

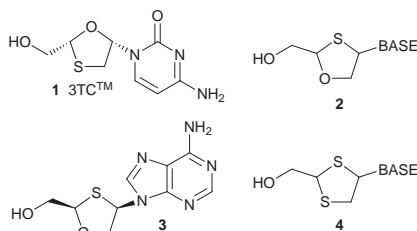
Synthesis and anti-HIV activity of 1,3-dithiolane nucleosides

Nghe Nguyen-Ba, William L. Brown, Laval Chan, Nola Lee, Livio Brasili, Dominique Lafleur and Boulos Zacharie*

BioChem Pharma Inc., 275 Armand-Frappier Blvd., Laval, Québec, Canada H7V 4A7

Received (in Corvallis, OR, USA) 3rd March 1999, Accepted 20th May 1999

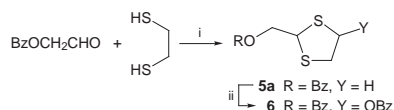
The potent activity displayed by 3'-azido-3'-deoxythymidine (AZT)¹ against human immunodeficiency virus (HIV) provides impetus for the development of novel nucleoside analogues.² Unfortunately, those compounds with the natural stereochemistry possess undesirable pharmacological properties³ and are susceptible to the development of resistant strains of HIV.^{3,4} In an attempt to overcome some of these detrimental side effects, the carbohydrate moiety of 2',3'-dideoxynucleoside analogs has been replaced by other five membered rings.^{3d,4} It has been demonstrated that hetero-substitution of these rings has a profound effect on the biological activity of the resulting nucleoside analogue⁵ as displayed by (–)-2'-deoxy-3'-thiacytidine (3TC, Efavir) **1**.^{5c,6}



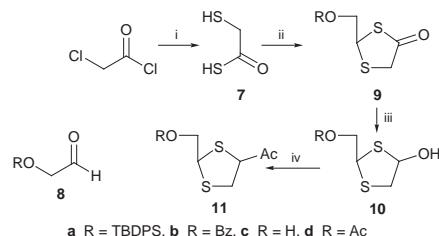
As part of an ongoing search for new anti-AIDS leads, we further explored this class of thioribonucleosides. These compounds possess improved metabolic stability to phosphor-ylases which cleave the glycosidic bond in nucleosides.⁷ Recently, we reported the anti-HIV activity of 2,4-disubstituted 1,3-oxathiolane nucleosides **2** by transposing the sulfur and oxygen atoms of **1**.^{4a} In this series, the (–)-adenine derivative **3** with the natural configuration was found to be twice as active as ddI in MT-4 cells. We further modified the oxathiolane ring by replacement of the oxygen atom of **2** with sulfur. This is exemplified by the general structure **4**. Here we report the synthesis and anti-HIV activity of this class of compounds.

The synthetic route to (±)-1,3-dithiolane compounds is based upon coupling a persilylated heterocyclic base with a dithiolane moiety **5** bearing a suitable leaving group Y at the 4-position under Vorbruggen's conditions.⁸ Two approaches were considered for the preparation of the ring **5**. The first route is the introduction of a leaving group at position 4 using peroxide derivatives. For example, dithiolane **6** was prepared by treating **5a** with benzoyl peroxide in refluxing benzene (Scheme 1).⁹

The second approach offers a more general route for the synthesis of the key intermediate 1,3-dithiolane **11**† with a variety of displaceable leaving groups at C-4. Our synthetic strategy was based on the preparation of 1,3-dithiolan-4-one **9**, followed by reduction and acylation to give 4-acyloxy-1,3-dithiolane **11** (Scheme 2). Thus, reaction of freshly distilled ClCH₂COCl with excess NaSH (3 equiv.) in absolute ethanol at –10 °C gave **7**¹⁰ in quantitative yield. The crude product was



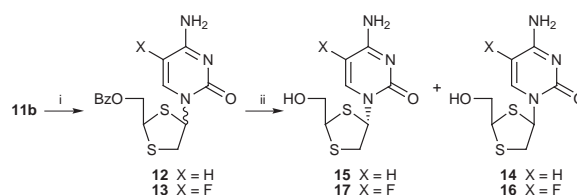
Scheme 1 Reagents and conditions: i, C₆H₆, TSA; ii, BzOOH, C₆H₆, heat.



