## Electrophilic Azidation of 2-Deoxy-aldono-1,5-lactones: an Alternative Route to 2-Azido-2-deoxy-aldopyranoses

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Electrophilic azidation of tri-*O*-benzyl-2-deoxy-D-galactono-1,5-lactone **3** with triisopropylphenylsulfonyl azide, followed by selective reduction with diisobutylaluminium hydride, yielded tri-*O*-benzyl-2-azido-2-deoxy-D-galactopyranose **5** as the sole product in 80% yield, while the same sequence of reactions with the 2-deoxy-glucono-1,5-lactone derivative **8** afforded only tri-*O*-benzyl-2-azido-2-deoxy-D-mannopyranose **10** in 65% yield.

Over the past decade the carbohydrate units of glycoconjugates (glycans) have received increasing attention in both academic and industrial sectors.<sup>1</sup> Particularly, the recent discovery of carbohydrate-mediated cell-cell interactions associated with inflammation and cancer metastasis has initiated intensive research focused on the development of carbohydrate-based therapeutics.<sup>2</sup> Glycans often contain 2-amino-2-deoxy-aldopyranosides as their building blocks. For glycan synthesis the 2-azido-2-deoxy derivatives of mono- and di-saccharides are versatile intermediates.<sup>3</sup> These azido sugars have been prepared by (a) azidonitration,<sup>4</sup> azidohalogenation<sup>5</sup> or azidophenylselenylation<sup>6</sup> of O-protected glycals, (b) azidolysis of the 2,3-epoxide ring in 1,6-anhydro-sugars,<sup>7</sup> (c) azide displacement of 2-O-sulfonate derivatives,<sup>8</sup> or (d) the direct<sup>9</sup> or stepwise<sup>10</sup> transformation from 2-amino-2-deoxy-sugars.

Electrophilic azide transfer to enolates using arylsulfonyl azides has been studied extensively and proven to be a general approach to the asymmetric synthesis of  $\alpha$ -amino acids. Herein, we report our preliminary findings that electrophilic azidation is highly stereoselective for the preparation of 2-azido-2-deoxy-aldopyranoses.

The 2-deoxy-aldono-1,5-lactones  $3^{12}$  and  $8^{13}$  were prepared in two steps from readily available tri-O-benzyl-D-glycals  $1^5$  and 6,  $1^4$  respectively (Scheme 1). Treatment of 1 and 6 in an acidic aqueous medium gave the corresponding 2-deoxy-aldopyranoses  $2^{\ddagger}$ , 8 and 7 without allylic rearrangement (the Ferrier reaction).  $1^5$  The subsequent oxidation was effected by portionwise addition of pyridinium chlorochromate (PCC), whereas the use of Me<sub>2</sub>SO with P<sub>2</sub>O<sub>5</sub>, Ac<sub>2</sub>O or (CF<sub>3</sub>CO)<sub>2</sub>O was

Scheme 1 Reagents and conditions: i, THF–H<sub>2</sub>O–conc. HCl (5:1:0.1), room temp. overnight, 85% 2 and 88% 7; ii, PCC (3 × 1 equiv. at 2 h intervals), 4 Å molecular sieves CH<sub>2</sub>Cl<sub>2</sub>, room temp. 5 h, 85% 3 and 90% 8; iii, KHMDS (1.1 equiv.), THF, -90 °C, 15 min; iv, trisyl azide (1.2 equiv.), 2 min; v, AcOH (1.2 equiv.), -90 °C  $\rightarrow$  room temp.; vi, DIBAL-H (2 equiv.), THF, -70 °C, 30 min; vii, H<sub>2</sub>O, -70 °C  $\rightarrow$  room temp. viii, 6 mol dm<sup>-3</sup> HCl (a few drops), 15 min, 80% 5 from 3 and 65% 10 from 8.

not satisfactory. In contrast to the previous report, <sup>12</sup> the direct oxidation of glycals to lactones was accompanied by  $\beta$ -elimination leading to the formation of  $\alpha$ ,  $\beta$ -unsaturated lactones. Based on the <sup>1</sup>H NMR data ( $J_{2ax,3} = J_{2eq,3} = J_{3,4} = 4.5$  Hz and NOE between H-2ax and H-5), the lactone **8**§ seems to adopt a  $B_{2,5}$  conformation rather than  ${}^4C_1$  as in the case for **3** (Scheme 2).

