

Regioselective oxyfunctionalization of peptides by dimethyldioxirane: tertiary C–H σ -bond oxygen atom insertion into leucine derivatives and leucine-containing dipeptides

Maurizio Mezzetti, Enrico Mincione* and Raffaele Saladino*†

Dipartimento Agrochimico Agrobiologico, Università degli studi di Viterbo 'La Tuscia', via San Camillo de Lellis, 01100 Viterbo, Italy

A simple and straightforward approach to selective C–H σ -bond oxygen atom insertion into Leu residues on peptides using dimethyldioxirane is described; this procedure is compatible with Gly, Ala, Val, Ile and Phe residues.

Proteins with altered functions can be prepared from existing molecules by genetic manipulations¹ or, particularly in the case of small peptides, by chemical methods, *via* side-chain alteration and backbone modifications. Several chemical methods for performing side-chain alterations have been reported. The tryptophan side-chain is efficiently transformed by Ru^{VIII}, generated *in situ* from RuCl₃ and NaIO₄, to that of aspartic acid.² A Ru^{VIII} reagent has subsequently been used to oxidize 13 to the 20 coded α -amino acids³ and for a novel protein backbone modification by C(α)–C side-chain scission.⁴ Moreover, selective modification of serine- and threonine-containing peptides has been accomplished with lead tetraacetate to give α -acyloxyglycine derivatives, which can be easily transformed into α -alkylthio- and α -halogeno-glycines.⁵ Recently, a simple approach to selective iodination of tyrosine residues on peptides using IPy₂BF₄ (Py = pyridine) has been described,⁶ and preliminary studies revealed that the method might be also suitable for iodination of phenylalanine derivatives.

In spite of extensive work on side-chain transformations of aromatic, heterocyclic and heteroatom moieties in peptides, no general methods have been described for oxygen atom insertion into the C–H σ -bond present in the side-chain of α -amino acids, with the exception of the glycine-selective chemical modification by Nickel peroxide,⁷ in which case a concomitant backbone modification was observed, probably because of the low reactivity generally shown by this bond.

A powerful and selective oxidant for this purpose,⁸ which performs under strictly neutral conditions, is dimethyldioxirane (DMD). The advantages of dioxiranes, especially DMD, in synthetic chemistry have been well documented in alkene epoxidations, heteroatom oxidations and C–H σ -bond insertions. To the best of our knowledge, DMD has been used in the chemistry of α -amino acids and peptides to perform the oxidation of α -diazo ketones derived from α -amino acids,⁹ and in the synthesis of enantiomerically pure N-protected β -amino- α -keto esters from α -amino acids and dipeptides.¹⁰ In each case, the possible oxygen atom insertion into the side-chain was not observed. We report here a simple, selective and straightforward approach to the C–H σ -bond oxygen atom insertion into Leucine (Leu) and Leu-containing dipeptides.

The oxidation of Boc-Leu-OMe **1** (1 mmol) performed with a freshly prepared solution of DMD (0.09 mol dm⁻³ acetone solution, 6.0 equiv.)¹¹ in CH₂Cl₂ (5 cm³) at 25 °C for 3 days afforded the 4,4-dimethyl-4-butanolide derivative **2** as the only recovered product in 42% isolated yield (Scheme 1),[‡] with some unreacted substrate. Compound **2** is probably formed by selective oxygen atom insertion into the tertiary C–H σ -bond of Leu followed by cyclization.

Some other protected α -amino acids bearing an alkyl side-chain (Boc-Gly-OMe, Boc-Ala-OMe, Boc-Val-OMe, Boc-Ile-

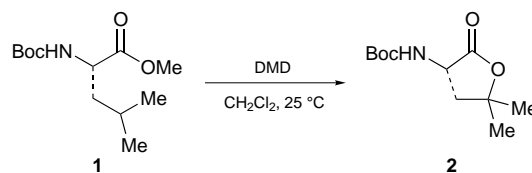
OMe and Boc-Phe-OMe) were then studied. The reactions performed with DMD (0.09 mol dm⁻³ acetone solution) in CH₂Cl₂ (5 cm³) at 25 °C for 3 days did not give products in appreciable amounts, even in the presence of a large excess of DMD and after longer reaction times.