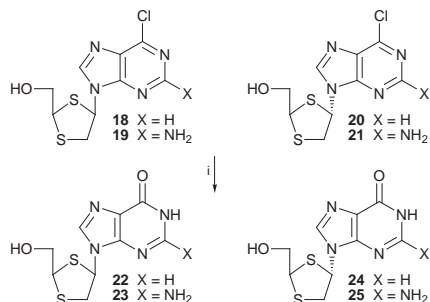
Scheme 2 Reagents and conditions: i, NaSH, absolute EtOH, –10 °C; ii, **8**, ZnI₂, CH₂Cl₂; iii, DIBAL-H, PhMe or BH₃·THF, B(MeO)₃; iv, AcOCl, Py.

immediately treated with aldehyde **8** in the presence of ZnI₂ as catalyst in CH₂Cl₂¹¹ to give the desired (±)-1,3-dithiolan-4-one **9** in moderate yield (50–60%). The initial synthetic procedure was based upon using the TBDPS protecting group for the hydroxy function of **9**. Reduction of **9a** with DIBAL-H (1.1 equiv.) in toluene gave thiolactol **10a** which was subsequently acylated to give the key intermediate **11a** in high yield. The silyl protecting group was later replaced by a benzoate group in order to facilitate the separation of the *cis* and *trans* nucleoside isomers. Applying the same conditions to reduce the benzoate **9b** did not result in any reduction product. However, using excess of DIBAL-H (3 equiv.) was successful and both the thiolactone and the benzoate function were reduced to give diol **10c** in 40% yield. Compound **10c** was then bis-acylated giving intermediate **11d** in high yield. Efforts were then directed to scale-up this procedure. Unfortunately, the DIBAL-H reduction proved particularly intractable. We therefore investigated other reducing agents that are selective and require little work-up. Only BH₃·THF (1.2 equiv.) catalyzed by B(OMe)₃ (1 equiv.) gave satisfactory results. The reduction was completed in 16 h and the product **10b** was obtained in 95% yield. This compound was then acylated to give the expected product **11b**. Following the same procedure, a number of different leaving groups (Bz, *m*-ClC₆H₄CH₂ and *p*-O₂NC₆H₄CH₂) were successfully introduced at C₄ of the sugar moiety **11**.

Compound **11b** is suitable for coupling with silylated cytosine or 5-fluorocytosine under refluxing conditions in CH₂Cl₂ and in the presence of SnCl₄ (Scheme 3). This gave the desired nucleoside analogue **12** or **13** as a 1 : 2 mixture of *cis* and *trans* isomers in moderate yields. Replacement of SnCl₄ with TMSI altered the ratio of the isomers. For example, compound **6** reacted with silylated N-acetylcytosine to give a mixture of the *cis* and *trans* nucleosides **12** in 62% yield with a slight predominance of the *cis* isomer.⁹ Similar results were obtained using other leaving groups at C₄. This did not improve the yield



Scheme 3 Reagents and conditions: i, 2,4-bis(trimethylsilyloxy)pyrimidine, ClCH₂CH₂Cl, SnCl₄, reflux, 16 h; ii, NH₃, MeOH.



