

A Stereoselective and Preparative Entry to 1,2-Anhydrosugars through Oxidation of Glycals with Perfluoro-*cis*-2,3-dialkyloxaziridines

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Perfluoro-*cis*-2,3-dialkyloxaziridines **1** perform the direct epoxidation of glycals **2** to give cleanly corresponding 1,2-anhydrosugars **3** with medium to complete diastereoselection; elaboration of these glycals to glycosyl fluorides and lipid conjugates is also reported.

1,2-Anhydrosugars **3** have good glycosyl donor properties, probably due to the ring strain in the alkoxy-epoxide moiety. While they have been used for the synthesis of glycosyl fluorides and sulfides^{1,2} and, more interestingly, in a reiterative strategy to α - or β -linked oligosaccharides and glyco conjugates,³⁻⁵ the full synthetic potential of these compounds has not been exploited, probably owing to a lack of broad availability of such oxiranes. In fact, direct epoxidation of glycals **2** with peracids affords products deriving from the reaction of initially produced 1,2-anhydrosugars with either solvent or co-formed acids.⁶

Perfluoro-*cis*-2,3-dialkyloxaziridines **1** are powerful, yet selective oxidizing agents which can work in aprotic and non-nucleophilic solvents.⁷⁻¹¹ The co-product of the oxidation reaction are perfluoroalkenes which are inert and volatile materials.^{12,13} For these reasons we thought that oxaziridines **1** could not only perform the epoxidation of glycals **2** to 1,2-anhydrosugars **3**, but also allow these latter products to be generated in a non-reactive medium. This is a prerequisite for the conjugation of the 1,2-anhydrosugar with a defined nucleophile. Here we report our preliminary results on this interesting epoxidation reaction.

When tri-*O*-acetyl-D-glucal **2a** was reacted with an equimolar amount of perfluoro-1-*n*-butyl-3-*n*-propyloxaziridine **1a** at room temperature and in trichlorofluoromethane solution a rapid reaction occurred to afford corresponding 1,2-anhydro derivatives **3a** and **4a** in nearly pure form and as a 1:1 mixture and in nearly quantitative yields (¹H NMR analyses of crude reaction mixture, Scheme 1). The stereoselectivity of the process and the chemical yields did not change substantially at lower (-78 °C) or higher temperatures (80 °C), but better results were obtained by using other solvents (CHCl₃, d.e. 18%; CCl₄, d.e. 30%; CFCl₂CF₂Cl, d.e. 42%). Perfluoro-1-*n*-pentyl-3-*n*-hexyloxaziridine **1b**, while showing a similar behaviour, allowed slightly higher diastereoselectivities to be obtained (e.g. CFCl₂CF₂Cl, d.e. 50%). In all cases chemical yields remained high and the prevailing diastereoisomer had the 1,2-anhydro- α -D-glucopyranose structure **3a**.[†]

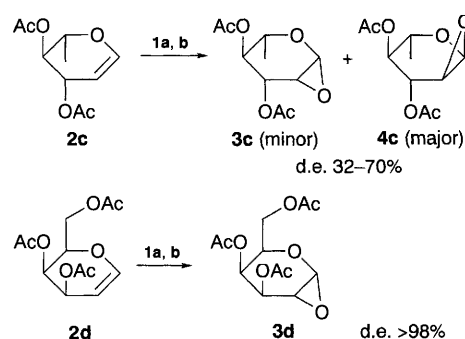
Best results were obtained by changing the protective groups at the hydroxy residues in the starting glucal, i.e. by using the tri-*O*-benzyl-D-glucal **2b** (**1a**, CHCl₃, d.e. 68%; **1b**, CHCl₃, d.e. 75%).

