

Investigation of the Stereospecificity of Clavamincic Acid Synthase in the Desaturation of Dihydroclavamincic Acid to Clavamincic Acid

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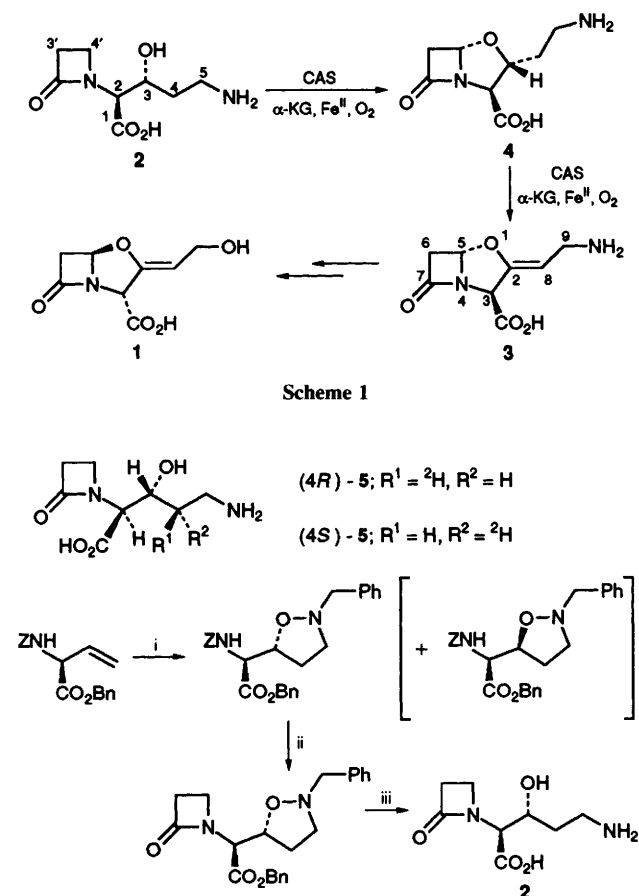
Incubations of (4*R*)- and (4*S*)-[4-²H₁]-proclavamincic acid with clavamincic acid synthase resulted in the stereospecific removal of the deuterium and hydrogen respectively from C-4, in their conversions to clavamincic acid, suggesting an enzyme catalysed *syn*-elimination for the desaturation of dihydroclavamincic acid to clavamincic acid.

Clavamincic acid synthase (CAS),¹ isolated from *Streptomyces clavuligerus* ATCC 27064,² is one of the enzymes involved in the biosynthesis of the β-lactamase inhibitor clavulanic acid 1.^{2,3} The key function of CAS is conversion of the monocyclic β-lactam, proclavamincic acid 2^{4,5} to the bicyclic clavamincic acid 3,⁴ via the dihydroclavamincic acid intermediate 4⁶ (Scheme 1). This process requires ferrous ion as co-factor and α-ketoglutarate (α-KG) and molecular oxygen as co-substrates and involves two distinct chemical events, namely closure of the oxazolidine ring to give 4 and subsequent desaturation to form the exocyclic double bond of 3.⁶

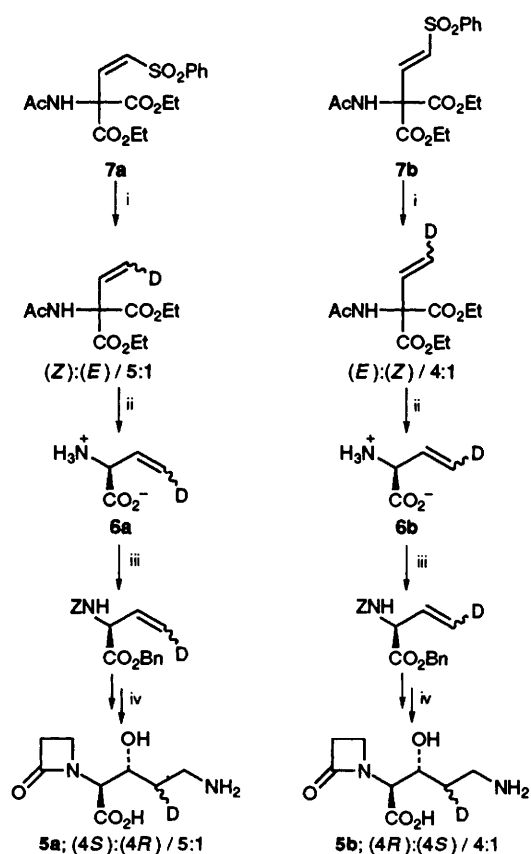
Labelling experiments have shown that the cyclisation of 2 to 3 (via 4) occurs stereospecifically,⁷ with the 4'-pro-(*S*) hydrogen being replaced by oxygen from the substrate. The aim of this work was to investigate the stereospecificity of the

desaturation of 4 to 3 by incubation of (4*R*)- and (4*S*)-[4-²H₁]-proclavamincic acids 5 with CAS.

