

Synthesis and Separation of Structural Isomers of Tri-*tert*-butylsubphthalocyaninatophenylboron(III)

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Tri-*tert*-butylsubphthalocyaninatophenylboron(III) is synthesized from phthalodinitrile and triphenylboron, whereby a mixture of two structural isomers is obtained; these isomers, with C_3 - and C_1 -symmetry, are separated by HPLC and characterized by ^1H NMR spectroscopy.

Subphthalocyanines **1** ($X = \text{Cl}, \text{Br}, \text{F}$) recently became increasingly interesting due to the fact that they can be used as precursors for the synthesis of unsymmetrically substituted phthalocyanines.^{1–8} As in the case of phthalocyanines, unsubstituted subphthalocyanines **1** ($X = \text{Cl}, \text{Br}, \text{F}$) are practically insoluble in common organic solvents. On the other hand, the tri-*tert*-butyl-substituted subphthalocyanine $\text{Bu}_3\text{SubPcBBr}$ is soluble in organic solvents, exhibiting similarity with tetra-*tert*-butyl-substituted phthalocyaninato analogues.

Recently we described for the first time a new subphthalocyanine **1** ($X = \text{Ph}$) in which the axial substituent is a phenyl group.⁷

We have now prepared the corresponding tri-*tert*-butyl-substituted derivative $\text{Bu}_3\text{SubPcBPh}$ and report here also on the successful chromatographic separation and spectroscopic characterization of its two possible structural isomers (C_1 and C_3) (Scheme 1). The first chromatographic separation of the four possible structural isomers (C_{4h} , D_{2h} , C_{2v} and C_s) of a tetra-substituted phthalocyanine was carried out in our group recently.^{9,10}

A mixture of tri-*tert*-butylsubphthalocyanines, $\text{Bu}_3\text{SubPcBCl}$ and $\text{Bu}_3\text{SubPcBPh}$, both soluble in common organic solvents, starting from *tert*-butylphthalodinitrile and dichloro-

phenylborane in chloronaphthalene as solvent at 230 °C is prepared first. As a side reaction, chlorination of the macrocycle by chlorine generated from PhBCl_2 under the reaction conditions is observed. The mixture of subphthalocyanines obtained is difficult to separate and purify. A careful HPLC-analysis shows, beside the chlorinated products, four peaks, which we assign to the two isomers of $\text{Bu}_3\text{SubPcBCl}$ and $\text{Bu}_3\text{SubPcBPh}$, respectively.

The preparation of only one subphthalocyanine, with a phenyl group as axial substituent, would facilitate the separation of the isomers. For this purpose triphenylboron as the boron-reagent and naphthalene instead of 1-chloronaphthalene as solvent are used. By this approach the unwanted chlorination of the macrocycle is avoided.

The reaction of *tert*-butylphthalodinitrile and BPh_3 (Scheme 1) is carried out in naphthalene at 218 °C. The obtained product $\text{Bu}_3\text{SubPcBPh}$ is prepurified by column chromatography on deactivated Al_2O_3 using toluene as the eluent.

The reaction of *tert*-butylphthalodinitrile with BPh_3 can lead to two structural isomers with the point group symmetry C_3 and C_1 , respectively (Scheme 1), whereby in the mixture four magnetically nonequivalent isoindolenine units should be observable. The C_3 -isomer contains only one magnetically equivalent isoindolenine whereas the C_1 -isomer has three nonequivalent isoindolenine units.

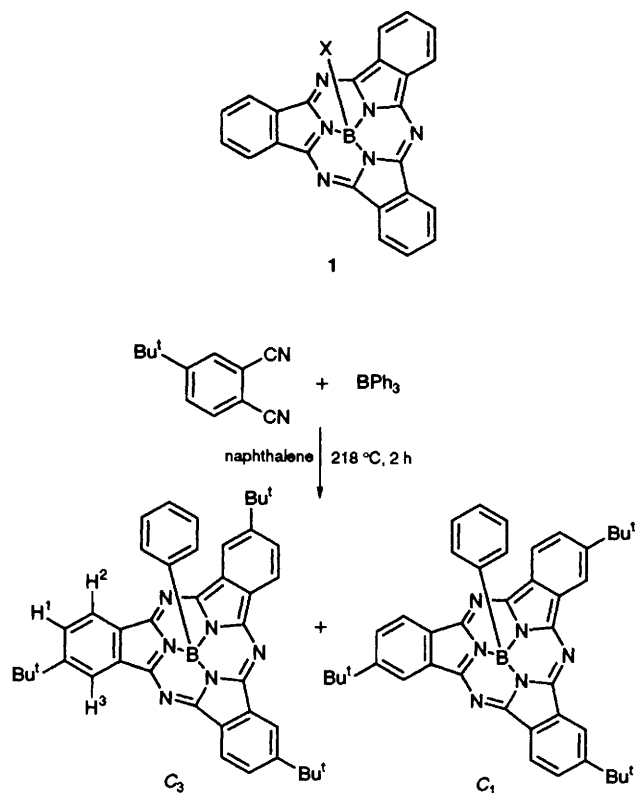
If the four signals for the *tert*-butyl group do not overlap, one should find one signal for the C_3 -isomer and three signals with equal intensity for the C_1 -isomer. The ^1H NMR spectrum of the prepurified $\text{Bu}_3\text{SubPcBPh}$ shows four singlets for the *tert*-butyl groups. This proves that two structural isomers were formed.

