Two-step synthesis of C-glycosyl juglones from unprotected sugars: a novel approach to angucycline antibiotics

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The two-step synthesis of C-glycosyl juglones, versatile synthetic intermediates for angucycline antibiotics, has been developed by the C-glycosidation of naphthalene-1,5-diol with an unprotected sugar and the subsequent regioselective photooxygenation of the resultant C-glycosyl naphthalenediol.

C-Glycosyl juglone, a 5-hydroxy-1,4-naphthoquinone bearing a C-glycoside at C6, is a common structural feature among the many members of angucycline antibiotics such as aquayamycin 1 and urdamycinone B 2 which show a variety of biological activity including antitumour activity.1 C-Glycosyl juglone is also a promising synthetic intermediate for this class of antibiotics. Therefore, several approaches to C-glycosyl juglone have been developed² and the syntheses of angucycline antibiotics via the C-glycosyl juglone have also been reported.^{3,4} Herein we report a novel and efficient approach to Cglycosyl juglones in two steps involving the C-glycosidation of naphthalene-1,5-diol with an unprotected sugar to form the aryl C-glycosidic linkage and the subsequent regioselective photooxygenation of the resultant C-glycosyl naphthalenediol to construct the juglone skeleton (Scheme 1). To the best of our knowledge, this is the first protocol for the synthesis of Cglycosyl juglones without any protecting groups on the aglycon or glycon moieties.

The first step is the C-glycosidations⁵ of naphthalene-1,5-diol **3** with several unprotected sugars **4–7**. The results are summarized in Table 1. D-Olivose **4** is the most representative sugar that appeared in many angucycline antibiotics. The C-glycosidation of **3** (2.0 equiv.) and **4** (1.0 equiv.) using trimethylsilyl trifluoromethanesulfonate (Me₃SiOTf) (0.2 equiv.)⁶ in MeCN at 25 °C for 1 h proceeded smoothly to give the unprotected aryl β -C-glycoside **8**[†] in 65% yield as a single isomer (entry 1 in Table 1). Unprotected methyl D-olivoside **5** was also coupled with **3** under similar conditions to afford **8** in good yield (entry 2 in Table 1). To enhance the synthetic utility of this reaction, we further examined the C-glycosidations of other unprotected 2,6-dideoxy sugars **6** and **7** with **3** mediated



Urdamycinone B 2

by a catalytic amount of Me₃SiOTf. It was found that these Cglycosidations also proceeded easily to furnish the unprotected aryl β -C-glycosides **9**[†] and **10**[†] in 61 and 65% yields, respectively (entries 3 and 4 in Table 1).

With the unprotected C-glycosyl naphthalenediols in hand, the second step, the oxygenation to the C-glycosyl juglones, was carried out. We first examined the oxygenation of **8** using ammonium cerium(IV) nitrate (CAN),⁷ potassium nitrosodisulfonate (Fremy's salt)⁸ and thallium(III) nitrate trihydrate (TTN)⁹ as the oxygenating agents, which are widely used for



Scheme 1



Table 1 C-Glycosidations of 3 and the unprotected sugars 4–7 to give the β -C-glycosides 8–10^a

Entry	Sugar	Product	Yield (%)
1	4	8	65
2	5	8	64
3	6	9	61
4	7	10	65

^a All reactions were carried out using 2.0 equiv. of **3** with respect to the sugars.

conversion of naphthol to quinone. However, in these cases, the desired C-glycosyl juglone 11‡ was either not detected at all or isolated in a very low yield while the regioisomer 14‡ was predominantly produced as shown in Table 2. Therefore, our attention next turned to the photooxygenation¹⁰ of 8. These results are summarized in Table 3. Remarkably, the regioselectivity of the oxygenation of 8 was dramatically changed and the desired C-glycosyl juglone 11 was predominantly obtained by choice of the appropriate reaction solvent. Thus, the photooxygenation of 8, without any reagent, was best effected by irradiating it with sunlight in Bu^oOH–CHCl₃ (1:3, 0.0069 M



12 $R^2 = R^4 = H, R^1 = R^3 = OH$ **13** $R^1 = R^3 = H, R^2 = R^4 = OH$

15 $R^2 = R^4 = H$, $R^1 = R^3 = OH$ **16** $R^1 = R^3 = H$, $R^2 = R^4 = OH$

Table 2 Oxygenations of 8 at 25 °C to give the C-glycosyl juglones 11 and 14

Entry	Reagent (equiv.)	Solvent	t/h	Yield (%)	
				11	14
1	CAN (2.5)	MeCN-H ₂ O	0.5	0	25
2	Fremy's salt (3.6)	MeOH	1	10	55
3	TTN (1.8)	MeOH	0.5	1	74

Table 3 Photooxy genations of 8–10 to give the C-glycosyl juglones 11-16

Entry	C-Glycosyl naphthalenediol	Solvent ^a		C-Glycosyl juglone/ Yield (%)				
1	8	MeOH	11	7	14	12		
2	8	EtOH	11	19	14	17		
3	8	Pr ⁱ OH	11	30	14	17		
4	8	Bu'OH	11	52	14	22		
5	8	ButOH-hexane(1:1)	11	41	14	13		
6	8	Bu ⁱ OH–PhH(1:1)	11	51	14	15		
7	8	Bu ^t OH–CHCl ₃ (1:1)	11	52	14	14		
8	8	$Bu^{t}OH-CHCl_{3}(1:3)$	11	57	14	13		
9	9	Bu ^t OH–CHCl ₃ (1:1)	12	56	15	17		
10	10	Bu ^t OH–CHCl ₃ (1:1)	13	55	16	18		

^а 0.0069 м for C-glycosyl naphthalenediol.

for 8) under O_2 at room temperature for 12 h to give the Cglycosyl juglone 11 in 57% yield along with a 13% yield of 14 (entry 8 in Table 3). Other C-glycosyl naphthalenediols 9 and 10 were also found to cleanly convert under similar conditions to the C-glycosyl juglones 12 and 13 in 56 and 55% yields, respectively (entries 9 and 10 in Table 3).

À typical experimental procedure for the preparation of **8** via C-glycosidation of **3** and **4**, followed by photoxygenation to give **11**, is as follows. To a mixture of **3** (183.8 mg, 1.148 mmol) and **4** (85.0 mg, 0.574 mmol) in dry MeCN (5.74 ml) was added Me₃SiOTf (0.022 ml, 0.115 mmol) under ice-cooling conditions. After stirring for 1 h at 25 °C, the reaction was quenched with Et₃N and the resulting mixture was then concentrated *in vacuo*. Purification of the residue by flash column chromatography with 10:1 chloroform–methanol gave **8** (108.8 mg, 05.3%, β only) as a white solid. A solution of **8** (108.8 mg, 0.375 mmol) in Bu'OH–CHCl₃ (3:1, 54.4 ml) was then irradiated with diffuse sunlight (a 75 W xenon lamp, Wacom Sunray Lamp, I-Sunsun) under O₂ for 12 h and then concentrated *in vacuo*. Purification of the residue by flash column chromatography with 2:1 hexane–acetone gave **11** (64.4 mg, 56.5%) and **14** (14.8 mg, 13.0%) as an orange solid.

Footnotes

† The β -configuration of C-glycoside was evident from the ¹H NMR spectra.

‡ Structure confirmation relies on the NOE experiments.

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