

Preparation of Activated Nucleotides and their Phosphorothioate and Phosphorodithioate Analogues

Colin B. Reese,* Louise H. K. Shek and Zhengyun Zhao

Department of Chemistry, King's College London, Strand, London, UK WC2R 2LS

The preparation of nucleoside phosphoro-(1,2,4-triazolides) (e.g. **9a**) and their thio- and dithio- analogues (e.g. **9b** and **c**, respectively), and a corresponding set of activated nucleotides (e.g. **10a**, **b**, **c**) derived from 1-hydroxybenzotriazole is described; the reactions between these derivatives and methylamine are also reported.

Ribonucleoside 5'-phosphorimidazolides **1a** were first prepared by Cramer and his coworkers¹ and were shown^{2,3} to behave as activated nucleotides in that they reacted with primary amines, alcohols and monoalkyl phosphates to give the corresponding phosphoramidates, phosphodiester and *P*¹,*P*²-dialkyl pyrophosphates, respectively. Hoard and Ott subsequently demonstrated⁴ that the reaction between 2'-deoxyribonucleoside 5'-phosphorimidazolides **1b** and inorganic pyrophosphate constituted a convenient method for the preparation of 2'-deoxyribonucleoside 5'-triphosphates **3a**. This procedure was extended⁵ to include the conversion of nucleoside 5'-thiophosphorimidazolides **2a** and **2b** into the corresponding nucleoside 5'- α -thiotriphosphates **3b** (R = OH and H).

Scheit⁶ showed that 1,1',1''-phosphinylidynetris-1*H*-imidazole **4a** was an effective reagent for the conversion of suitably-protected nucleoside derivatives into nucleoside 5'-phosphorimidazolides **1**, and Eckstein and Gindl⁵ found that the corresponding thio-compound **4b** could be used in the same way in the preparation of 5'-thiophosphorimidazolides **2**. We now report that related phosphorylating agents† derived from 1*H*-1,2,4-triazole (i.e. **5a** and **b**) and 1-hydroxybenzotriazole (i.e. **6a** and **b**) can readily be used in the preparation of activated nucleotides and their phosphorothioate and phosphorodithioate analogues.

Two substrates (**7** and **8**)⁷ were used in our studies. When the thymidine derivative **7** was treated (Table 1, entry 1) with a threefold excess of **5a**⁸⁻¹⁰ (Table 1, footnote *a*) in THF

