

Enantiospecific Synthesis of the Fluoro and Epimeric Derivatives of 5'-Noraristeromycin

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The synthesis of derivatives of 5'-noraristeromycin **1** in which its C-4' hydroxy group has been (i) replaced by a fluorine atom (**5**) and (ii) inverted (**6**) is described starting from the diacetate of (*Z*)-cyclopentene-3,5-diol.

Inhibition of (*S*)-adenosyl-L-homocysteine (AdoHcy) hydrolase by derivatives of adenosine (e.g. carbocyclic adenosine or aristeromycin, **1**) has been a productive area in the pursuit of antiviral agents.¹ A common problem with these compounds that limits their potential usefulness, however, is the associated toxicity arising from conversion to the 5'-phosphates.^{2,3} To circumvent this undesirable consequence recent efforts have focused on derivatives of **1** either less likely to undergo phosphorylation (e.g. **2**)⁴ or incapable of doing so (e.g. **3**⁵ and **4**⁶). Both **2** and **3** are promising antiviral candidates that inhibit AdoHcy hydrolase while showing little or no toxicity.

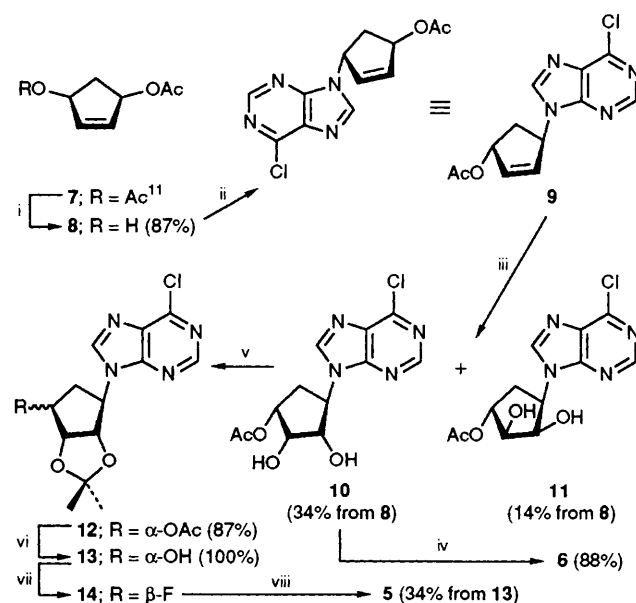
In exploring the lead provided by **2** and **3**, the fluoro derivative **5** arose as a meaningful target compound. Compound **5** is related (i) to **2** owing to the similarity of the fluorine atom to the hydroxy group in size, electronegativity and ability to participate in hydrogen bonding,⁷ and (ii) to **3** because structural modifications in which a hydrogen is replaced by a fluorine atom have found relevance in the production of biologically useful compounds⁸ owing to similarities in the van der Waals radii of hydrogen and fluorine but dramatic differences in their electronegativities.⁹ The synthetic approach to **5** was designed also to avail a means to the epimer of **2** (that is, **6**) to provide a molecule that could be used to ascertain the biological consequences of C-4' hydroxy inversion. There is little literature precedence^{6a,10} to predict how significant the configuration at the C-4' centre of carbocyclic adenosines (e.g. **1**) is to the inhibition of AdoHcy hydrolase.

The synthesis of both **5** and **6** (Scheme 1) began with subjecting the diacetate of (*Z*)-cyclopentene-3,5-diol **7**¹¹ to hydrolysis with *Pseudomonas cepacia* lipase (PCL)¹² under carefully monitored conditions (pH) to assure monohydrolysis.^{6b,13} The resultant monoacetate (+)-**8**[†] underwent the Mitsunobu reaction¹⁵ with 6-chloropurine to yield **9**. Standard glycolization conditions on **9** led to a mixture of the desired **10**^{‡§} and **11**^{‡§} in approximately a 2:1 ratio. Following separation of this mixture using flash column chromatography, **10** and **11** were distinguished by a 1D nuclear