The epoxidation reaction had been performed successfully also on the di-*O*-acetyl-L-rhamninal **2c** (Scheme 2). The stereochemical preference paralleled strictly that observed on glucal **2a** as was expected given the fact that the two compounds

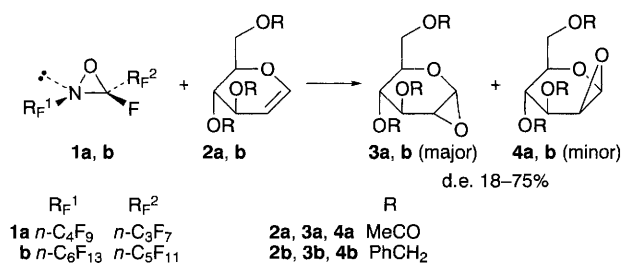
have the same relative configurations. The epoxide **4c**, in which the oxirane ring is *trans* to the ring appendages at C-3 and C-5 of the sugar moiety, was in fact formed preferentially (d.e. 32-70%) and once again ¹H NMR of the crude reaction mixture showed that nearly quantitative yields were obtained. Finally, on epoxidation of tri-*O*-acetyl-D-galactal **2d**, the same diastereofacial preference was observed, but the process occurred with complete stereoselection using both **1a** and **1b** and the 1,2-anhydro-3,4,6-tri-*O*-acetyl- α -D-galactopyranose **3d** was obtained exclusively (yields > 90%).

Spectroscopic properties of anhydrosugars **3a-d** and **4a-c** did not enable a direct assignment of their configuration to be established.² For this reason and as a first exploitation of their effectiveness as glycosyl donors, epoxides **3** were opened with fluorine and oxygen nucleophiles to give corresponding glycosyl fluorides^{14,15} **5** and lipid conjugates **6**. For instance, when the crude anhydrosugar **3d** was reacted with NBu₄F¹⁶ (TBAF, Aldrich, THF solution) a single fluoride **5d** was formed which possessed spectroscopic parameters diagnostic of the configuration shown at C-1 and C-2.¹⁷ The α -D-galactopyranosyl configuration was assigned to this compound and consequently to the precursor anhydrosugar **3d**.[‡] Similarly, the configuration of epoxides **3a** and **4a** was established from the fact that on treatment with TBAF they afforded two pyranosyl fluorides **5a** and **5b** having α -D-*gluco* and α -D-*manno* configurations, respectively.

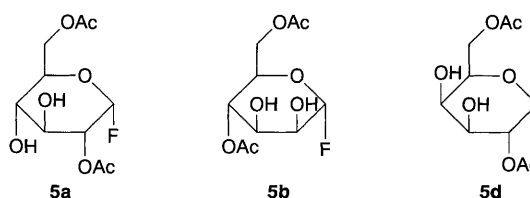
Furthermore, reaction of crude epoxides obtained from glucal **2a** with cholesterol in the presence of zinc(II) chloride afforded cholesteryl β -D-*gluco*-pyranoside **6a** and α -D-*manno*-pyranoside **6b** in 62% isolated yield.[§] Similarly, when (*S*)-citronellol was used, the corresponding glyco conjugates **6c** and **6d**, having the β -D-*gluco* and α -D-*manno* configurations, respectively, were formed.[¶]

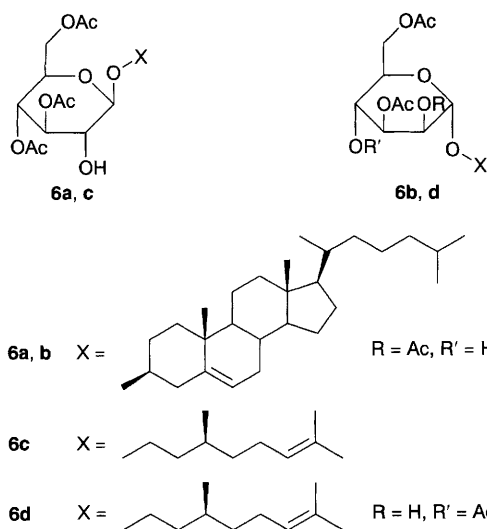


Scheme 2



Scheme 1





In conclusion, a new and preparative approach to 1,2-anhydrosugars is described. A stereospecific elaboration to give pyranosyl fluorides and glyco conjugates is also reported thus showing that the anhydrosugars produced through our methodology are pure enough to undergo subsequent conjugation with a defined nucleophile. Other reactions to afford interesting disaccharides are under study.