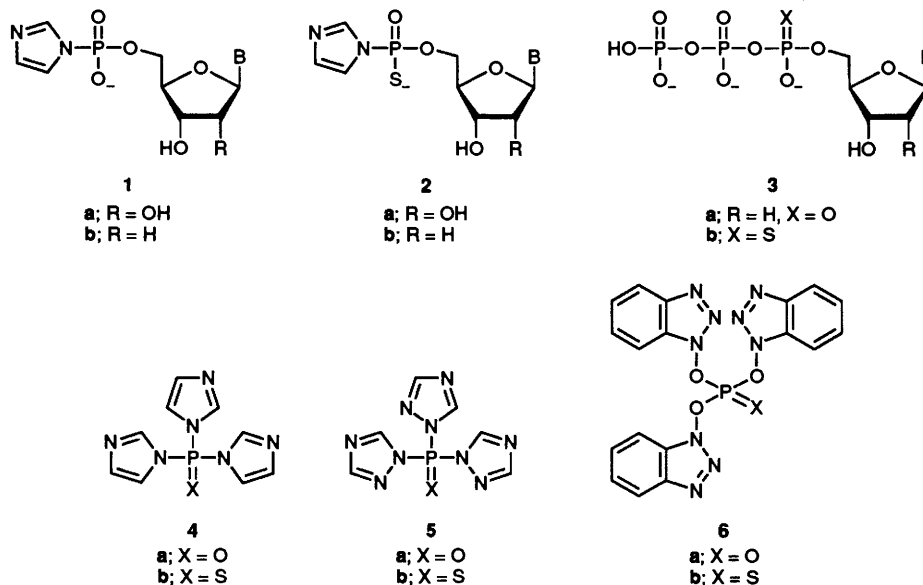
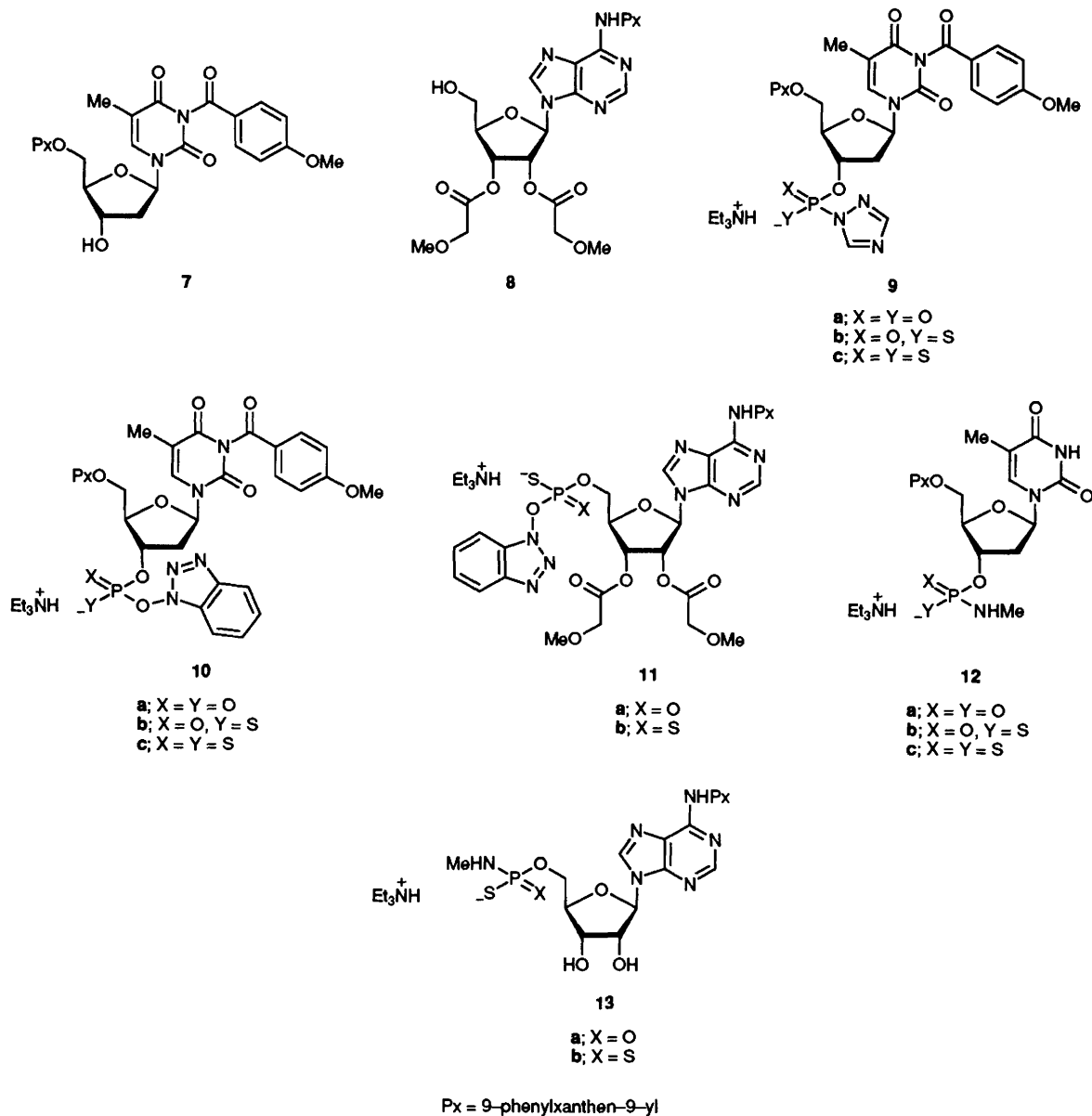


Table 1 Preparation of activated nucleotide derivatives

Entry	Substrate	Phosphorylating agent ^a	Work-up procedure ^b	Product	Eluting solvent (% EtOH) ^c	Yield (%) ^d	δ_p ^e	Purity (%) ^f
1	7	5a	A	9a	15–20	42	–11.1	94
2	7	5b	A	9b	8–10	80	43.1	81
3	7	5b	B	9c	8–10	76	96.3	77
4	7	6a	A	10a	8–10	81	–0.5	100
5	7	6a	B	10b	2–4	49	59.1, 59.2	— ^g
6	7	6b	A	10b	2–4	88	59.1, 59.2	100
7	7	6b	B	10c	2–4	82	125.0	100
8	8	6b	A	11a	2–4	70	59.9	99
9	8	6b	B	11b	2–4	63	125.5	100