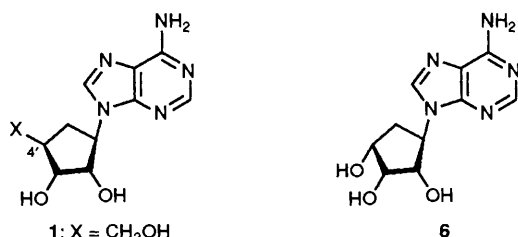
Overhauser enhancement determination. In this regard, pre-irradiation of H-2' in **10** resulted in enhancement of the H-3', H-4' and H-5'_a protons; no enhancement was observed for H-1' and H-5'_b. This confirmed the *cis*-relationship between the hydroxy groups and the acetate.

Ammonolysis of **10** provided **6**^{‡¶} whereas isopropylideneation (to **12**) followed by hydrolysis resulted in **13**.[§] Using diethylaminosulfur trifluoride (DAST),¹⁶ **13** was converted into **14**. Ammonolysis of **14** with subsequent deprotection yielded the target compound **5**.^{‡||}

The biological properties of **5** and **6** will be reported as they become available.



Scheme 1 Reagents and conditions: i, *Pseudomonas cepacia* lipase, 0.1 mol dm⁻³ phosphate buffer, pH 7, 25 °C; ii, 6-chloropurine-PPh₃-diethyl azodicarboxylate in tetrahydrofuran (THF), room temp.; iii, OsO₄-*N*-methylmorpholine *N*-oxide in THF-H₂O, room temp.; iv, NH₃ in MeOH, 100 °C; v, 2,2-dimethoxypropane-catalytic *p*-Me-C₆H₄SO₃H in acetone, room temp.; vi, K₂CO₃ in aq. MeOH, room temp.; vii, DAST in CH₂Cl₂, -78 °C to room temp.; viii, (a) NH₃ in MeOH, 100 °C; (b) dil. HCl.



- 1: X = CH₂OH
2: X = OH
3: X = H
4: X = Me
5: X = F

6

† [α]_D²⁵ + 67.30 (c 0.27, CHCl₃) {lit.^{14a} [α]_D²³ + 66.30 (c 1.53, CHCl₃), enantiomeric excess 99%; lit.^{14b} [α]_D²⁰ + 65.60 (c 2.3, CHCl₃ + 1% EtOH)}.

‡ Satisfactory microanalytical data was obtained for this compound.

§ Satisfactory ¹H and ¹³C NMR spectral data were recorded for this compound.

¶ Data for **6**: pale-yellow solid; m.p. 220 °C (decomp.); [α]_D²⁵ - 43.7 (c 1.0, dimethylformamide); ¹H NMR [(CD₃)₂SO] δ 1.90-1.94 (m, 1 H, H-5'), 2.50-2.75 (m, 1 H, H-5'), 3.20 (br, 1 H, OH), 3.45 (m, 1 H, H-1'), 3.86 (m, 1 H, H-4'), 4.15 (m, 2 H, H-2' and H-3'), 5.09 (m, 2 H, OH), 7.19 (s, 2 H, NH₂), 8.13 (s, 1 H, H-2), 8.18 (s, 1 H, H-8); ¹³C NMR [(CD₃)₂SO] δ 39.50, 52.77, 71.62, 73.52, 78.72, 140.59, 141.02, 151.37, 151.86, 155.70. The spectral data for this compound is different from **2**⁴ assuring that no inversion of configuration had occurred in the conversion of **10** into **6**.

|| Data for **5**: white solid; m.p. 128-130 °C; [α]_D²⁵ - 40.38 (c 1.0, MeOH); ¹H NMR [(CD₃)₂SO] δ 2.10-2.86 (m, 2 H, H-5'), 3.40 (br, 2 H, OH), 4.11 (m, 1 H, H-1'), 4.60 (m, 1 H, H-4'), 5.03-5.62 (m, 2 H, H-2' and H-3'), 7.22 (s, 2 H, NH₂), 8.16 (s, 1 H, H-2), 8.18 (s, 1 H, H-8); ¹³C NMR [(CD₃)₂SO] δ 33.56 (d, J_{CCF} 21.97 Hz), 52.66, 74.01, 74.49 (d, J_{CCF} 24.41 Hz), 95.40 (d, J_{CF} 178.23 Hz), 140.48, 140.69, 150.08, 152.45, 156.41.