The separation of the C_3 - and C_1 -isomers is carried out by HPLC,[†] the peak detection was done by a UV-detector in the region λ 190–600 nm. The HPLC chromatogram shows two completely separated peaks, which were collected by preparative HPLC[‡] (fraction A and fraction B).

The ^1H NMR spectra of both fractions show three signals (δ 5.748–5.786, 6.517–6.581, 6.624–6.695) with an integration ratio of 2:2:1 for the protons of the axial phenyl group. The position of the *tert*-butyl group has no influence on these signals.

The ^1H NMR spectrum of fraction A (250 MHz, C_6D_6)[§] exhibits signals at δ 1.312, 5.748–5.786, 6.517–6.581, 6.624–6.695, 7.604–7.644, 8.839–8.875, 9.050–9.059 with an integration ratio of 27:2:2:1:3:3:3. In the region of the *tert*-butyl group only one signal (δ 1.312) is found. The aromatic protons appear as a doublet of doublets for the H^1 proton with $^3J_{2,1}$ 8.2–8.5 Hz, $^4J_{3,1}$ 1.5–1.8 Hz, a doublet of doublets for the H^2 proton with $^3J_{1,2}$ 8.2–8.5 Hz, $^5J_{3,2}$ 0.6–0.9 Hz and a doublet of doublets for the H^3 proton, $^4J_{1,3}$ 1.8 Hz, $^5J_{2,3}$ 0.6 Hz (Scheme 1). This confirms that fraction A contains the pure C_3 -isomer.

The ^1H NMR spectrum of fraction B (250 MHz, C_6D_6)[§] shows signals at δ 1.301–1.329, 5.748–5.786, 6.517–6.581, 6.624–6.695, 7.615–7.677, 8.854–8.897, 9.053–9.058 with an integration ratio of 27:2:2:1:3:3:3. For the *tert*-butyl group three signals (δ 1.301, 1.316, 1.329) with the same intensity are observed. In the aromatic region two doublets of doublets for the H^1 proton with an integration ratio of 1:2 are found, in addition to two doublets of doublets with the same integration



ratio for the H² proton. Due to the overlap of the expected two doublets of doublet, the signal for H³ shows only one doublet. The signals of the *tert*-butyl group and the aromatic protons respectively prove that fraction B contains the pure C₁-isomer.

In summary we have described for the first time the synthesis and HPLC-separation of the structural isomers of a trisubstituted subphthalocyanine, Bu^t₃SubPcBPh. The point groups are determined unequivocally by ¹H NMR spectroscopy.

Received, 3rd June 1994; Com. 4/03304C

Footnotes

† Beckmann system Gold 5.1: column; Macherey-Nagel, ET 250/8/4, Nucleosil 5NO₂: solvent; hexane-toluene (1:1); sample collection, 1.5 ml min⁻¹

‡ Harley Systems Peakmaster, Sepacon Peakmaster Auto Interface: column; Macherey-Nagel VarioPrep ET250/21, Nucleosil 100-5NO₂: solvent; hexane-toluene (2:3); sample collection, 26 ml min⁻¹

§ Isomer 1: ¹H NMR (C₆D₆) δ 1.312 (s, 27H), 5.748–5.786 (dd, 2H),

6.517–6.581 (m, 2H), 6.624–6.695 (tt, 1H), 7.604–7.644 (dd, 3H), 8.839–8.875 (dd, 3H), 9.050–9.059 (dd, 3H). Isomer 2: ¹H NMR (C₆D₆) δ 1.301–1.329 (3s, 27H), 5.748–5.786 (dd, 2H), 6.517–6.581 (m, 2H), 6.624–6.695 (tt, 1H), 7.615–7.677 (2 dd, 3H), 8.854–8.897 (2 dd, 3H), 9.053–9.058 (d, 3H)

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