

## A New Tricarbocyclic Diterpene Structure from the Soft Coral *Xenia florida*

Tetsuo Iwagawa,<sup>\* a</sup> Jun-ich Kawasaki,<sup>a</sup> Tsunao Hase,<sup>a</sup> Chao-Mei Yu,<sup>b</sup> John A. Walter<sup>b</sup> and Jeffrey L. C. Wright<sup>\* b</sup>

<sup>a</sup> Department of Chemistry, Faculty of Science, Kagoshima University, 1-21-35 Korimoto, Kagoshima 890, Japan

<sup>b</sup> National Research Council, Institute for Marine Biosciences, 1411 Oxford St, Halifax, Nova Scotia B3H 3Z1, Canada

Floridicin **1**, a diterpene containing a [4.3.1] bicyclic ring system, has been isolated from the soft coral *Xenia florida*, and represents a new class of tricarbocyclic diterpenes.

Coelenterates belonging to the order Alcyonacea (soft corals), Gorgonacea (gorgonians) or Pennatulacea (sea pens), are a rich source of unusual diterpenes, representing a diverse group of structural types.<sup>1,2</sup> This diversity reflects a different folding pattern of a common diterpenoid precursor, and diterpenes isolated from the soft coral *Xenia* spp. usually contain mono- or di-carbocyclic structures.<sup>3</sup> During our investigations of the soft coral *Xenia florida*,<sup>†</sup> collected in the area of Bonotsu, Kagoshima Prefecture, a major component of the lipid-soluble extract was identified as the tricarbocyclic diterpene floridicin **1**.

Frozen coral (12 kg) was extracted with methanol, and this initial extract was partitioned against methylene chloride. Primary purification of this extract was achieved by Si-gel adsorption chromatography and elution with mixtures of hexane, methylene chloride and methanol. The fractions eluted with methanol–methylene chloride mixtures gave oily residues that were purified by further Si-gel chromatography and finally C<sub>18</sub> reversed-phase HPLC and elution with water–methanol mixtures. Following these steps, **1** was obtained as colourless needles (300 mg). The mass spectral data suggested a molecular formula of C<sub>20</sub>H<sub>30</sub>O<sub>4</sub> (6 double bond equivalents)<sup>‡</sup> that was supported by the <sup>13</sup>C NMR data which showed twenty identifiable resonances. The assignment of a single carbonyl ( $\delta$  195.45), and four olefinic resonances ( $\delta$  124.8, 137.69, 146.46, 153.82), required three ring systems to account for the remaining degrees of unsaturation.

Further inspection of the <sup>1</sup>H and <sup>13</sup>C NMR data revealed two olefinic methyl groups ( $\delta$  1.74, 18.72; 1.78, 26.15), weakly coupled to an olefinic proton ( $\delta$  4.96, 124.8), characteristic of a dimethyl allyl group. The remaining double bond was identified as part of a conjugated aldehyde system (UV  $\lambda_{\text{max}}$ /nm 235; IR  $\nu/\text{cm}^{-1}$  2700 and 1680; <sup>1</sup>H NMR  $\delta$  9.30; <sup>13</sup>C NMR  $\delta$  195.45) and the low-field olefinic resonance at  $\delta$  6.38 was assigned to the  $\beta$  proton (H-11) of this conjugated system. The 2D COSY, TOCSY and HMQC data of **1** established the partial structure **a**. An identical moiety is found in a portion of the diterpene **3** isolated from another soft coral species, *Efflatounaria* sp.<sup>4</sup> Structure **3** was established on the basis of NMR and X-ray crystallographic data which at the time represented a new class of coral diterpene molecules. The 1D NOE and 2D NOESY data confirmed that the stereochemistry of structure **a** was similar to that of **3**; irradiation of H-9 resulted in a strong NOE (7.5%) with H-13, as well as

with H-17 (7%). However, despite this structural similarity between portions of **1** and **3**, there was little commonality of NMR data for the remainder of the molecule. An important observation was that in **1**, two ring systems remained unaccounted for, and this was significant since coral diterpenoids possessing more than two carbocyclic rings are extremely rare.<sup>2</sup>

