Mechanism of oxygenation of vitamin K hydroquinone

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When the monoanion of vitamin K hydroquinone is treated with ¹⁸O₂ in THF, the product, vitamin K oxide, carries a full **atom of 18O at the epoxide oxygen and partial incorporation (34%) of a second atom of 18O at the carbonyl oxygen, which is most reasonably ascribed to the dioxetane mechanism.**

Vitamin K in its hydroquinone form, vitamin $KH₂$, is required as a cofactor for a microsomal enzyme that converts N-terminal glutamates in proteins of the blood clotting cascade to γ -carboxyglutamates.¹ Vitamin KH₂ is transformed into vitamin K oxide concurrent with the abstraction of the γ -hydrogen of glutamate, leading to γ -carboxyglutamate.² In studies of the mechanism of action of vitamin K, the role of molecular oxygen in the formation of vitamin K oxide was explored.^{3,4} Key features of this mechanism are the rearrangement to a strong alkoxide base **5** *via* the dioxetane intermediate **4** (Scheme 1), proposed on the basis of results of a parallel oxygen-16 and oxygen-18 experiment,3 and the corresponding oxidation of a model system.4

The nonenzymic model approach that supports the chemical principle used 2,4-dimethylnaphthol in place of vitamin KH_{2} .⁴ However, the most important difference between the model and the vitamin K dependent carboxylase resides in the structure of vitamin K. Vitamin K is a $1,4$ -dioxygenated naphthalene, whereas the model carries only one oxygen. Since molecular oxygen can add to the 2-position of vitamin KH_{2} 1 yielding a hydroperoxy anion **7** which might produce vitamin K oxide **6**, 5 it was interesting to compared the *in vivo* oxygenation reaction of vitamin K hydroquinone itself, in the absence of the enzyme.

It is well-established that hydroquinones are unstable in air and rapidly oxidize to yield quinone. Indeed, when a chloroform solution of vitamin K hydroquinone **1** was stirred under an oxygen atmosphere, vitamin K was formed within 30 min. By contrast, when the monoanion of vitamin KH_{2} was stirred under an oxygen atmosphere, vitamin K oxide **6** was isolated in 88% yield (Scheme 2). These spontaneous oxidations in the chemical model system are clearly important for the mechanism

Scheme 1

of action of vitamin KH_2 in biological systems. Therefore, experiments employing oxygen-18 were next used to examine the involvement of the dioxetane intermediate, as shown in Scheme 1, because 2-hydroperoxide **8** could be differentiated from the dioxetane mechanism by 18O labelling experiments.

Typical experimental conditions consisted of stirring a THF solution of a monoanion of vitamin KH₂ 2 at room temperature with 18-crown-6 and delivering ${}^{18}O_2$ from a sealed tube to yield vitamin K oxide **6**.† Examination of the mass spectrum reveals that an atom of 18O has been incorporated into the vitamin K oxide **6**, since the molecular ion peak moves from *m/z* 466 to 468. The 18O label could, in principle, be located at any of the three oxygen positions, but the mass spectrum is most consistent with location of the ¹⁸O at the epoxide position. Thus, the m/z 423 peak is unchanged when comparing the unlabelled with the labelled spectrum; the $M^+ - 43$ peak at m/z 423 arises from cleavage of the epoxide and loss of a MeCO fragment.3 Moreover, the intensity of the peak at *m/z* 470 was 41% of the parent peak at *m/z* 468, which is much larger than would be expected on the basis of natural abundance 13C.6 After correcting for natural abundance 13C, the presence of 36% of a second atom of ¹⁸O in the vitamin K oxide from the labelling experiment is indicated, and this is most reasonably ascribed to

the dioxetane mechanism (Scheme 3).
In EtOH, the product of oxygenation of vitamin $KH-2$ under ¹⁸O₂ resulted in a shift of the m/z 466 peak to 468, with no increase in the intensity of the M^+ + 4 peak. The path leading to

*Chem. Commun***., 1997 929**

the formation of vitamin K oxide in an aprotic solvent could be different from that in protic solvent. In the oxygenation of the vitamin K anion **2** in EtOH, the 2-peroxy anion intermediate **7** might predominate and might be protonated by EtOH, yielding the hydroperoxide **8**. If the phenolic hydroxy group of peroxide **7** is deprotonated by the resulting ethoxide, and with the assistance of a hydrogen bonding of solvent, the hydroperoxide could yield vitamin K oxide.

