

The Synthesis of Eurystatin A¹

Ulrich Schmidt* and Steffen Weinbrenner

Institut für Organische Chemie und Isotopenforschung der Universität Stuttgart, Pfaffenwaldring 55, D-70569 Stuttgart, Germany

Eurystatin A, a new prolyl endopeptidase inhibitor isolated from *Streptomyces eurhythmus* has now been synthesized.

The thrombin inhibitors cyclotheonamide A and B,² the prolyl endopeptidase inhibitor poststatin,³ as well as the prolyl endopeptidase inhibitors eurystatin A and B (**1a** and **1b**)⁴ all contain a β -amino- α -oxocarboxylic acid moiety, which appears to be the common active centre in these enzyme-inhibiting peptides.

During the synthesis of cyclotheonamide B,⁵ the β -amino- α -oxocarboxylic acid unit was formed by oxidation of the corresponding β -amino- α -hydroxy compound. The latter was prepared by the reaction of an α -acylaminoaldehyde with tris(methylthio)methyl lithium and subsequent Hg²⁺-mediated hydrolysis of the trithioorthoester.

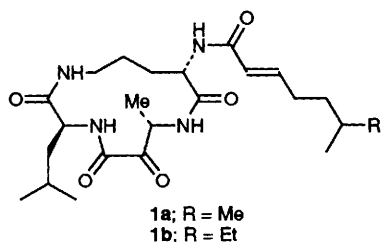
Here, we describe the first total synthesis of eurystatin A (**1a**) in which we have utilized the Passerini reaction for the construction of the diastereoisomeric β -amino- α -hydroxy compound **5**.

The synthesis of the β -amino- α -hydroxybutyric acid and its conversion into dipeptide **4** can be achieved in one step by the Passerini reaction of (*S*)-Z-alanine (**3**) with methyl (*S*)-2-isocyano-4-methylpentanoate (**2**). Compound **2** was obtained in an optically pure form by the triphosgene dehydration⁶ of (*S*)-*N*-formylleucinate. No optical induction can be observed during the Passerini reaction and the product is a 1 : 1 mixture of the diastereoisomers.

Saponification of **4** and replacement of the benzyloxycarbonyl group by a *tert*-butyloxycarbonyl protecting function furnished the carboxylic acid **5**. Coupling with benzyloxycarbonyl ornithine methyl ester gave the diastereoisomeric products **6a** and **6b**, whose configurations could not be elucidated. In an attempt to avoid the difficulties associated with the reactions and the characterisation of diastereoisomeric mixtures in subsequent steps, the mixture of **6a**, **6b** was oxidized. However, saponification of the α -oxocarboxylate was not successful and only decomposition products were isolated. Thus, the diastereoisomers **6a**, **6b** were separated by medium pressure LC and subsequent reactions were carried out with the pure diastereoisomers.

For the peptide ring closure we introduced the pentafluorophenyl ester.⁷ This procedure has been our method of choice for several years in the synthesis of natural biologically active cyclopeptides. The construction of the cyclopeptide alkaloids zizyphine A and B,⁸ mucronine,⁹ dihydrozizyphines A, B¹⁰ and G¹¹ and frangulane,¹² of the cytostatic cyclotetrapeptides chlamydocine¹³ and WF-3161¹⁴ and of the antibiotic glidobactin¹⁵ have been mostly realized in 80–95% yields by catalytic hydrogenation of the corresponding linear ω -Z-amino pentafluorophenyl esters with palladium/charcoal. This cyclisation process takes place on the surface of the palladium where the free amino group is adsorbed after the Z-group has been cleaved.