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Footnotes

† In a standard reaction, a solution of the oxaziridine **1** in the halogenated solvent was added dropwise to a solution of the glycol **2** in the same solvent at room temperature. After 5 min, the reaction mixture was washed with perfluorotri-*n*-butyl amine, the co-formed perfluoroazaalkene was removed and a solution of nearly pure anhydrosugar **3** was obtained. Diastereoisomer ratios were established through ^1H NMR of crude reaction mixtures. *Selected spectral properties:* 1,2-anhydro-3,4,6-tri-*O*-acetyl- α -D-glucopyranose **3a**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.23 (dd, 1H, J 1.2 and 8.2 Hz, H-3), 5.03 (dd, 1H, J 10.4 and 8.2 Hz, H-4), 5.02 (d, 1H, J 2.2 Hz, H-1), 4.32 (dd, 1H, 3J 4.1 Hz, J_{gem} 12.5 Hz, H-6), 4.07 (dd, 1H, 3J 2.4 Hz, J_{gem} 12.5 Hz, H-6'), 3.98 (ddd, 1H, J 4.1, 2.4 and 10.4 Hz, H-5), 3.01 (d, 1H, J 2.2 Hz, H-2), 2.086, 2.069 and 2.027 (s, 3H each, CH_3CO); 1,2-anhydro-3,4,6-tri-*O*-acetyl- β -D-manno-pyranose **4a**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.37–5.28 (m, 2H, H-3 and H-4), 5.00 (d, 1H, J 2.8 Hz, H-1), 4.19 (dd, 1H, 3J 4.6 Hz, J_{gem} 12.5 Hz, H-6), 4.10 (dd, 1H, 3J 2.4 Hz, J_{gem} 12.5 Hz, H-6'), 3.94 (ddd, 1H, J 4.6, 2.4 and 7.0 Hz, H-5), 3.44 (dd, 1H, J 2.8 and 2.1 Hz, H-2), 2.094, 2.075 and 2.022 (s, 3H each, CH_3CO); 1,2-anhydro-6-deoxy-3,4-di-*O*-acetyl- β -L-manno-pyranose **3c**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.15 (dd, 1H, J 1.2 and 8.3 Hz, H-3), 4.90 (br s, 1H, H-1), 4.75 (dd, 1H, J 10.2 and 8.3 Hz, H-4), 3.85 (dd, 1H, J 10.2 and 6.3 Hz, H-5), 2.97 (d, 1H, J 2.3 Hz, H-2), 2.044 and 1.982 (s, 3H each, CH_3CO), 1.14 (d, 3H, J 6.3 Hz, Me); the acetyl groups gave two singlets at δ 2.07 and 2.04 and were not assigned; 1,2-anhydro-6-deoxy-3,4-di-*O*-acetyl- α -L-glucopyranose **4c**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.24 (dd, 1H, J 2.3 and 8.6 Hz, H-3), 5.09 (dd, 1H, J 10.2 and 8.6 Hz, H-4), 4.91 (br s, 1H, H-1), 3.81 (dd, 1H, J 10.2 and 6.3 Hz, H-5), 3.41 (dd, 1H, J 2.4 and 2.4 Hz, H-2), 2.070 and 2.037 (s, 3H each, CH_3CO), 1.15 (d, 3H, J 6.3 Hz, Me); 1,2-anhydro-3,4,6-tri-*O*-acetyl- α -D-galactopyranose **3d**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.22 (dd, 1H, J 1.2 and 4.3 Hz, H-3), 5.17 (dd, 1H, J 1.0 and 4.3 Hz, H-4), 5.03 (d, 1H, J 2.4 Hz, H-1), 4.12–3.99 (m, 3H, AMX system, H-5,

H-6 and H-6' strongly coupled), 2.98 (dd, 1H, J 2.4 and 1.2 Hz, H-2), 2.197, 2.054 and 2.048 (s, 3H each, CH_3CO).

‡ Values of fluorine chemical shift ($\delta_{\text{F}} - 150.8$) and heteronuclear coupling constants ($^3J_{\text{F-H-2}} = 24.8$ Hz) show that F and H-2 are in a *trans*-diaxial relationship; furthermore the homonuclear coupling constants $^3J_{\text{H-1-H-2}} = 2.7$ Hz and $^3J_{\text{H-2-H-3}} = 9.5$ Hz are consistent with H-1 equatorial and H-2 and H-3 axial in a six-membered $^4C_1(\text{D})$ conformation. These data are not consistent with the alternative structure of an α -*tal*o-pyranosyl fluoride and consequently the β -*tal*o-pyranose structure was ruled out for the precursor anhydrosugar. The stereochemistry of **3b,c** and **4b,c** was assigned through spectral correlation with **3a,d** and **4a**. The TBAF ring-opening of **3a**, **4a** and **3d** was accompanied by the release of one acetate group yielding the corresponding di-*O*-acetyl derivatives. Scrambling of acetyl groups was also observed.