^a Phosphorylation reactions were carried out in the following way: POCl₃ (1.5 mmol) or PSCl₃ (1.5 mmol) and Et₃N (4.5 mmol) were added to a stirred suspension of 1*H*-1,2,4-triazole (4.5 mmol) or 1-hydroxybenzotriazole (4.5 mmol) in dry THF (20 cm³) under N₂ at room temp. After 30 min, a solution of substrate (**7** or **8**, 0.5 mmol) in THF (20 cm³) and 1-methylimidazole (1.0 mmol) were added. Phosphorylation reactions were generally complete within 1 h. ^b Procedure A: Et₃N (18 mmol) and water (1.0 cm³) were added to the stirred products at room temp. which, after 20 min, were concentrated under reduced pressure and fractionated; procedure B: Et₃N (18 mmol) was added and dry H₂S gas was bubbled through the mixture for 15–30 min, followed by N₂ for 30 min. The products were then filtered, concentrated under reduced pressure and fractionated. ^c Percentage EtOH by volume in CHCl₃-EtOH mixtures used for short column chromatography on silica gel. ^d Yields relate to precipitated triethylammonium salts, isolated as colourless solids. ^e NMR spectra were measured in (CD₃)₂SO solution; in all cases satisfactory ³¹P and ¹H spectra were obtained. ^f Purity was estimated by HPLC on a Jones APEX ODS column eluted with mixtures of 0.1 mol dm⁻³ aqueous triethylammonium acetate (pH 7.0) and acetonitrile. ^g This product was not examined by HPLC but was found to be pure by ³¹P and ¹H NMR spectroscopy.



solution and the products worked-up with aqueous triethylamine according to procedure A (footnote *b*), the activated nucleotide **9a** was obtained and isolated as a colourless solid in 42% yield. The corresponding thiophosphoro-(1,2,4-triazolide) **9b** was obtained (entry 2) in 80% isolated yield when **5b** was used as the acylating agent and the products were worked-up in the same way. However, when substrate **7** was treated with the same acylating agent **5b** and the products were worked-up with triethylamine and hydrogen sulfide under anhydrous conditions (entry 3) according to procedure B (footnote *b*), the dithiophosphoro-(1,2,4-triazolide) **9c** was obtained in 76% isolated yield. Although, possibly with the exception of the phosphoro-(1,2,4-triazolide) **9a**, satisfactory yields of 1*H*-1,2,4-triazole derived activated nucleotide derivatives (entries 1–3) were obtained, the estimated purity (by HPLC) of the products was disappointing.

When substrate **7** was allowed to react with a threefold excess of tri(benzotriazol-1-yl) phosphate **6a** (entry 4) and the products then worked-up according to procedure A and fractionated by chromatography on silica gel, the nucleotide derivative **10a** was obtained and isolated as a colourless solid in 81% yield. This material was found to be completely homogeneous by HPLC. When the products of the same phosphorylation reaction were worked-up with triethylamine

and hydrogen sulfide (entry 5) according to procedure B, the corresponding phosphorothioate **10b** was obtained. However, the preparation of the latter product **10b** was effected more easily (entry 6) by acylating substrate **7** with tri(benzotriazol-1-yl) phosphorothioate **6b** and then working the products up with aqueous triethylamine according to procedure A. A high yield (88%) of HPLC-homogeneous product was then obtained. When the products of the same acylation reaction were worked-up with triethylamine and hydrogen sulfide under anhydrous conditions (entry 7) according to procedure B, the corresponding phosphorodithioate derivative **10c** was obtained and isolated as an HPLC-homogeneous colourless solid in 82% yield. It can be seen (Table 1) that the 1-hydroxybenzotriazole derived acylating agents **6a** and **b** gave rise to products that were eluted from silica gel with less polar solvent mixtures than were required for the products obtained in reaction involving the 1*H*-1,2,4-triazole derived reagents **5a** and **b**. The fact that the benzotriazole derivatives (**10a**, **b** and **c**) were easier to purify by chromatography on silica gel probably led to generally higher yields of purer products being isolated (Table 1) from reactions involving the 1-hydroxybenzotriazole derived acylating agents **6a** and **b**.