It is known that the DMD oxygen atom insertion into the C–H σ -bond shows a pronounced tertiary–secondary–primary selectivity.⁸ Furthermore, this transformation is extremely sensitive to solvent¹² and stereoelectronic effects. In particular, a dipolar group close to the reactive centre may influence the reaction, favouring a chemo- and stereo-specific attack,¹³ or preventing the oxidation.¹⁴

We can put forward only hypotheses to explain the unexpected stability of the valine substrate in the DMD oxidation. A rationalization based on simple electronic deactivation by the amino functionality on the more proximate tertiary C–H σ -bond with respect to leucine seems reasonable. This hypothesis is in accord with the recently reported procedure¹⁵ for the selective oxyfunctionalization of unactivated C–H σ -bonds of alkylamines in acid medium by methyl(trifluoromethyl)dioxirane, in which the strong electron-withdrawing nature of the ammonium group deactivates the oxidation of even tertiary C–H σ -bonds at the α and β positions. In our case, the reaction appears to be operative only when a tertiary C–H σ -bond is in the remote C(γ) position with respect to the amino acid functionality.

The feasibility of transformation of **1** by DMD with respect to the other α -amino acids bearing an alkyl side-chain prompted us to evaluate the possibility to obtain a chemo- and regio-selective modification of Leu-containing dipeptides. The oxidation of Boc-Ala-Leu-OMe **3** and Boc-Leu-Ala-OEt **5** with an excess of DMD in CH₂Cl₂ (5 cm³) at 25 °C for 3 days afforded the butanolide derivate **4**§ and the peptide derivative **6**¶ in 35 and 38% isolated yield, respectively, plus some unreacted substrates (Scheme 2). Compound **4**, which is obtained by selective modification of the Leu residue, may be formed by selective oxygen atom insertion into the tertiary bond present in the side-chain of Leu, followed by cyclization. On the other hand, the presence of a free alcoholic moiety in **6** (as shown by the OH group absorption at 3390 cm⁻¹ in the IR spectrum) suggests that, in this case, a spontaneous cyclization process is not operative. This hypothesis was further confirmed by the absence of a detectable amount of the butanolide derivative **2** in the reaction mixture.

Finally, we studied the oxidation of the dipeptide Boc-Leu-Leu-OMe **7** by DMD under similar experimental conditions.¹⁶

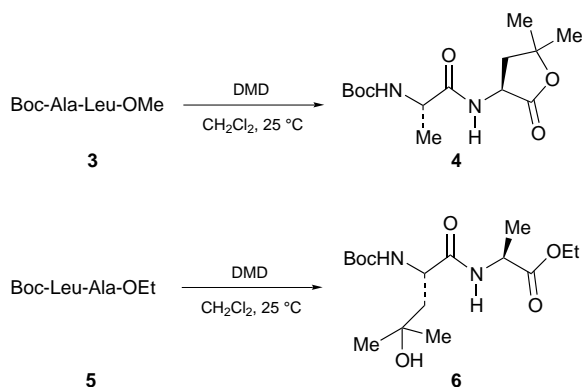


Scheme 1

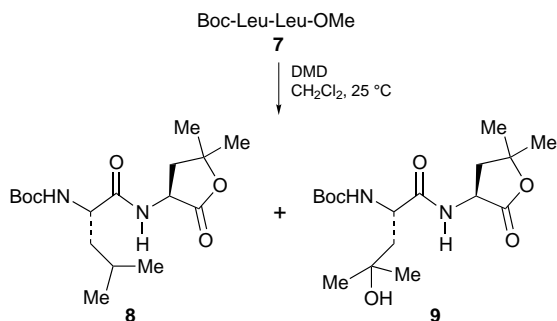
The oxidation of **7** with DMD (0.09 mol dm⁻³ acetone solution, 6.0 equiv.) in CH₂Cl₂ (5 cm³) at 25 °C for 3 days afforded the butanolide derivative **8** in 33% isolated yield (Scheme 3).|| Minor amounts of **9** were also formed, as shown by ¹H and ¹³C NMR spectral analyses of the crude reaction mixture prior to purification.

These data show that the reactivity of the Leu residue towards DMD is extremely sensitive to the position (C/N-terminal position) of the residue into the peptide. In particular, C-terminal Leu appears to be more reactive than the N-terminal residue towards DMD. In this case, steric factors may operate, since dioxiranes are generally sensitive to steric hindrance,⁸ even if electronic deactivation by the two amino acid functionalities of the Leu N-terminal residue cannot be ruled out.