This solvent-dependent regioselectivity observed in the oxygenation of the vitamin K anion **2** might be rationalized in terms of the stability of the peroxy anion intermediates **3** and **7**. Since it is known that the potassium ion associates with crown ether more strongly than with protic solvents,7 the peroxy anion intermediate would be naked. The resulting free peroxy anion **7** at the 2-position might experience electronic repulsion by the carbonyl group, whereas the peroxy anion **3** at the 4-position is stabilized by hydrogen bonding with the *gem*-hydroxy group.8 therefore, the addition of oxygen using THF with 18-crown-6 might be preferred in the 4-position (Fig. 1).

The peroxy anion intermediate in EtOH seems to undergo stabilization by association with the counter cation K^+ and partly also by solvation through hydrogen bonding.7 In this case, the stabilization of the peroxy anion **7** at the 2-position appears to be due to chelation with the carbonyl group of the intermediate.

The biological transformation of vitamin K oxide **6** appears to be shielded from water by the enzyme or membrane environment.9 Therefore the chemical formation of vitamin K oxide **6** has parallels in the biological formation. However, another possibility for incorporation of 18O into vitamin K oxide **6** would involve label exchange between 18O labelled hydroxide and the product in THF, even though formation of hydroxide anion might be unfavourable under aprotic conditions. This path

Fig. 1 Proposed solvent-dependent regioselectivity of peroxide in the oxygenation of the vitamin K hydroquinone monoanion

cannot be ruled out at present, and the appropriate control reactions remain to be conducted.

This research was generously supported by a grant from the Ministry of Education (BSRI-96-3416). The authors also acknowledge partial support from Chung-Ang University.

Footnotes

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 \dagger Typical experiment: oxygenation of 2 with ${}^{18}O_2$. A solution of 142.8 mg (0.316 mmol) of vitamin K hydroquinone **1** in 4 ml of dry THF was added to a suspension of 12.6 mg (0.315 mmol) of potassium hydride and 1 ml of dry THF *via* a double tipped needle and stirred for 10 min under an atmosphere of argon at room temperature. 18-Crown-6 (60.0 mg, 0.227 mmol) was then added. ${}^{18}O_2$ [96% enriched in 18-oxygen (ICON), 100 ml] was added through a break seal. After stirring for 20 min at room temperature, the reaction mixture was quenched with 5 ml of water and extracted with diethyl ether (3×10 ml). The combined diethyl ether layers were washed with saturated aqueous potassium chloride, dried over MgSO₄ and concentrated to yield 101.8 mg of the crude product as a yellow oil, which was checked by GC–MS (Hewlett-Packard 5890 Series II gas chromatograph and a Hewlett-Packard 5970 mass selective detector).

References

- 1 A. F. Wagner and K. Folkers, *Vitamins and Coenzymes*, Interscience, New York, 1964, pp. 407–434.
- 2 J. W. Suttie, *Biofactors*, 1988, **1**, 55; J. W. Suttie, *Annu. Rev. Biochem.*, 1985, **54**, 459; R. E. Olson, *Annu. Rev. Nutr.*, 1984, **4**, 281; P. Dowd, R. Hershline, S. W. Ham and S. Naganathan, *Science*, 1995, **269**, 1684.
- 3 P. Dowd, S. W. Ham and S. J. Geibs, *J. Am. Chem. Soc.*, 1991, **113**, 7734; P. Dowd, S. W. Ham and R. Hershline, *J. Am. Chem. Soc.*, 1992, **114**, 7613.
- 4 S. W. Ham and P. Dowd, *J. Am. Chem. Soc.*, 1990, **112**, 1660.
- 5 K. Alder, F. H. Flock and H. Baumling, *Chem. Ber.*, 1960, **93**, 1896. Compound **8** is also proposed as a vitamin K intermediate for the basepromoted pathway; J. W. Suttie, A. E. Larson, L. M. Canfield and T. L. Carlisle, *Fed. Proc. Am. Soc. Exp. Biol.*, 1978, **37**, 2605.
- 6 The calculated ratio of the *m/z* 468, 469, 470 peaks is 100: 34.4: 6.3: J. H. Beynon, *Mass Spectrometry and its Applications to Organic Chemistry*, Elsevier, Amsterdam, 1960, p. 524.
- 7 R. Alexander and A. J. Parker, *J. Am. Chem. Soc.*, 1967, **89**, 5549.
- 8 A. Nishinaga, T. Itahara, T. Shimizu and T. Matsuura, *J. Am. Chem. Soc.*, 1978, **100**, 1820.
- 9 J. J. Mctigue and J. W. Suttie, *J. Biol. Chem.*, 1983, **258**, 12 129; D. A. Anton and P. A. Friedman, *J. Biol. Chem.*, 1983, **258**, 14 084.

Received in Cambridge, UK, 17th February 1997; Com. 7/01091E