Finally, the adenosine derived building block **8** was allowed to react with **6b**, and the products were worked-up either with

Table 2 Reactions between activated nucleotide derivatives and methylamine in ethanol solution

Entry	Substrate	Reaction time/h ^a	Product	Yield (%) ^b	δ_p^c	Purity (%) ^d
1	9a	2(0.5)	12a	83		93
2	10a	1(0.2)	12a	84	6.6	100
3	9b	8(0.7)	12b	93		97
4	10b	8(0.5)	12b	95	60.3, 60.4	97
5	9c	18(8)	12c	69		97
6	10c	18(6)	12c	79	108.2	98
7	11a	23	13a	96	60.1, 60.3	100
8	11b	18	13b	86	108.1	99

^a Reactions were carried out by dissolving the substrates (ca. 0.2 mmol) in a very large excess (20 cm³) of 8 mol dm⁻³ alcoholic methylamine at room temp. The numbers in parentheses after the actual reaction times represent the estimated (by HPLC) times required for the reactions to go to completion. ^b Isolated yields as precipitated solids following chromatography on silica gel. ^c NMR spectra were measured in (CD₃)₂SO solution; satisfactory ¹H and ³¹P NMR spectra were obtained for all products. ^d Purity was estimated by HPLC (see Table 1, footnote f).

aqueous triethylamine according to procedure A (entry 8) or with triethylamine and hydrogen sulfide according to procedure B (entry 9). In this way, the corresponding phosphorothioate and phosphorodithioate derivatives **11a** and **b**‡ were obtained and isolated as pure colourless solids in 70 and 63% yield, respectively.

All of the activated nucleotides (**9a–c**, **10a–c**, **11a** and **b**) were found to be effective acylating agents in that they reacted with 8 mol dm⁻³ ethanolic methylamine to give the corresponding *N*-methylphosphoramidates in satisfactory to excellent yields (Table 2). In the case of the thymidine derivatives (**9a–c**, **10a–c**), the more readily isolable (see above) 1-hydroxybenzotriazole derivatives (**10a–c**; entries 2, 4 and 6)

were found to be somewhat more reactive and to lead to slightly higher isolated yields of purer products than the corresponding 1*H*-1,2,4-triazole derivatives (**9a–c**; entries 1, 3 and 5). A comprehensive study of the reactions between the above types of activated nucleotide derivatives and various nucleophiles is currently being undertaken in our laboratory.

We thank the K. C. Wong Foundation and the University of London for the award of research scholarships (to Z. Z. and L. H. K. S., respectively).

Received, 19th August 1993; Com. 3/05044K

Footnotes

† **5a** and **6a** are the putative structures of the products generated *in situ* (Table 1, footnote a) from phosphoryl trichloride and 1-*H*-1,2,4-triazole and 1-hydroxybenzotriazole, respectively; **5b** and **6b** are likewise the putative structures of the corresponding products similarly prepared from thiophosphoryl trichloride.

‡ The site of attachment of the 1*H*-1,2,4-triazole and 1-oxybenzotriazole residues to phosphorus has not been firmly established, either in the case of these (**11a** and **b**) or of the other activated nucleotide derivatives (**9a–c**, **10a–c**) described above.

References

- 1 F. Cramer, H. Schaller and H. A. Staab, *Chem. Ber.*, 1961, **94**, 1612.
- 2 H. Schaller, H. A. Staab and F. Cramer, *Chem. Ber.*, 1961, **94**, 1621.
- 3 F. Cramer and H. Neunhoeffer, *Chem. Ber.*, 1962, **95**, 1664.
- 4 D. E. Hoard and D. G. Ott, *J. Am. Chem. Soc.*, 1965, **87**, 1785.
- 5 F. Eckstein and H. Gindl, *Biochim. Biophys. Acta.* 1967, **149**, 35.
- 6 K.-H. Scheit, *Chem. Ber.*, 1968, **101**, 1141.
- 7 C. B. Reese, L. H. K. Shek and Z. Zhao, unpublished results.
- 8 W. Walter and M. Radke, *Liebigs Ann. Chem.*, 1979, 1756.
- 9 A. Kraszewski and J. Stawinski, *Tetrahedron Lett.*, 1980, **21**, 2935.
- 10 K. J. Divakar and C. B. Reese, *J. Chem. Soc., Perkin Trans. 1*, 1982, 1171.