replacement of 1-Ph-Im by either 1-methyl-5-chloroimidazole or 1-methylimidazole in the reaction with **2** also afforded **3'** via instantaneous formation of six coordinated low spin intermediate **4**.

On the basis of the UV–VIS spectral changes shown in Fig. 1, the O–O bond cleavage of **4** was found to be first order in [**4**], consistent with the reaction mechanism shown in Scheme 2. Values of the rate constants $k_{\text{hetero}}^{(\text{Im})}$ for the formation of **3'** in the presence and absence of a series of imidazole derivatives are summarized in Table 1. Apparently, electron-rich imidazoles accelerate the heterolytic O–O bond-cleavage step, whereas electron-poor imidazole is less effective. These observations of the push effect mediated by *N*-substituted imidazoles provide the first direct evidence to support the proposed role of strong electron donor, anionic imidazole of proximal histidine, for the formation of Compound I in the peroxidase reactions.² The results are consistent with the push effect of substituents at the *meso*-positions of a porphyrin ring on the heterolysis of the O–O bond.^{6g}

In order to determine the activation parameters for the O–O bond cleavage, the formation of **3'** in the presence and absence of 1-Ph-Im was examined between –40 and –90 °C. Linear Arrhenius plots were obtained based on the results listed in Table 2. The activation energies for the formation of **3'** in the presence and absence of 1-Ph-Im were determined to be 3.9 and 5.9 kcal mol^{–1} (1 cal = 4.184 J), respectively. The result suggests that the coordination of a sixth ligand stabilizes the transition state in the O–O bond-cleavage reaction.

In conclusion, it has been shown that the electron-rich sixth ligands strongly enhance the heterolysis of the O–O bond of acylperoxo-iron(III) TDMPP-imidazole complexes to yield the corresponding oxo-ferryl porphyrin cation radicals.

Table 2. Temperature dependence of the first-order rate constants for the O–O bond cleavage of acylperoxo-Fe^{III}TDMPP and acylperoxo-Fe^{III}TDMPP(1-Ph-Im)

T/°C	$k_{\text{hetero}}^{(\text{Im})}/10^{-3}\text{s}^{-1}$	
	Without Im	1-Ph-Im
–40	3.9 ± 0.2	11.2 ± 0.5
–50	2.4 ± 0.1	8.4 ± 0.4
–60	1.3 ± 0.1	5.5 ± 0.2
–70	0.6 ± 0.2	3.4 ± 0.1
–80	0.3 ± 0.2	2.0 ± 0.1
–90	<i>b</i>	1.2 ± 0.1

^a 1.2 equiv. of *m*-chloroperbenzoic acid was used in CH₂Cl₂. ^b Not available.

References

- For example: R. E. White and M. J. Coon, *Ann. Rev. Biochem.*, 1980, **50**, 315; P. R. Ortiz de Montellano, *Oxygen activation and transfer*, ed. P. R. Ortiz de Montellano, Plenum Press, New York, 1986, pp. 217–271; J. H. Dawson, *Science*, 1988, **240**, 433.
- T. L. Poulos, B. C. Finzel, I. C. Gunsalus, G. C. Wagner and J. Kraut, *J. Biol. Chem.*, 1985, **260**, 16 122; T. L. Poulos, B. C. Finzel and A. J. Howard, *Biochemistry*, 1986, **25**, 5314; T. L. Poulos and A. J. Howard, *Biochemistry*, 1987, **26**, 8165; V. Thanabal, J. S. de Ropp and G. N. La Mar, *J. Am. Chem. Soc.*, 1988, **110**, 3027; M. R. N. Murthy, T. J. Reid III, A. Sicignano, N. Tanaka and M. G. Rossmann, *J. Mol. Biol.*, 1981, **152**, 465.
- J. H. Dawson, R. H. Holm, J. R. Trudell, G. Barth, R. E. Linder, E. Bunnenberg, C. Djerassi and S. C. Tang, *J. Am. Chem. Soc.*, 1976, **98**, 3707; T. L. Poulos, *Adv. Inorg. Biochem.*, 1987, **7**, 1.
- T. G. Traylor, W. A. Lee and D. V. Stynes, *J. Am. Chem. Soc.*, 1984, **106**, 755; T. G. Traylor and R. Popovitz-Biro, *J. Am. Chem. Soc.*, 1988, **110**, 239; T. G. Traylor and F. Xu, *J. Am. Chem. Soc.*, 1990, **112**, 178.
- W. A. Lee and T. C. Bruice, *J. Am. Chem. Soc.*, 1985, **107**, 513; M. Z. Zippies, W. A. Lee and T. C. Bruice, *J. Am. Chem. Soc.*, 1986, **108**, 4433; L. C. Yuan and T. C. Bruice, *J. Am. Chem. Soc.*, 1986, **108**, 1643; T. C. Bruice, P. N. Balasubramanian, R. W. Lee and J. R. Lindsay Smith, *J. Am. Chem. Soc.*, 1988, **110**, 7890; R. Panicucci and T. C. Bruice, *J. Am. Chem. Soc.*, 1990, **112**, 6063.
- (a) J. T. Groves and Y. Watanabe, *Inorg. Chem.*, 1986, **25**, 4808; (b) J. T. Groves and Y. Watanabe, *J. Am. Chem. Soc.*, 1986, **108**, 7834 and 7836; (c) J. T. Groves and Y. Watanabe, *J. Am. Chem. Soc.*, 1988, **110**, 8443; (d) R. Labeque and L. J. Marnett, *J. Am. Chem. Soc.*, 1989, **111**, 6621; (e) T. Higuchi, S. Uzu and M. Hirobe, *J. Am. Chem. Soc.*, 1990, **112**, 7051; (f) A. Robert, B. Loock, M. Momenteau and B. Meunier, *Inorg. Chem.*, 1991, **30**, 706; (g) K. Yamaguchi, Y. Watanabe and I. Morishima, *Inorg. Chem.*, 1992, **31**, 156.
- J. T. Groves and Y. Watanabe, *Inorg. Chem.*, 1987, **26**, 785.
- TDMPP is 5,10,15,20-tetrakis(2,6-dimethylphenyl)porphyrin and hydroxoiron(III) TDMPP was characterized by Bruice *et al.*: T. C. Woon, A. Shirazi and T. C. Bruice, *Inorg. Chem.*, 1986, **25**, 3845.
- The absorption maxima for **4** was found at 416, 567 and 602 nm typical for the ferric low-spin porphyrin adducts. (a) E. W. Ainscough, A. W. Addison, D. Dolphin and B. R. James, *J. Am. Chem. Soc.*, 1978, **100**, 7585; (b) R. Quinn, M. Nappa and J. S. Valentine, *J. Am. Chem. Soc.*, 1982, **104**, 2588; (c) K. Tajima, J. Jinno, K. Ishizu, H. Sakurai and H. Ohya-Nishiguchi, *Inorg. Chem.*, 1989, **28**, 709; (d) K. Tajima, *Inorg. Chim. Acta*, 1990, **169**, 211.
- These values are similar to PPIXDBFe(OAr)(1-MeIm)^{9a} (PPIXDBE = protoporphyrin IX di-*tert*-butyl ester) but different from TPPFe(Im)(OOBu) reported by Tajima *et al.*^{9c,d} and TPPFe(ImH)₂Cl.^{9b}
- J. T. Groves, R. C. Haushalter, M. Nakamura, T. E. Nemo and B. J. Evans, *J. Am. Chem. Soc.*, 1981, **103**, 2884.