Electrophilic azidation of 3/8 was carried out according to the procedure reported by Evans and Britton, <sup>11</sup> *i.e.* enolization with potassium bis(trimethylsilyl)amide (KHMDS), triazine formation with triisopropylphenylsulfonyl azide (trisyl azide)<sup>16</sup> and quenching the reaction with AcOH. Since the azidolactones 4\sqrt{s} and 9\sqrt{s} decomposed slowly during work-up, the azidation was followed by selective reduction of lactone to lactol with diisobutylaluminium hydride (DIBAL-H) in the same pot, furnishing 2-azido-2-deoxy-aldopyranoses 5 and 10\sqrt{s} in 80 and 65\% overall yields, respectively. For the analysis, small amounts of 4 and 9 could be isolated by flash column chromatography on silica gel (6:1 hexane–EtOAc).

A general procedure for the one-pot reaction  $(3 \rightarrow 4 \rightarrow 5 \text{ and } 8 \rightarrow 9 \rightarrow 10)$  is as follows: a solution of 3/8 in anhydrous THF was cooled to -90 °C and a 0.5 mol dm<sup>-3</sup> solution of KHMDS (1.1 equiv.) was added dropwise with vigorous stirring. After 15 min, a precooled 0.2 mol dm<sup>-3</sup> solution of trisyl azide in THF (1.2 equiv., -90 °C) was added dropwise. After another 2 min, AcOH (1.2 equiv.) was added and the mixture was warmed gradually to room temp. and stirred for 15 min. The mixture was again cooled to -70 °C and precooled DIBAL-H (2 equiv.) was added. After 30 min, H<sub>2</sub>O was added and the mixture was warmed to room temp. The mixture was then acidified by a few drops of 6 mol dm<sup>-3</sup> HCl and stirred for 15 min. The product was extracted with CH<sub>2</sub>Cl<sub>2</sub> and purified by flash column chromatography on silica gel (20:1 toluene–acetone).

The high stereochemistry of this azidation reaction, is comparable to (for the *lyxo*-series, *i.e.* 3) or better than (for the *arabino*-series, *i.e.* 8) those observed in the known azide additions to glycals<sup>4-6</sup> mentioned above. For the azidation of 8, the production of another diastereoisomer, the 2-azido-2-deoxy-glucopyranose derivative, was not detected. It seems that the azidation yields preferentially an equatorial azido group. Therefore, the electrophilic azidation to 2-deoxy-aldono-1,5-lactones provides a highly stereoselective alternative to the existing methods for preparing 2-azido-2-deoxy-aldopyranoses.

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$$\begin{array}{c} \text{BnO} \\ \text{BnO} \\ \text{H} \\ \text{O} \\ \text{BnO} \\$$

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## **Footnotes**

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- ‡ All new compounds exhibited satisfactory spectral and high-resolution MS data.