The high selectivity observed for the C_γ-H σ-bond of Leu towards DMD offers a novel synthetic route to modified peptides without backbone modifications. Work is in progress in our laboratories to study the reactivity pattern of the other coded α-amino acids towards DMD oxidation, as well as to



Scheme 2



Scheme 3

transform the newly introduced hydroxy moieties into other useful functionalities.

Financial support from Italian MURST is acknowledged.

Footnotes

† E-mail: saladino@unitus.it

‡ Compound **2** was purified by flash-chromatography. MS analysis for **2** (ESI-MS) shows *m/z* 229 [M]⁺, in agreement with the proposed structure.

§ Compound **4** was purified by flash-chromatography. MS analysis for **4** (ESI-MS) shows *m/z* 299 [M + H]⁺, in agreement with the proposed structure.

¶ Compound **6** was purified by flash-chromatography. MS analysis for **6** (ESI-MS) shows *m/z* 334 [M]⁺, in agreement with the proposed structure.

|| Compound **8** was purified by flash-chromatography. MS analysis for **8** (ESI-MS) shows *m/z* 330 [M]⁺, in agreement with the proposed structure.

References

- M. Mutter, *Angew. Chem., Int. Ed. Engl.*, 1985, **24**, 639; M. Mutter and S. Vuilleumier, *Angew. Chem., Int. Ed. Engl.*, 1989, **28**, 535.
- S. Ranganathan, D. Ranganathan and D. Bhattacharyya, *J. Chem. Soc., Chem. Commun.*, 1987, 1085.
- S. Ranganathan, D. Ranganathan, D. Bhattacharyya, S. Bamezai, W. P. Singh, G. P. Battacharyya, G. P. Singh, R. K. Rathi, S. Saini, N. Jayaraman and B. K. Patel, *Pure Appl. Chem.*, 1990, **62**, 1433; S. Ranganathan, D. Ranganathan and D. Bhattacharyya, *Tetrahedron Lett.*, 1991, **32**, 5616.
- D. Ranganathan, N. K. Vaish and K. J. Shah, *J. Am. Chem. Soc.*, 1994, **116**, 6545.
- G. Apitz and W. Steglich, *Tetrahedron Lett.*, 1991, **27**, 3163.
- J. Barlunga, M. A. García-Martin, J. M. Gonzàles, P. Clapés and G. Valencia, *Chem. Commun.*, 1996, 1505.
- C. J. Easton, S. K. Eichinger and M. J. Pitt, *J. Chem. Soc., Chem. Commun.*, 1992, 1295.
- W. Adam, R. Curci and J. O. Edwards, *Acc. Chem. Res.*, 1989, **22**, 205; R. Curci, in *Advances in Oxygenated Processes*, ed. A. L. Baumstark, JAI Press, Greenwich, CT, 1990, vol. 2.
- W. Adam and L. P. Hadjarapoglou, *Top. Curr. Chem.*, 1993, **164**, 45.
- P. Darkins, N. McCarthy, T. McKervey and T. Ye, *J. Chem. Soc., Chem. Commun.*, 1993, 1222.
- W. Adam, J. Bialas and L. P. Hadjarapoglou, *Chem. Ber.*, 1991, **124**, 2377.
- R. W. Murray and D. Gu, *J. Chem. Soc., Perkin Trans. 2*, 1994, 451.
- R. Ballini, F. Papa and P. Bovicelli, *Tetrahedron Lett.*, 1996, **37**, 3507 and references cited therein.
- P. Bovicelli, P. Lupattelli, E. Mincione and V. Fiorini, *Tetrahedron Lett.*, 1993, **34**, 6103.
- G. Asensio, M. E., Gonzales-Núñez, C. B. Bernardini, R. Mello and W. Adam, *J. Am. Chem. Soc.*, 1993, **115**, 7250.
- Recently, Rosner and co-workers reported the importance of the dileucine motifs in biological systems. In particular, dileucine motifs have been implicated in the internalization of degradation of many membrane proteins: P. Morrison, K. C. Chung and M. R. Rosner, *Biochemistry*, 1996, **35**, 14 618.

Received in Liverpool, UK, 10th March 1997; Com. 7/01650F