Acetylation of **1** yielded a crystalline diacetate **2** which provided better resolved NMR spectra and so this was used for most of the remaining structural elucidation studies.<sup>§</sup> The unsaturated aldehyde grouping was unperturbed, as judged from the UV ( $\lambda_{\text{max}}$ /nm 235), IR ( $\nu/\text{cm}^{-1}$  2700 and 1680) and NMR (<sup>1</sup>H  $\delta$  9.34) data, which also confirmed the acetate groups (IR:  $\nu/\text{cm}^{-1}$  1745; <sup>1</sup>H NMR  $\delta$  2.05, 3H, and  $\delta$  2.06, 3H) and a remaining tertiary alcohol group ( $\nu/\text{cm}^{-1}$  3600). The <sup>13</sup>C NMR DEPT spectral data supported a molecular formula of C<sub>24</sub>H<sub>34</sub>O<sub>6</sub> (8 double bond equivalents), and confirmed the presence of two acetate groups ( $\delta$  21.22, 168.95 and 21.27, 170.63), an aldehyde group ( $\delta$  192.70) and two double bonds ( $\delta$  121.69, 137.07 and 145.09, 150.78).

The 2D-COSY and TOCSY data of **2** revealed a major spin system that contained H-7, H-8, H-9, H-1, the unsaturated aldehyde group, the dimethyl allyl group, as well as H-12 and H-13, the latter characteristically deshielded following acetylation of the hydroxy group at C-13. This corresponds to the partial structure **a** already identified in **1**, and a characteristic homoallylic coupling between H-9 and H-12 ( $J$  3.7 Hz), also reported in **3**, was now clearly observed in the diacetate. The 2D-NOESY data of the diacetate also confirmed that the stereochemistry of this partial structure was unchanged. In a second spin system, the low-field proton at  $\delta$  4.65 was assigned to H-5, geminal to the second acetate group and part of a system that included the methylenes H-4 ( $\delta$  1.64) and H-3 ( $\delta$  1.71 and 1.98). There remained a singlet methyl resonance at  $\delta$  0.95 and an isolated methylene group H-14, which appeared as an AB quartet at  $\delta$  1.67 and 1.88 ( $J$  14.5 Hz) and was reminiscent of a methylene bridge in a bicyclic ring system.

In both **1** and **2** (excluding the two acetate groups), the partial structure **a** accounts for all but two of the double bond equivalents, which can only be accommodated by two additional ring systems. These ring systems were established

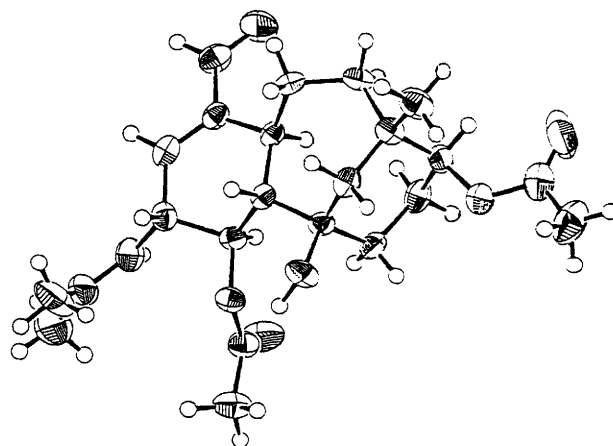
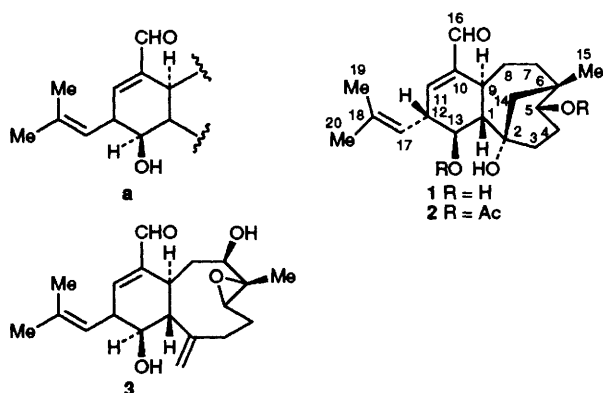