§ Selected physical properties of compounds. 2: colourless oil, 3.5:1 (α:β); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 5.45 (br d, 1 H, J 4.0 Hz, H-1α), 4.13 (ddd, 1H, J 5.5, 5.5, ca. 0 Hz, H-5α), 3.98 (ddd, 1 H, J 11.0, 4.5, 3.0 Hz,  $H-3\alpha$ ), 3.87 (br s, 1 H,  $H-4\alpha$ ), 3.81 (br s, 1 H,  $H-4\beta$ ), 3.63 (dd, 1 H, J 9.5, 6.0 Hz, H-6β), 3.58 (dd, 1 H, J 9.5, 5.5 Hz) and 3.50 (dd, 1 H, J 9.5, 5.5 Hz) (2 × H-6 $\alpha$ ), 2.21 (ddd, 1 H, J 12.0, 11.0, 4.0 Hz,  $\text{H-2ax}\alpha$ ), 2.15 (br d, 1 H, J 12 Hz, H-2eq $\beta$ ) and 2.01 (ddd, 1 H, J 12.0, 4.5, ca. 0 Hz, H-2eqα). HRFABMS: 457.2001 (C<sub>27</sub>H<sub>30</sub>NaO<sub>5</sub> [M + Na]<sup>+</sup>, calc. 457.1991). **4**: colourless oil,  $[\alpha]_D$  +63 (*c* 0.6, CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 4.59 (d, 1 H, J 10.5 Hz, H-2), 4.31 (ddd, 1 H, J 9.0, 6.0, 2.0 Hz, H-5), 4.15 (br s, 1 H, H-4), 3.70 (dd, 1 H, J 10.0, 9.0 Hz) and 3.65 (dd, 1 H, J 10.0, 6.0 Hz) (2 × H-6) and 3.67 (dd, 1 H, J 10.5, 2.0 Hz, H-3). HRFABMS: 496.1839 (C<sub>27</sub>H<sub>27</sub>N<sub>3</sub>NaO<sub>5</sub> [M + Na]+, calc. 496.1848). **8**: colourless crystals, mp 79 °C, [α]<sub>D</sub> +37 (c 0.6, CHCl<sub>3</sub>) {lit.  $^{13}$  mp 83 °C, [ $\alpha$ ]<sub>D</sub> +48 (c 1.0, EtOH)};  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  4.30 (ddd, 1 H, J 7.5, 4.0, 4.0 Hz, H-5), 3.94 (ddd, 1 H, J 4.5, 4.5, 4.5 Hz, H-3), 3.89 (dd, 1 H, J 7.5, 4.5 Hz, H-4), 3.73 (dd, 1 H, J 10.5, 4.0 Hz) and 3.70 (dd, 1 H, J 10.5, 4.0 Hz) (2 × H-6), 2.84 (dd, 1 H, J 15.0, 4.5 Hz, H-2ax) and 2.74 (dd, 1 H, J 15.0, 4.5 Hz, H-2eq). 9: colourless oil, [ $\alpha$ ]<sub>D</sub> +6 (c 0.06, CHCl<sub>3</sub>); <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$ 4.33 (m, 1 H, H-5), 4.14 (d, 1 H, J 3.5 Hz, H-2), 4.05 (dd, 1 H, J 3.5, 1.5 Hz, H-3), 3.90 (dd, 1 H, J 6.0, 1.5 Hz, H-4) and 3.66 (d, 2 H, J 4.5 Hz, 2 × H-6). HRFABMS: 496.1848 ( $C_{27}H_{27}N_3NaO_5$  [M + Na]+ calc. 496.1848). **10**: colourless oil, 3.5:1 ( $\alpha:\beta$ ); <sup>1</sup>H NMR (500 MHz,  $CDCl_3)$   $\delta$  5.19 (br s, 1 H, H-1 $\alpha$ ), 4.69 (br s, 1 H, H-1 $\beta$ ), 4.10 (dd, 1 H, J 9.0, 4.0 Hz, H-3 $\alpha$ ), 3.99 (ddd, 1 H, J 9.5, 5.0, 2.5 Hz, H-5 $\alpha$ ), 3.93 (m, 1 H, H-2 $\beta$ ), 3.92 (dd, 1 H, J 4.0, 2.5 Hz, H-2 $\alpha$ ), 3.82 (dd, 1 H, J 9.5, 9.5 Hz, H-4 $\beta$ ), 3.78 (dd, 1 H, J 9.0, 9.0 Hz, H-4 $\alpha$ ), 3.71 (dd, 1 H, J 9.0, 3.5 Hz, H-3 $\beta$ ), 3.69 (m, 2 H, 2 × H-6 $\beta$ ) and 3.65 (m, 2 H, 2 × H-6 $\alpha$ ). HRFABMS:  $498.2010 (C_{27}H_{29}N_3NaO_5 [M + Na]^+ calc. 498.2005)$ .

¶ On the basis of the  $^1$ H NMR data, the azidolactones **4** and **9** seem to exist similarly to **3** and **8** as  $^4C_1$  and  $B_{2,5}$  conformations, respectively. NOE between H-2 and H-5 observed in **9** implies the  $B_{2,5}$  conformation.

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