## Stereochemical Course of Ring Formation in Fumitremorgin B and Verruculogen, Metabolites of *Penicillium verruculosum:* Investigation into the Loss of Stereochemical Integrity of the Geminal Methyl Groups

## Robert Vleggaar,\* a R. Marthinus Horak b and Vinesh J. Maharaj b

- <sup>a</sup> Department of Chemistry, University of Pretoria, Pretoria 0002, South Africa
- b Division of Food Science and Technology, CSIR, PO Box 395, Pretoria 0001, South Africa

Incorporation studies of [2-2H<sub>3</sub>,2-13C] acetate and different (2H,13C)-labelled mevalonolactones into verruculogen established the stereochemical course of ring C formation in fumitremorgin B, which results in the loss of stereochemical integrity of the C-22 methyl groups, and of the formation of the eight-membered peroxide ring at C-25 in verruculogen.

An earlier incorporation study in which (3RS)-[2-13C]mevalonolactone was administered to cultures of Penicillium verruculosum has established that although C-23 and C-29 in verruculogen 1 are labelled to the same extent, a lower, but significant enrichment is also observed for C-24.1 A similar phenomenon was observed for fumitremorgin B 2 obtained from the same feeding experiment. As expected (3RS)-[2-13C]mevalonolactone labelled the 27-pro-E methyl group, C-29, of fumitremorgin B but it is C-23, the 22-pro-Z methyl group of the 2,2-dimethylvinyl moiety, which is enriched. In addition an enrichment is also observed for the 22-pro-E methyl group, C-24.1 This finding is confirmed in the present study by the one-bond (13C-13C) couplings observed in the <sup>13</sup>C{<sup>1</sup>H} NMR spectra of verruculogen 1 and fumitremorgin B 2 derived from (3RS)- $[2,3^{-13}C_2]$  mevalonolactone: the enhancement of the C-24 signal was ca. 25% of that of the C-23 and C-29 signals. It is evident that the stereochemical integrity of the C-22 diastereotopic methyl groups in verruculogen 1 is lost during the formation of fumitremorgin B 2. The focus of interest in this paper is the stereochemical course of this process.

The most likely explanation of this result is that an intermediate, which allows rotation around a C(21)–C(22) single bond, is involved in the formation of ring C in fumitremorgin B. In order to test this hypothesis the fate of the hydrogen atoms in the biosynthesis of verruculogen was studied using  $^2H$  in association with  $^{13}C$  as a reporter nucleus and either the  $\alpha^{-2.3}$  or  $\beta$ -isotope shifts $^{3.4}$  in the  $^{13}C$  NMR spectra. Verruculogen was used in these studies as the stereochemical course of the change in hybridisation of the C-22 centre in going from fumitremorgin B to verruculogen is known.

The incorporation of different (2H,13C)-labelled mevalonolactones† into verruculogen established the origin and fate of the C-24 hydrogen atoms. The  $\beta$ -isotope shifts of -0.068and -0.132 ppm observed for C-27 and C-22, respectively, in the <sup>13</sup>C{<sup>1</sup>H} NMR spectrum of verruculogen derived from (3RS)- $[6-2H_3,3-13C]$ mevalonolactone (>98 atom% <sup>2</sup>H, 99 atom % <sup>13</sup>C) indicate that three deuterium atoms are retained at both C-28 and C-24 in verruculogen. The presence of three deuterium atoms at C-28 is based on the established mechanism for the transformation of mevalonolactone to 3,3dimethylallyl pyrophosphate as well as the β-shift value observed for a similar system in viridicatumtoxin.<sup>5</sup> The single β-shifted <sup>13</sup>C signal for C-22 excludes a mechanism involving a C-24 sp<sup>2</sup>-hybridised intermediate. This finding was corroborated by the observed  $\alpha$ -isotope shifts in the <sup>13</sup>C{<sup>2</sup>H, <sup>1</sup>H} NMR spectrum of verruculogen on incorporation of (3RS)-[6-2H<sub>3</sub>,6-13C]mevalonolactone (99 atom% <sup>13</sup>C) containing <sup>13</sup>C,<sup>2</sup>H<sub>3</sub>-(55 mol%) and <sup>13</sup>C,<sup>2</sup>H<sub>2</sub>-labelled (45 mol%) species at C-6. The relative intensity of the two  $\alpha$ -isotopically shifted signals observed for the C-24 resonance and the magnitude of the  $\alpha$ -isotope shifts ( $\Delta\delta$  -0.55 and -0.83 ppm) in the major