Fig. 1 Perspective ORTEP drawing of floridicin diacetate **2**

by interconnection of the major spin systems through a series of  $^1\text{H}$ -detected long-range  $^1\text{H}$ - $^{13}\text{C}$  correlation experiments (HMBC). A key result was the correlation of the singlet methyl resonance H-15 ( $\delta$  0.95), with C-14 ( $\delta$  43.94), C-7 ( $\delta$  38.10), C-6 ( $\delta$  35.76) and C-5 ( $\delta$  75.50) which established that *all* the spin systems terminated at the quaternary carbon C-6. Another crucial correlation was between the tertiary hydroxy carbon C-2 ( $\delta$  73.57) and the methine H-1 ( $\delta$  2.10) as well as H-14 ( $\delta$  1.67, 1.88), which thus linked these two spin systems through C-2. The [4.3.1] bicyclic structure was completed by linking the spin system containing H-3, H-4 and H-5, through the same low-field quaternary C-2.

The relative orientation of the groups and the *trans*-fusion of the cyclohexene ring system of **2** was established by 2D-NOE correlations between H-1 and H-12 (but not between H-1 and H-13) and between H-9 and H-13. Further proof of the orientation of the isobutenyl group came from an NOE correlation between H-13 and H-17. The stereochemistry of the bicyclic ring system was established by the observation of a crucial NOE correlation between the H-14 methylene bridge proton at  $\delta$  1.88 and H-1, and between the singlet methyl resonance H-15 and both protons ( $\delta$  1.67 and  $\delta$  1.88) of the methylene bridge. No correlation was observed between H-15 and H-5.

Final confirmation of the structure for this new tricyclic diterpene came from X-ray analysis of a crystal of the diacetate $\ddagger$  and the 3-D structure obtained from the analysis is depicted in Fig. 1. This structure represents a new class of tricyclic diterpenes among the plethora of diterpenoid compounds already reported from corals. Interestingly, careful examination of other related *Xenia* spp. from around the Kagoshima area, using the same chemical procedures, failed to uncover any other tricyclic compounds. Further investigations are underway to determine if other examples of the [4.3.1] bicyclic system are produced by *X. florida*.

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### Footnotes

$\dagger$  Specimen sample was identified as *Xenia florida* by Mr F. Iwase, Kushimoto Marine Park Centre, and a voucher sample is deposited at this institute.

$\ddagger$  Data for **1**: Colourless needles (mp 182–183 °C);  $[\alpha]_{\text{D}} -94.5$  (MeOH, *c.* 0.635); UV (MeOH)  $\lambda_{\text{nm}}$  235 ( $\epsilon$  8880); IR (Nujol)  $\nu/\text{cm}^{-1}$

3300, 2700, 1680, 1630, 980, 940, 870; EIMS  $m/z$  334; HREIMS  $m/z$  316.2037  $[(\text{M} - \text{H}_2\text{O})^+]$ , calc.  $\text{C}_{20}\text{H}_{28}\text{O}_3$ ,  $\Delta$  0 mmu];  $^1\text{H}$  NMR (400 MHz;  $\text{CD}_3\text{OD}$ )  $\delta$  (rel. to  $\text{SiMe}_4$ ) 0.99 (s, H-15), 1.4, 1.58 (m, H-7), 1.42, 2.14 (m, H-8), 1.75, 2.07 (m, H-4), 1.72 (m, H-14), 1.74 (d,  $J$  1.47 Hz, H-19), 1.78 (m, H-10), 1.78 (d,  $J$  1.47, H-20), 1.84 (br dd,  $J$  5.1 and 15.4 Hz, H-3), 2.02 (br t,  $J$  13.9 Hz, H-3), 2.11 (dd,  $J$  4.7 and 12.4 Hz, H-1), 2.54 (m, H-9), 3.33 (br s, H-5), 3.36 (dd,  $J$  1.5 and 9.2 Hz, H-12), 3.58 (dd,  $J$  9.2 and 10.4 Hz, H-13), 4.96 (dq,  $J$  1.5 and 9.2 Hz, H-17), 6.38 (t,  $J$  1.8 Hz, H-11), 9.3 (s, H-16);  $^{13}\text{C}$  NMR (100 MHz;  $\text{CD}_3\text{OD}$ )  $\delta$  ( $\text{SiMe}_4$ ) 18.72 (q, C-19), 26.15 (q, C-20), 28.86 (t, C-3), 29.32 (t, C-4), 29.48 (t, C-8), 31.27 (q, C-15), 37.99 (s, C-6), 39.85 (t, C-7), 40.44 (d, C-9), 45.05 (t, C-14), 48.08 (d, C-12), 57.67 (d, C-1), 74.84 (d, C-5), 75.3 (d, C-13), 76.39 (s, C-2), 124.8 (d, C-17), 137.69 (s, C-18), 146.46 (s, C-10), 153.82 (d, C-11), 195.45 (s, C-16).