In the cases of the synthesis of the thiazole-containing

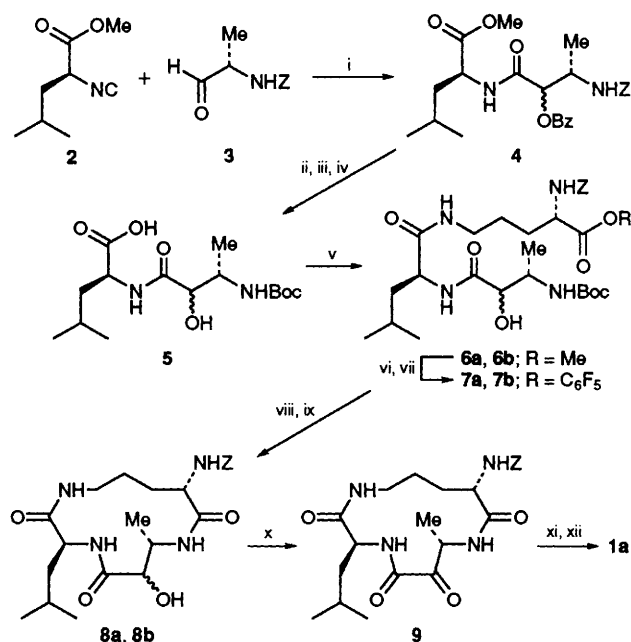


cyclopeptides—ulicyclamide,¹⁶ ulithiacyclamide,¹⁷ patellamide B¹⁸ and the dolastatin³ isomers¹⁸—where catalytic deprotection of the benzyloxycarbonyl group is not possible, ring closure has been achieved by reacting the ω -amino pentafluorophenyl esters (obtained from ω -Boc-amino pentafluorophenyl esters) with a base in dioxane. If this reaction is performed in the two-phase system chloroform–hydrogen carbonate–water without the use of high dilution conditions often the yield is so high¹⁹ within a few minutes that we believe that the conformations of the transition state for the cyclisation is the same as those of the linear substrate.

The synthesis of the cytostatic didemins,²⁰ the antibiotics biphenomycin A²¹ and B,²² the encyminhibitor OF-4949²³ and of the ACE inhibitors lyciumin A and B²⁴ have been realized in high yields by this procedure.

Saponification of **6a** and **6b** and esterification with pentafluorophenol afforded the esters **7a** and **7b**, the starting materials for the ring closure step. The Boc protection was removed to leave the hydrochloride and ring closure was realized in good yields (72% for **8a**, 63% for **8b**, each over four steps) in the previously described two-phase system composed of chloroform and aqueous sodium hydrogen carbonate.

Swern oxidation, which has been used with success for linear β -acylamino- α -hydroxycarboxylic acid derivatives (such



Scheme 1 Reagents and conditions: i, benzoic acid, CH₂Cl₂, room temp., 48 h, 85%; ii, MeOH, Cs₂CO₃, 15 min, room temp., 98%; iii, MeOH, Boc₂O, Pd/C, H₂, 2 h, 96%; iv, LiOH, H₂O–dioxane, room temp., 96%; v, Z-(H)-Orn*HCl, 2-(1H-benzotriazol-1-yl)-1,1,3,3-tetramethyluronium tetrafluoroborate (TBTU), CH₂Cl₂, 35% **6a**, 25% **6b**; vi, LiOH, H₂O–dioxane, room temp.; vii, C₆F₅OH, *N*-(3-dimethylaminopropyl)-*N'*-ethylcarbodiimide hydrochloride (EDC), –15 to 20 °C, 16 h; viii, 6 mol dm⁻³ HCl, dioxane, 2 h, 20 °C; ix, CHCl₃–NaHCO₃, room temp., 6 h, vi–ix, **6a** to **8a** 72%, **6b** to **8b** 63%; x, pyridinium dichromate, DMF, room temp., 48 h, **8a** to **9** 75%, **8b** to **9** 30%; xi, MeOH–CH₂Cl₂, Pd/C, H₂, 2 h; xii, DMF–CH₂Cl₂, (*E*) 6-methylhept-2-enoyl chloride, pyridine, room temp., 3 h, xi + xii, 50%

as **6**), failed in the cases of the cyclic compounds **8a** and **8b**. However, the required transformation was achieved with pyridinium dichromate and it was found that the diastereoisomer **8a** reacted to give a higher yield (75%) than **8b** (30%).

Subsequent hydrogenolytic deprotection of **9†** and acylation with (*E*)-6-methylhept-2-enoyl chloride then completed the synthesis of eurystatin A (**1a**).‡ The product thus synthesized was found to be identical in all respects (NMR, optical rotation, melting point and MS) with the naturally occurring compound.

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Footnotes

† δ_{H} (250 MHz, $(\text{CD}_3)_2\text{SO}$) 8.83 (d, *J* 7.7, 1H), 8.23 (d, *J* 8.1, 1H), 7.45–7.15 (m, 7H), 5.05 (d, *J* 12.5, 1H), 4.98 (d, *J* 12.5, 1H), 4.69–4.64 (m, 1H), 4.12–4.10 (m, 2H), 3.04 (br, 2H), 1.61–1.48 (m, 7H), 1.16 (d, *J* 6.7, 3H), 0.89 (d, *J* 5.7, 3H) and 0.82 (d, *J* 5.7, 3H).

‡ δ_{H} (250 MHz, $(\text{CD}_3)_2\text{SO}$) 8.82 (d, *J* 7.7, 1H), 8.21 (d, *J* 8.5, 1H), 8.02 (d, *J* 7.3, 1H), 7.43 (br, 1H), 6.61 (dt, *J* 15.4, 6.8, 1H), 6.03 (d, *J* 15.4, 1H), 4.76–4.69 (m, 1H), 4.21–4.18 (m, 1H), 4.12–4.06 (m, 1H), 3.10–3.03 (m, 2H), 2.17–2.10 (m, 2H), 1.82–1.48 (m, 8H), 1.32–1.23 (m, 2H), 1.15 (d, *J* 6.7, 3H), 0.91–0.89 (m, 12H).

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