The synthesis of hydroxyornithine, based on a 1,3-dipolar cycloaddition of *N*-benzyl nitron to vinyl glycine, has been described by Wityak and Gould.⁸ Elaboration of a β-lactam moiety onto the cycloadduct and hydrogenation provides Townsend's route to proclavamincic acid (Scheme 2).⁹ In order to incorporate the deuterium label in 5, (*E*)- and (*Z*)-[4-²H₁]-vinyl glycines 6 were required, the stereospecific olefinic synthesis of these amino acids in racemic form had been described by Sawada and Hill from the (*E*)- and (*Z*)-vinyl sulfones 7, respectively, by reduction with aluminium amalgam.¹⁰ Despite repeated efforts the stereospecificity of the sulfone reduction could not be reproduced in greater than a 5:1 ratio of retention to inversion. This was considered adequate to proceed as the (*E*):(*Z*) ratio of deuterium



Scheme 2 Reagents and conditions: i, (a) *N*-benzyl nitron, benzene, reflux, (b) separation of diastereoisomers by chromatography; ii, (a) HBr in AcOH, (b) *tert*-butyl acrylate, (c) TFA, (d) PPh₃, dipyridyl disulfide, MeCN, reflux; iii, Pd(OH)₂/C, H₂ (3 atm.), EtOH:H₂O (1:1 v/v)



Scheme 3 Reagents and conditions: i, Al/Hg (D₂O), dioxan, 10 °C, 48 h; ii, (a) 6 mol dm⁻³ HCl, reflux, 4 h (b) ClCH₂COCl, 1 mol dm⁻³ NaOH, Et₂O:H₂O (1:1 v/v); (c) enzymatic resolution using Acylase I from porcine kidney; iii, (a) *N*-benzyloxycarbonyloxysuccinimide, CH₂Cl₂, NEt₃, then H₃O⁺, extract; (b) PhCHN₂; iv, reagents as shown in Scheme 2

incorporation in the vinyl compounds was clearly discernible by 500 MHz NMR and, since the protons at C-4 in **2** are diastereotopic, the ratio of deuterium incorporation in **5** could also be clearly established. The (2*S*)-[4-²H₁]-vinyl glycines **6a** and **6b** were obtained by enzymatic resolution¹¹ of the racemic olefins; a necessary procedure for the synthesis of the optically pure proclavaminc acids. The route to (4*R*)- and (4*S*)-**5** is shown in Scheme 3. Electrospray ionisation mass spectrometry revealed the deuterium incorporation in **5** to be greater than 98%; 500 MHz NMR showed **5a** to have 5:1 (*S*):(*R*) stereochemistry at C-4 and **5b** to have 4:1(*R*):(*S*) stereochemistry at C-4.

Incubation of **5a** with CAS† resulted in the formation of **3a** as the major enzymatic product, whereas incubation of **5b** with CAS resulted in the formation of **3b** as the major enzymatic product (Scheme 4). The ratio of **3a**:**3b** in each case was consistent with the ratio of deuterium incorporation at C-4 in the substrates. The products from the incubation mixture were isolated by reverse phase HPLC and characterised by ¹H NMR spectroscopy and mass spectrometry [¹H NMR data as previously reported⁶ with the absence of the triplet at δ 4.86 in the case of **3a** (*m/z*, Table 1)]. Recovered **5** from each incubation mixture showed the same (*S*):(*R*) ratio for deuterium incorporation at C-4 as in the fed substrate, implying that a kinetic isotope effect was not observable for

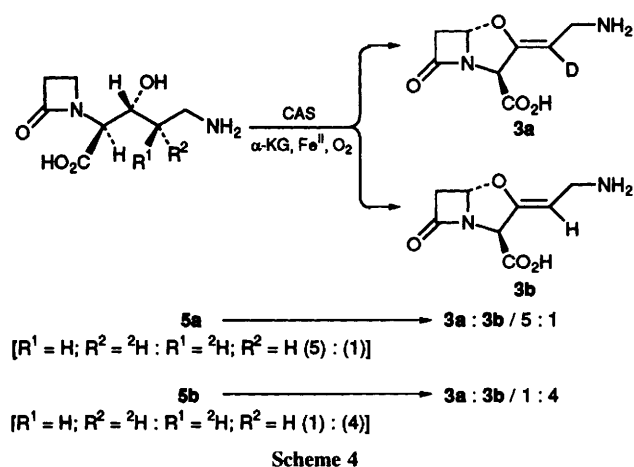


Table 1 Electrospray ionisation mass spectrometry results, *m/z* for M + H, for clavaminic acid **3a/b** produced by incubation of **2**, **5a** and **5b** with CAS; (a) incubation of **2**, (b₁, b₂) incubation of **5a**, (c₁, c₂) incubation of **5b**.

| <i>m/z</i> (%) | 198 | 199 | 200 | 201 | 202 | Ratio of 3a : 3b |
|-------------------|-----|------|------|------|-----|--------------------------------|
| (a) | 0.0 | 100 | 10.5 | 10.0 | 0.0 | — |
| (b ₁) | 2.0 | 19.5 | 100 | 13.0 | 6.0 | 5.0:1 |
| (b ₂) | 0.0 | 18.0 | 100 | 11.0 | 6.0 | 5.4:1 |
| (c ₁) | 0.5 | 100 | 37.5 | 6.0 | 2.5 | 1:3.7 |
| (c ₂) | 3.0 | 100 | 33.5 | 6.0 | 4.0 | 1:4.3 |

hydrogen/deuterium abstraction at this centre. The experiment was repeated with identical results.

These results, combined with the previously reported structure of dihydroclavaminc acid **4**⁶ and the known double bond geometry in clavaminic acid **3**, suggest that the desaturation step occurs *via* a CAS catalysed *syn*-elimination. This stereochemical result is the same as was observed for the desaturation of stearic acid to oleic acid in bacteria¹² and algae.¹³

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Footnote

† Incubations were carried out under standard conditions⁶ utilising 1.1–1.2 IU of CAS with 0.5 mg of the deuterated substrate.

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