$\S$  Data for **2**: colourless needles (mp 214–216 °C);  $[\alpha]_{\text{D}} -80$  ( $\text{CHCl}_3$ , *c.* 0.05); UV (MeOH)  $\lambda_{\text{max}}/\text{nm}$  231  $\epsilon$  (20 600); IR (Nujol)  $\nu/\text{cm}^{-1}$  3600, 2700, 1745, 1680, 1640, 1455, 1220; EIMS  $m/z$  418; HREIMS  $m/z$  418.2350 ( $\text{M}^+$ , calc.  $\text{C}_{24}\text{H}_{34}\text{O}_6$ ,  $\Delta$  -0.5 mmu);  $^1\text{H}$  NMR (500 MHz;  $\text{CDCl}_3$ )  $\delta$  0.95 (s, H-15), 1.45, 2.18 (m, H-11), 1.48, 1.69 (m, H-7), 1.64 (m, H-4), 1.67, 1.88 (br d,  $J$  14.5 Hz, H-14), 1.70 (br s, H-19), 1.71, 1.98 (m, H-3), 1.73 (br s, H-20), 2.05 (s, *Ac Me*), 2.06 (s, *Ac Me*), 2.10 (ddd,  $J$  1.3, 11.1 and 11.1 Hz, H-1), 2.63 (m, H-9), 3.56 (dddd,  $J$  1.9, 3.7, 9.8 and 10.1 Hz, H-12), 4.65 (t,  $J$  2.6 Hz, H-5), 4.85 (dq,  $J$  1.4 and 10.1 Hz, H-17), 5.06 (dd,  $J$  9.5 and 10.8 Hz, H-13), 6.34 (t,  $J$  2.0 Hz, H-11), 9.34 (br s, H-16);  $^{13}\text{C}$  NMR (125 MHz;  $\text{CDCl}_3$ )  $\delta$  18.19 (q, C-19), 21.22 (q, C-22), 21.27 (q, C-24), 25.68 (t, C-3), 25.80 (q, C-20), 27.57 (t, C-8), 28.70 (t, C-4), 30.21 (q, C-15), 35.76 (s, C-6), 38.1 (t, C-7), 40.54 (d, C-9), 43.94 (t, C-14), 45.29 (d, C-12), 56.34 (d, C-1), 73.57 (s, C-2), 75.38 (d, C-13), 75.50 (d, C-5), 121.69 (d, C-17), 137.07 (s, C-18), 145.09 (s, C-10), 150.78 (d, C-11), 168.95 (s, *Ac CO*), 170.63 (s, *Ac CO*), 192.70 (s, C-16).

$\P$  Crystal data for **2**:  $\text{C}_{24}\text{H}_{34}\text{O}_6$ , colourless prisms,  $0.1 \times 0.5 \times 0.1$  mm, orthorhombic space group  $P2_12_12_1$ ,  $a = 11.293(1)$ ,  $b = 25.753(4)$ ,  $c = 7.876(1)$  Å,  $U = 2290.4(5)$  Å<sup>3</sup>,  $Z = 4$ ,  $\mu(\text{Cu-K}\alpha) = 6.64$  cm<sup>-1</sup>. Data were collected on an AFC5R Rigaku diffractometer, with graphite monochromated Cu-K $\alpha$  radiation and a 12 kW rotating anode generator; 2004 reflections, of which 1626 had  $I > 3\sigma$  (1), were collected in the range  $6 < \theta < 120.2^\circ$ . The structure was solved by MITHRIL and extended with DIRDIF.<sup>5</sup> It was refined by full-matrix least-squares using the TEXSAN system. It converged with  $R = 0.06$ ,  $R_w = 0.076$  for the 1626 reflections. Atomic coordinates, bond lengths and angles, and thermal parameters have been deposited at the Cambridge Crystallographic Data Centre. See Information for Authors, Issue No. 1.

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