isotopomer were essentially the same as those for the C-28 resonance ( $\Delta\delta$  -0.53 and -0.79 ppm), which served as an internal reference as no deuterium loss occurs from this position during the biosynthesis. Similar  $\alpha$ -isotopically shifted signals, but with greatly reduced intensity, were evident for the C-23 resonance and suggest that a C-23 sp<sup>2</sup>-hybridised intermediate can be excluded. This was confirmed by the  $\alpha$ -isotope shifts observed in the  ${}^{13}C\{{}^{2}H,{}^{1}H\}$  NMR spectrum of verruculogen derived from sodium [2-2H<sub>3</sub>,2-13C]acetate. The observed  $\alpha$ -isotopically shifted signals (-0.27 ppm per <sup>2</sup>H atom) confirm the expected presence of three and two deuterium atoms at C-28 and C-29, respectively. The intensity and number of the  $\alpha$ -shifted signals observed for C-23 (two <sup>2</sup>H atoms) and C-24 (three <sup>2</sup>H atoms), on comparison with the C-28 and C-29 signals, confirms that none of the C-23 or C-24 hydrogen atoms is lost in the process which results in the loss of the stereochemical integrity of the C-22 methyl groups.

A possible mechanism for this process, which must occur during the formation of ring C in fumitremorgin B, is as follows. The introduction of the 3,3-dimethylallyl moiety at N-1 and C-2 of the indole nucleus in the putative precursor 3 occurs with inversion of configuration. The loss of one of the C-20 diastereotopic methylene protons generates an allylic carbocation 4 which allows rotation around the C(20)–C(21) bond and concomitant loss of the stereochemical integrity of the C-22 methyl groups. Attack of N-19 on the 20Si face of the allylic carbocation generates the correct stereochemistry at C-20 in fumitremorgin B. The process could proceed by either an ionic or radical mechanism.

 $<sup>\</sup>dagger$  The details on the synthesis of the (<sup>2</sup>H, <sup>13</sup>C)-labelled mevalonolactones used in this study will be reported in full elsewhere.

Earlier studies on the incorporation of (3RS)- $[5-2H_2]$ mevalonolactone into verruculogen established that a single deuterium atom is retained at both C-20 and C-25.1 The question as to which of the two diastereotopic C-5 protons of mevalonate is retained in each case, is answered in the present study by incorporation of stereospecifically labelled (3RS,5S)and (3RS,5R)-[5-2H,4-13C]mevalonolactone into verruculogen. The retention of deuterium at C-20 in verruculogen derived from the 5S stereoisomer was evident from the  $\beta$ -isotope shift of -0.094 ppm observed for the C-21 resonance in the  $^{13}C\{^{1}H\}$  NMR spectrum. No  $\beta$ -shifted signal was observed for the C-26 resonance. This pattern was reversed in the spectrum of verruculogen derived from the 5R stereoisomer: a  $\beta$ -shifted signal ( $\Delta\delta$  -0.077 ppm) was observed for the C-26 resonance but none for the C-21 resonance. The results establish that the 5Si proton of mevalonate is retained at C-20 of verruculogen whereas the 5Re proton is retained at C-25.

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## References

- 1 R. M. Horak and R. Vleggaar, J. Chem. Soc., Chem. Commun., 1987, 1568.
- M. J. Garson and J. Staunton, *Chem. Soc. Rev.*, 1979, 539; M. J. Garson, R. A. Hill and J. Staunton, *J. Chem. Soc.*, *Chem. Commun.*, 1977, 921.
- 3 P. E. Hansen, Annu. Rep. N.M.R. Spectrosc., 1983, 15, 105.
- 4 C. Abell and J. Staunton, J. Chem. Soc., Chem. Commun., 1981, 856
- 5 R. M. Horak, V. J. Maharaj, S. F. Marais, F. R. van Heerden and R. Vleggaar, J. Chem. Soc., Chem. Commun., 1988, 1562.