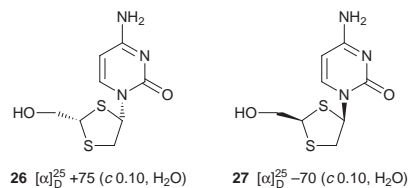
Scheme 4 Reagents and conditions: i, 25% aq. $\text{Me}_3\text{N}\cdot\text{H}_2\text{O}$, 55–65%.

or the *cis:trans* ratio. The next step was the separation of the isomers **12** or **13**. This was achieved by flash chromatography on silica gel by prior acetylation of the amino group (cytosine) or by reverse chromatography HPLC after deprotection (5-fluorocytosine). The protecting groups were then removed by treatment with methanolic ammonia to give the desired nucleosides **14–17** in high yields. The relative stereochemistry of these products was assigned by difference NOE spectra.

Similarly, uracil, thymine, adenine and guanine derivatives were produced from **11b** using the same conditions. However, in the case of hypoxanthine and guanine analogs **22–25**, synthesis was undertaken by treating the corresponding 6-chloropurine or 2-amino-6-chloropurine derivatives **18–21** with 20 equiv. of an aqueous Me_3N solution in water (Scheme 4).

The anti-HIV activity of (\pm)-1,3-dithiolane nucleoside analogues was evaluated in MT-4 (human T helper) cells at concentrations up to $100\ \mu\text{g}\ \text{ml}^{-1}$ and compared with 3TCTM (EpiVir)¹² and the 5-fluoro derivative (FTC).¹³ In this assay, only *cis* cytosine and 5-fluorocytosine derivatives **14** and **16** displayed inhibitory activity at ID_{50} of 9.3 and $4.8\ \mu\text{g}\ \text{ml}^{-1}$ and were not cytotoxic at $100\ \mu\text{g}\ \text{ml}^{-1}$, whereas 3TCTM and FTC showed anti-HIV activity at 0.3 and $0.14\ \mu\text{g}\ \text{ml}^{-1}$, respectively. All the other nucleosides did not exhibit antiviral activity with no cytotoxicity up to $100\ \mu\text{g}\ \text{ml}^{-1}$. In contrast, *cis* and *trans* 6-chloropurine derivatives **18** and **20** showed cytotoxicity at CD_{50} of $10\ \mu\text{g}\ \text{ml}^{-1}$.

Described herein is a novel class of anti-HIV (\pm)-1,3-dithiolane nucleoside analogues. The biological results demonstrate that replacement of an oxygen atom of the oxathiolane with sulfur causes reduction in antiviral activity. It should be noted that compounds **14** and **16** are racemic. Resolution of the enantiomers may improve the activity. Therefore, enantiomeric separations of the racemic *cis* **14** was undertaken by chiral HPLC.¹⁴ This gave the two enantiomers **26** and **27** in a



reasonable yield. It was found that isomer **27** possesses the natural configuration as evidenced by enzymatic resolution of the racemic mixture. Thus treatment of the mixture of the two enantiomers with cytidine deaminase converted only **27** to its corresponding uracil derivative. However, the unnatural enantiomer **26** was recovered and characterized by comparison of HPLC retention time and optical rotation with the previously isolated isomer. Both enantiomers were submitted for anti-HIV evaluation. Neither of the two compounds displayed improved antiviral activity.

We thank Drs T. Bowlman and R. Storer for reading the manuscript, Drs P. Hopewell and N. Cammack of Glaxo Group Research for testing the compounds, Ms L. Bernier and J. Dugas

for technical assistance with HPLC separation of enantiomers, and Ms L. Marcil for secretarial and technical assistance.

