

Cyclisation and Aromatisation of Carotenoids during Sediment Diagenesis

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A novel diaryl isoprenoid with an additional aromatic ring (C₄₀H₅₈), formed from the diaromatic carotenoid isorenieratene by cyclisation and aromatisation during sediment diagenesis, is identified in carbonaceous sedimentary rocks.

A number of carbonaceous sediments contain substantial amounts of the diaryl isoprenoid isorenieratane **1**,^{1–3} which is derived from the diaromatic carotenoid isorenieratene **2**, a carotenoid exclusively biosynthesized by photosynthetic green sulfur bacteria (Chlorobiaceae).⁴ Upon GC analyses of the polyaromatic fractions of extracts of a number of sediments (*e.g.* Kimmeridge Clay, Schistes Cartons, Calcaires en Plaquettes, Allgäu Formations) compound **3** elutes just before **1** and possesses a mass spectrum [*m/z* 538(20), 173(12), 134(36), 133(100), 119(22%)], suggesting that it is isorenieratane **1** with

an additional aromatic ring formed by cyclisation and aromatisation of the isoprenoidal acyclic part of **1**. The MS, however, does not reveal any strong indications for the position of the additional ring. This is not surprising since MS of 1,2-dialkylbenzenes hardly contain fragment ions due to cleavage of one of the alkyl side chains β to the aromatic ring, but instead are dominated by a fragment formed by loss of both alkyl side chains.⁵ Indeed, the MS of **3** contains a fragment ion at *m/z* 119, which is more abundant than in the MS of **1** (22 vs. 10%), suggesting the presence of a trialkylbenzene moiety.

The genetic relationship between **1** and **3** was further established by comparison of their ¹³C content in two sediments (Table 1) as determined by isotope ratio monitoring-GC-MS. Both **1** and **3** are uniquely enriched by *ca.* 14 parts per thousand relative to lipids derived from algae. This is consistent with their derivation from photosynthetic green sulfur bacteria since these organisms fix CO₂ through the reversed TCA cycle leading to biomass anomalously enriched in ¹³C.⁶

To fully determine the structure of **3**, it was isolated from the sedimentary rock extract of the Lower Jurassic Allgäu Formation by column chromatography and subsequent reversed phase HPLC using a Polygosil C₁₈ preparative column eluting with MeOH–EtOAc (90–10; *v/v*). The purest fraction (2.3 mg) consisting of 74% of **3** (other 26% consisted of a complex mixture of hydrocarbons, none representing more than 3% of the total fraction) was analysed by ¹H and ¹³C NMR.† These data allowed discrimination between the five possible isomers **3–7**, which can be formed by cyclisation and aromatisation of **1**. From the COSY spectrum of **3** it was deduced that all three aliphatic methyl groups were coupled with protons attached to carbon atoms not α to the aromatic rings as indicated by their chemical shifts in the range δ 1.5–1.8. Furthermore, four CH₂ groups next to an aromatic ring were clearly revealed in the COSY spectrum. Since **3** is the only structure which shows these features, its identity was established. Other shifts are in agreement with this assignment.

The similarity in structure and ¹³C content of **1** and **3** establishes a direct link between these sedimentary components and their precursor, isorenieratene **2**. The identification of **3** is the first evidence for the process of cyclisation and aromatisation of carotenoids during sediment diagenesis. In fact, the formation of **3** is only one step in this process since components

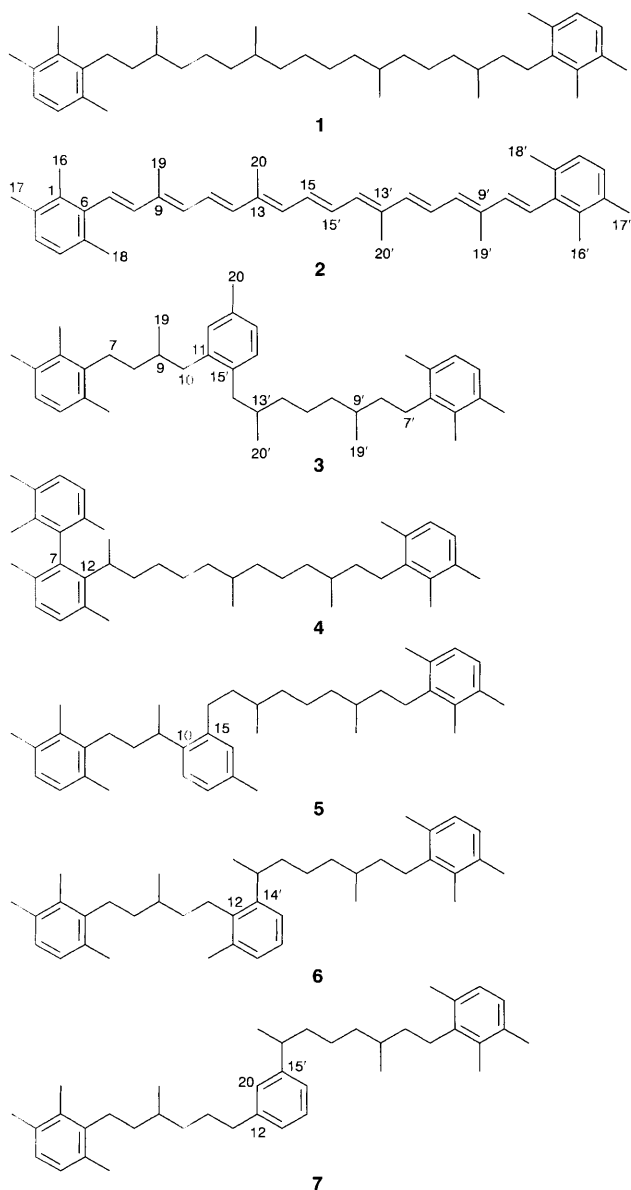


Table 1 δ¹³C (parts per thousand vs. PDB^a) values of selected sedimentary components

Formation name	δ(¹³ C)			
	1	3	Pristane	Phytane
Kimmeridge Clay	−16.6 ±0.3 ^b	−16.3 ±0.2	−30.0 ±0.3	−30.1 ±0.5
Allgäu	−19.0 ±0.4	−19.0 ±0.3	−33.5 ±0.8	−33.5 ±0.2

^a PDB indicates PeeDee belemnite, the stable carbon isotope standard.

^b Standard deviation from three replicates

with the isorenieratene carbon skeleton containing up to four additional (condensed) aromatic rings have been identified in sediments.⁷

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Footnote

† Selected NMR data for **3**: ¹H (400 MHz, CDCl₃), 2.65 (m, CH₂, C-7'), 2.58 (m, CH₂, C-7), 2.55 (m, CH₂, C-10), 2.41 (m, CH₂, C-14'), 2.23 (s, Me, C-20), 1.86 (m, CH, C-9), 1.68 (m, CH, C-13'), 1.52 (m, CH, C-9'), 1.00 (d, *J* 6.7 Hz, Me, C-19), 0.97 (d, *J* 6.7 Hz, Me, C-19'), 0.85 (d, *J* 6.7 Hz, Me, C-20); Shifts for C-8, C-8', C-10' and C-12' were not determined. ¹³C (100 MHz; CDCl₃) 19.6 (Me, C-19'), 19.5 (Me, C-19),

19.5 (Me, C-20'); these assignments may be interchanged, 21.0 (Me, C-20).

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