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References

- 1 E. De Clercq, *Biochem. Pharmacol.*, 1987, **36**, 2567; M. Cools and E. De Clercq, *Biochem. Pharmacol.*, 1989, **38**, 1061.
 - 2 D. L. Hill, S. Straight, P. W. Allan and L. L. Bennett, Jr., *Mol. Pharmacol.*, 1971, **7**, 375; L. L. Bennett, Jr., R. W. Brockman, L. M. Rose, P. W. Allan, S. C. Shaddix, Y. F. Shealy and J. D. Clayton, *Mol. Pharmacol.*, 1985, **27**, 666.
 - 3 L. L. Bennett, Jr., P. W. Allan and D. L. Hill, *Mol. Pharmacol.*, 1968, **4**, 208.
 - 4 S. D. Patil, S. W. Schneller, M. Hosoya, R. Snoeck, G. Andrei, J. Balzarini and E. De Clercq, *J. Med. Chem.*, 1992, **35**, 3372.
 - 5 M. S. Wolfe and R. T. Borchardt, *J. Med. Chem.*, 1991, **34**, 1521.
 - 6 (a) M. S. Wolfe, Y. Lee, W. J. Bartlett, D. R. Borchardt and R. T. Borchardt, *J. Med. Chem.*, 1992, **35**, 1782; (b) S. M. Siddiqi, S. W. Schneller, S. Ikeda, R. Snoeck, G. Andrei, J. Balzarini and E. De Clercq, *Nucleosides Nucleotides*, 1993, **12**, 185.
 - 7 P. J. Card, *J. Carbohyd. Chem.*, 1985, **4**, 451; J. Mann, *Chem. Soc. Rev.*, 1987, **16**, 381; T. Tsuchiya, *Adv. Carbohyd. Chem. Biochem.*, 1990, **48**, 91.
 - 8 J. T. Welch, *Tetrahedron*, 1987, **43**, 3123.
 - 9 M. Schlosser, *Tetrahedron*, 1978, **34**, 3.
 - 10 J. C. Yeh, R. T. Borchardt and A. Vedani, *J. Comput.-Aided Mol. Des.*, 1991, **5**, 213.
 - 11 D. R. Deardorff, D. C. Myles and K. D. MacFerrin, *Tetrahedron Lett.*, 1985, **26**, 5615.
 - 12 X. Chen, S. M. Siddiqi and S. W. Schneller, *Tetrahedron Lett.*, 1992, **33**, 2249.
 - 13 S. M. Siddiqi, X. Chen and S. W. Schneller, *Nucleosides Nucleotides*, in the press.
 - 14 (a) D. R. Deardorff, A. J., Matthews, D. S. McMeekin and C. L. Craney, *Tetrahedron Lett.*, 1986, **27**, 1255; (b) K. Laumen and M. P. Schneider, *J. Chem. Soc., Chem. Commun.*, 1986, 1298.
 - 15 O. Mitsunobu, *Synthesis*, 1981, 1; M. Iwakawa, B. M. Pinto and W. A. Szarek, *Can. J. Chem.*, 1978, **56**, 326; W. A. Szarek, C. Depew, H. C. Jarrell and J. K. N. Jones, *J. Chem. Soc., Chem. Commun.*, 1975, 648; T. F. Jenny, N. Previsani and S. A. Benner, *Tetrahedron Lett.*, 1991, **32**, 7029; T. F. Jenny, K. C. Schneider and S. A. Benner, *Nucleosides Nucleotides*, 1992, **11**, 1257; Y. Wu and P. M. Woster, *J. Med. Chem.*, 1992, **35**, 3196.
 - 16 K. Biggadike, A. D. Borthwick, D. Evans, A. M. Exall, B. E. Kirk, S. M. Roberts, L. Stephenson and P. Youds, *J. Chem. Soc., Perkin Trans. 1*, 1988, 549.
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