Efficient Synthesis of α -Purine Nucleosides of 2-Deoxyribofuranose and 2-Deoxyribopyranose

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A high yield synthesis of α -purine nucleosides of 2-deoxyribofuranose and 2-deoxyribopyranose is reported starting from 1,3,5-tri-*O*-acetyl-2-deoxyribopyranose with silylated purines under Lewis acid (SnCl₄) catalysed conditions.

Structural variants of oligodeoxynucleotides are valuable tools for studying protein-DNA interactions and are promising antisense agents in controlling gene expression.¹ α -Anomers of oligodeoxynucleotides² have been shown to form duplexes with complementary DNA or RNA via Watson-Crick hydrogen bonds.³ The increased thermal stability of α/β duplexes⁴ and their nuclease resistance⁵ make them promising antisense therapeutic agents for in vivo application. Hence it is highly desirable to have stereoselective procedures that allow large scale synthesis. We have investigated the coupling reactions of various 2-deoxyribose derivatives with silvlated bases in presence of Lewis acids. Here we report that the stannic chloride mediated coupling reactions of 1,3,5-tri-O-acetyl-2-deoxyribofuranose and 1,3,4-tri-O-acetyl-2-deoxyribopyranose with suitably protected purine derivatives proceed in very high yields and give excellent α -anomeric stereoselectivities. Suitably protected purines yield the desired N-9 regiochemistry while unprotected purines lead to mixtures of N-7 and N-9 regioisomers.⁶ In comparison with the previous methods for preparation of α -anomers by Lewis acid catalysed conditions,⁷ our method is highly efficient. Although our yields are comparable to the recently reported phase transfer glycosidation methods⁸ our procedure gives higher α -anomeric selectivity in the case of adenosine. More significantly, our base protection scheme is superior for oligonucleotide synthesis by phosphoramidite or phosphodiester methods.

1,3,5-tri-O-acetyl-2-deoxyribose 1 was prepared starting from 2-deoxyribose.⁹ N-benzoyl adenine (1.3 equiv.) was silylated using N,O-bistrimethylsilyl acetamide (BSA) (2.6 equiv.) in acetonitrile and the silyl derivative thus obtained was condensed with the sugar derivative 1 in the presence of SnCl₄ (1.3 equiv.) as devised by Vorbruggen *et. al.*¹⁰ After



Scheme 1 Reagents and conditions: i, $B-[Si(Me)_3]_2$, $SnCl_4$, MeCN, room temp.

12 h of stirring at room temp., TLC showed the complete disappearance of starting material. After aqueous workup and chromatographic separation, compound 2 was isolated in 95% yield[†] with an α : β ratio of 78: 22.[‡] The sugar acetate 1 under the reaction conditions does not give rise to any side product, whereas the 1-O-methyl-3,5-di-p-toluoyl-2-deoxyribose under identical conditions gave the acyclic nucleosides arising via silvlation of the ring oxygen¹¹ along with the expected cyclic nucleoside 2. Furthermore the reaction proceeds to completion with only 1.3 equiv. of silvlated base in a shorter period of time, yielding higher α -anomeric ratios. The choice of solvent, acetonitrile, proved to be critical for the success of the reaction. The reaction conducted in 1,2-dichloroethane under similar conditions resulted in a mixture of products from which the required nucleoside 2 was isolated in 60% yield with an anomeric ratio of α : $\beta = 68:32$.

Extension of this study to guanine nucleosides was then attempted. Of the several protecting groups¹² available for 6-O- of guanine, we chose diphenylcarbamoyl group,¹³ since it gives an organic-soluble derivative for further elaborations. Treatment of the sugar acetate 1 with silylated 2-N-acetyl-6-Odiphenylcarbamoyl guanine (1.3 equiv.) in the presence of $SnCl_4$ (1.3 equiv.) in acetonitrile led to a mixture of products. Analysis of the products showed that the sugar acetate as well as the silvlated guanine derivative undergoes decomposition. Similar guanine decomposition was also observed in the case of the reaction of 1-O-methyl-3,5-di-p-toluoyl-2-deoxyriboside with silylated guanine derivatives and 1.0 equiv. of SnCl₄. This observation prompted us to study the reaction with increasing amounts of SnCl₄. At 10 equiv. of SnCl₄ the reaction was complete in 16 h and the required nucleoside 3 was obtained in 80% yield with an anomeric ratio of α : β = 68:32. The reaction in 1,2-dichloroethane gave the nucleoside **3** in only 40% yield, with an anomeric ratio of α : $\beta = 75$: 25. These results show that the success of the nucleoside synthesis depends not only on the reaction conditions, the sugar derivatives and the Lewis acid catalyst but also on the reacting nucleobase. The acetate protection for the hydroxy groups is a better choice than benzoyl, p-nitrobenzoyl or p-toluoyl groups, since the O-acetyl groups can be selectively cleaved without affecting the N-acetyl group.14 Thus it avoids the additional steps needed in protecting the amino groups of purine nucleosides prior to phosphitylation.7,15

In the case of 2-deoxyribopyranose nucleosides, a recent report¹⁶ showed the adenine derivative to be obtained in only 50% yield under Lewis acid (SnCl₄) catalysed conditions in 1,2-dichloroethane. Hence we reinvestigated this reaction using acetonitrile as the solvent. 1,3,5-tri-*O*-acetyl-2-deoxyribopyranose **4** was prepared by reacting 2-deoxyribose with acetic anhydride in pyridine. Reaction of the pyranose sugar acetate with silylated adenine (1.3 equiv.) in the presence of SnCl₄ (1.3 equiv.) gave the required nucleoside **5** in 85% yield, with an anomeric ratio of $\alpha:\beta = 52:48$. Extending the reaction to guanine nucleoside was also successful. Treatment of the sugar derivative and the silylated 2-*N*-acetyl-6-*O*-diphenylcarbamoyl guanine in the presence of SnCl₄ (10 equiv.) gave the nucleoside **6** in 80% yield after chromatographic separation, with an anomeric ratio of $\alpha:\beta = 60:40$.