§ Minor amounts (18% yield) of other cholesteryl pyranosides having the β -D-*gluco* and α -D-*manno* configuration and formed by scrambling of the acetyl residues were also obtained. *Selected spectral properties:* cholesteryl 3,4,6-tri-*O*-acetyl- β -D-*gluco*-pyranoside **6a**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.36 (ddd, 1H, J 1.7, 1.7 and 5.2 Hz, H-6), 5.11 (dd, 1H, J 9.3 and 9.3 Hz, H-3'), 5.00 (dd, 1H, J 9.3 and 9.9 Hz, H-4'), 4.45 (d, 1H, J 7.8 Hz, H-1'), 4.26 (dd, 1H, J 5.1 and 12.2 Hz, H-6'), 4.08 (dd, 1H, J 2.6 and 12.2 Hz, H-6''), 3.67 (ddd, 1H, J 2.6, 5.1 and 12.2 Hz, H-5'), 3.57 (m, 1H, H-3), 3.53 (dd, 1H, J 7.8 and 9.4 Hz, H-2'), 2.065, 2.058 and 2.014 (s, 3H each, CH_3CO). Cholesteryl 2,3,6-tri-*O*-acetyl- α -D-*manno*-pyranoside **6b**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.37–5.33 (m, 1H, H-6), 5.23 (dd, 1H, J 3.4 and 9.8 Hz, H-3'), 5.17 (dd, 1H, J 1.8 and 3.4 Hz, H-2'), 4.94 (d, 1H, J 1.8 Hz, H-1'), 4.50 (dd, 1H, J 5.0 and 12.2 Hz, H-6'), 4.33 (dd, 1H, J 2.2 and 12.2 Hz, H-6'') 3.96 (ddd, 1H, J 2.2, 5.0 and 9.9 Hz, H-5'), 3.81 (dd, 1H, J 9.9 and 9.9 Hz, H-4'), 3.53–3.43 (m, 1H, H-3), 2.124, 2.114 and 2.076 (s, 3H each, CH_3CO).

¶ *Selected spectral properties:* citronellyl 3,4,6-tri-*O*-acetyl- β -D-*gluco*-pyranoside **6c**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.10 (ddd, 1H, J 9.4 and 9.4 Hz, H-3'), 5.07 (m, 1H, H-6), 5.00 (dd, 1H, J 9.4 and 9.8 Hz, H-4'), 4.33 (d, 1H, J 7.8 Hz, H-1'), 4.26 (dd, 1H, J 4.9 and 12.2 Hz, H-6'), 4.10 (dd, 1H, J 2.5 and 12.2 Hz, H-6''), 3.94 (ddd, 1H, J 5.7, 7.8 and 9.6 Hz, H-1a), 3.67 (ddd, 1H, J 2.5, 4.9 and 9.9 Hz, H-5'), 3.57 (ddd, 1H, J 6.9, 7.5 and 9.6 Hz, H-1b) 3.54 (dd, 1H, J 7.8 and 9.4 Hz, H-2'), 2.057, 2.055 and 2.009 (s, 3H each, CH_3CO). Citronellyl 3,4,6-tri-*O*-acetyl- α -D-*manno*-pyranoside **6d**: ^1H NMR (CDCl_3 , 400 MHz, 303 K) δ 5.32 (dd, 1H, J 9.8 and 9.8 Hz, H-4'), 5.26 (dd, 1H, J 9.8 and 2.9 Hz, H-3'), 5.10 (m, 1H, H-6), 4.86 (d, 1H, J 2.0 Hz, H-1'), 4.27 (dd, 1H, J 5.2 and 12.2 Hz, H-6'), 4.12 (dd, 1H, J 2.3 and 12.2 Hz, H-6''), 4.02 (m, 1H, H-2'), 3.97 (ddd, 1H, J 2.3, 5.2 and 9.8 Hz, H-5'), 3.73 (ddd, 1H, J 7.4, 7.4 and 9.7 Hz, H-1a), 3.51 (ddd, 1H, J 5.8, 7.4 and 9.7 Hz, H-1b), 2.087, 2.082 and 2.034 (s, 3H each, CH_3CO).

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