Notes and references

† Selected data for **10b**: colorless oil; δ_{H} (CDCl_3): 8.03 (m, 2H), 7.57 (dt, 1H, J 7, 1), 7.44 (t, 2H, J 7), 5.85 (m, 1H), 4.84 and 4.76 (t's, 1H, J 7), 4.68 and 4.39 (m's, 1H), 4.48 and 4.27 (m's, 1H), 3.30 (m, 2H), 2.85 (m, 1H). For **16**: δ_{H} ($\text{DMSO}-d_6$) 8.34 (d, 1H, J 7.5), 7.83 (br s, 1H), 7.59 (br s, 1H), 6.39 (m, 1H), 5.57 (t, 1H, J 5.5), 4.62 (t, 1H, J 6), 3.75 (t, 2H, J 6), 3.59 (m, 2H). For **17**: δ_{H} ($\text{DMSO}-d_6$) 8.10 (d, 1H, J 7 Hz), 7.78 (br s, H), 7.54 (br s, 1H), 6.41 (t, 1H, J 2), 5.36 (t, 1H, J 5.5), 4.81 (t, 1H, J 7 Hz), 3.45 (m, 4H). For **19**: δ_{H} ($\text{DMSO}-d_6$) 8.45 (s, 1H), 7.03 (br s, 2H), 6.40 (t, 1H, J 4), 5.53 (t, 1H, J 6), 4.73 (t, 1H, J 7), 3.80 (dd, 1H, J 13, 4.5), 3.74 (t, 2H, J 6), 3.63 (dd, 1H, J 13, 5); δ_{C} ($\text{DMSO}-d_6$) 159.80, 153.49, 150.01, 141.95, 123.61, 65.57, 64.52, 56.23, 42.93; HRMS (FAB): M^+ calc. for $\text{C}_9\text{H}_{11}\text{ClN}_5\text{OS}_2$ 304.00937, found 304.00880. For **21**: δ_{H} ($\text{DMSO}-d_6$) 8.34 (s, 1H), 7.02 (br s, 2H), 6.44 (t, 1H, J 3), 5.41 (t, 1H, J 6), 4.86 (t, 1H, J 7), 3.58 (m, 3H), 3.49 (m, 1H); HRMS (FAB): M^+ calc. for $\text{C}_9\text{H}_{11}\text{ClN}_5\text{OS}_2$ 304.00937, found 304.00840. For **26**: mp 108–110 °C; δ_{H} ($\text{DMSO}-d_6$) 8.06 (d, 1H, H-6', J 7.63), 7.20 (br d, 2H, NH_2), 6.47 (t, 1H, J 4.4), 5.74 (d, 1H, J 7.42) 5.50 (t, 1H), 4.61 (t, 1H, J 6.5 Hz), 3.74 (t, 2H, J 6.00 Hz), 3.46 (dd, 1H, J 4.30, 12.9), and 3.36 (dd, 1H, J 4.1, 10.4); HRMS (FAB): M^+ calc. for $\text{C}_8\text{H}_{12}\text{N}_3\text{O}_2\text{S}_2$ 246.03709, found 246.03610. For **27**: mp 200–202 °C (decomp.); δ_{H} ($\text{DMSO}-d_6$) 7.90 (d, 1H, J 7.35), 7.17 (br d, 2H), 6.48 (d, 1H, J 3.72), 5.69 (d, 1H, J 7.47), 5.38 (t, 1H), 4.74 (t, 1H, J 6.83), 3.43 (m, 4H); HRMS (FAB): M^+ calc. for $\text{C}_8\text{H}_{12}\text{N}_3\text{O}_2\text{S}_2$ 246.03709, found 246.03640.