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Footnotes

† All reported compounds showed satisfactory spectral data.

[‡] Anomeric ratios were determined using ¹H NMR (300 MHz) spectrum by integration of 1'-H, 3'-H or 8-H protons for adenosine and guanosine derivatives. The α -anomer of 2-N-3',5'-tri-O-acetyl-6-O-diphenylcarbamoyl-2'-deoxyguanosine is separable from the mixture by fractional crystallization (dichloromethane-diethyl ether = 1:1). Separation of the α -anomer of 6-N-benzoyl-3',5'-di-O-acetyl-2'-deoxyadenosine proved to be difficult. However, α - and β -anomers of 6-N-benzoyl-5'-dimethoxytrityl-2'-deoxyadenosine are easily separable by silica gel chromatography (0-5% methanol in chloroform containing 0.1% pyridine).

References

- 1 J. Goodchild, *Bioconjugate Chem.*, 1990, 1, 16; E. Uhlmann and A. Peyman, *Chem. Rev.*, 1990, **90**, 543; S. L. Beaucage and R. P. Iyer, *Tetrahedron*, 1993, **49**, 6123.
- 2 U. Sequin, Experientia. 1973, 29, 1059.
- 3 For example, see J. S. Sun, U. Asseline, D. Rouzaud, T. Montenary-Garestier, N. T. Thuong and C. Helene, *Nucl. Acids Res.*, 1987, 15, 6149; F. Morvan, B. Rayner, J. L. Imbach, D. K. Chang and J. W. Lown, *Nucl. Acids Res.*, 1987, 15, 4241; F. Morvan, B. Rayner, J. L. Imbach, M. Lee, J. A. Hartley, D. K. Chang and J. W. Lown, *Nucl. Acids Res.*, 1987, 15, 7027.
- 4 C. Gagner, J. R. Bertrand, S. Thenet, M. Lemaitre, F. Morvan,

B. Rayner, C. Malvy, B. Lebleu, J. L. Imbach and C. Paoletti, Nucl. Acids Res., 1987, 15, 10419.

- 5 F. Morvan, B. Rayner, J. L. Imbach, S. Thenet, J. R. Bertrand, J. Paoletti, C. Malvy and C. Paoletti, *Nucl. Acids Res.* 1987, 15, 3421; N. T. Thuong, U. Asseline, U. V. Roig, M. Takasugi and C. Helene, *Proc. Natl. Acad. Sci.*, USA. 1987, 84, 5129.
- 6 L. W. Dudycz and G. E. Wright, J. Med. Chem., 1984, 27, 175; H. Kawakami, H. Mathushita, M. Shibagaki, Y. Naoi, K. Itoh and H. Yoshikoshi, Chem. Lett., 1989, 1365. M. J. Robins, R. Zou, F. Hansike, D. Madej and D. L. T. Tyrell, Nucleosides and Nucleotides, 1989, 8, 625. P. Garner and S. Ramakanth, J. Org. Chem., 1988, 53, 1294.
- 7 T. Yamaguchi and M. Sameyoshi, *Chem. Pharm. Bull.*, 1984, **32**, 1441; M. Imazawa and F. Eckstein, *J. Org. Chem.*, 1978, **43**, 3044.
- 8 M. V. Baud, C. Chavis, M. Lucas and J. L. Imbach, *Tetrahedron Lett.*, 1990, **31**, 4437.
- 9 E. Fischer, Ber. Dtsch. Chem. Ges., 1945, 28, 1145; M. Hoffer, Chem. Ber., 1960, 93, 2777; Y. Ichikawa, H. Kubota, K. Fujita, T. Okauchi and K. Narasaka, Bull. Chem. Soc. Jpn., 1989, 62, 845.
- U. Niedballa and H. Vorbruggen, J. Org. Chem., 1974, 39, 3654.
 H. Vorbruggen, K. Krolikiewicz and B. Bennua, Chem. Ber., 1981, 114, 1234.
- P. T. Jorgensen, E. B. Pedersen and C. Neilsen, Synthesis, 1992, 1299; A. A. El-Barbary, A. I. Khodair and E. B. Pedersen, J. Org. Chem., 1993, 58, 5994; S. Janardhanam and K. P. Nambiar, Tetrahedron Lett. submitted for publication.
- 12 F. P. Clausen and J. L. Christensen, Org. Prep. Proc. Intl., 1993, 375.
- 13 R. Zuo and M. J. Robins, Can. J. Chem., 1987, 65, 1436.
- 14 H. Koster, K. Kulikowski, T. Liese, W. Heikens and V. Kohli, *Tetrahedron*, 1981, **37**, 363.
- 15 G. S. Ti, B. L. Gaffney and R. A. Jones, J. Am. Chem. Soc., 1982, 104, 1316.
- 16 M. Baud, C. Chavis, M. Lucas and J. L. Imbach, *Tetrahedron*, 1991, 47, 9993.