- H. Mitsuya, J. K. Weinhold, P. A. Furman, M. H. St-Clair, S. Nusinoff-Lehrman, R. C. Gallo, D. Bolognesi, D. W. Barry, S. Broder, *Proc. Natl. Acad. Sci. U.S.A.*, 1985, **82**, 7096.
- H. Mitsuya and S. Broder, *Proc. Natl. Acad. Sci. U.S.A.*, 1986, **83**, 1911; R. Yarchoan, H. Mitsuya, R. V. Thomas, J. M. Pluda, N. R. Hartman, C.-F. Perno, K. S. Marczyk, J.-P. Allain, D. G. Johns and S. Broder, *Science*, 1989, **245**, 412; T.-S. Lin, R. F. Schinazi and W. H. Prusoff, *Biochem. Pharmacol.*, 1987, **36**, 2713.
- (a) R. W. Klecker, J. M. Collins, R. Yarchoan, R. Thomas, J. F. Jenkins, S. Broder, C. E. Myers, *Pharmacol. Ther.*, 1987, **47**, 407; (b) M. S. Hirsch and J. C. Kaplan, *Antimicrob. Agents Chemother.*, 1987, **31**, 939; (c) R. Yarchoan, *Lancet*, 1988, **i**, 76; (d) M. L. Peterson and R. Vince, *J. Med. Chem.*, 1991, **34**, 2787 and references cited therein.
- (a) B. Belleau, L. Brasili, L. Chan, M. D. DiMarco, B. Zacharie, N. Nguyen-Ba, H. J. Jenkinson, J. A. V. Coates, J. M. Cameron, *Bioorg. Med. Chem. Lett.*, 1993, **3**, 1723; (b) M. J. Bamford, D. C. Humber, R. Storer, *Tetrahedron Lett.*, 1991, **32**, 271 and references cited therein; (c) J. Branalt and I. Kvarnstrom, *J. Org. Chem.*, 1996, **61**, 3604 and references cited therein.
- (a) T. S. Mansour, C. A. Evans, M. A. Siddiqui, M. Charron, B. Zacharie, N. Nguyen-Ba, N. Lee and B. Korba, *Nucleosides Nucleotides*, 1997, **16**, 993; (b) H. Soudeyans, X. J. Yao, Q. Gao, B. Belleau, J.-L. Kraus, N. Nguyen-Ba, B. Spira and M. A. Wainberg, *Antimicrob. Agents Chemother.*, 1991, **35**, 1386; (c) J. A. V. Coates, N. Cammack, H. J. Jenkinson, I. M. Mutton, B. A. Pearson, R. Storer, J. M. Cameron and C. R. Penn, *Antimicrob. Agents Chemother.*, 1992, **36**, 202; (d) M. W. Chun, D. H. Shin, H. R. Moon, J. Lee, H. Park and L. S. Jeong, *Bioorg. Med. Chem. Lett.*, 1997, **7**, 1475.
- U.S. Pat. 05047407, BioChem Pharma Inc.; M. A. Nowak, S. Bonhoeffer, A. M. Hill, R. Bochner, H. C. Thomas and H. McDade, *Proc. Natl. Acad. Sci. U.S.A.*, 1996, **93**, 4398.
- J. A. Secrist III, K. N. Tiwari, A. T. Shortnacy-Fowler, L. Messini, J. M. Riordan and J. A. Montgomery, *J. Med. Chem.*, 1998, **41**, 3865; M. R. Dyson, P. L. Coe and R. T. Walker, *J. Med. Chem.*, 1991, **34**, 2782.
- H. Vorbruggen, K. Krolikiewicz and B. Bennua, *Chem. Ber.*, 1981, **114**, 1234.
- N. Nguyen-Ba, W. Brown, N. Lee and B. Zacharie, *Synthesis*, 1998, 759.
- M. Therien, J. Y. Gauthier and R. N. Young, *Tetrahedron Lett.*, 1988, **29**, 6733.
- J. Y. Gauthier, T. Henien, L. Lo, M. Therien and R. N. Young, *Tetrahedron Lett.*, 1988, **29**, 6729.
- J. M. Cameron, P. Collis, M. Daniel, R. Storer and P. Wilcox, *Drugs Future*, 1993, **18**, 319 and references cited therein.
- L. W. Frick, L. St-John, L. C. Taylor, G. R. Painter, P. A. Furman, D. C. Liotta, E. S. Furfine and D. J. Nelson, *Antimicrob. Agents Chemother.*, 1993, **37**, 2285.
- Chiral Column: Cyclobond I 2000 Beta-RSP 4.6 mm ID \times 250 mm; mobile phase 10% MeCN–0.05% (AcOH–Et₃N, pH 6.74); pressure 965 psi and flow rate 0.50 ml min⁻¹. For **26**: t_{R} = 19.880 min; for **27**: t_{R} = 22.201 min.

Communication 9/01927H