

A highly water-soluble 2:1 β -cyclodextrin–fullerene conjugateSalvatore Filippone,^{ab} Frank Heimann^a and André Rassat^{*a}^a Département de Chimie, École Normale Supérieure and CNRS, 24 Rue Lhomond, F75231 Paris Cedex 05, France. E-mail: andre.rassat@ens.fr; Fax: +33 14432 3325; Tel: +33 14432 3266^b Laboratoires de Biophysique et de Photobiologie, Muséum National d'Histoire Naturelle, CNRS and INSERM, 43 Rue Cuvier, F-75231 Paris Cedex 05, France

Received (in Cambridge, UK) 8th March 2002, Accepted 15th May 2002

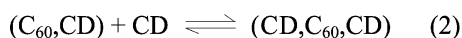
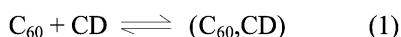
First published as an Advance Article on the web 11th June 2002

A novel 2:1 (permethylated- β -cyclodextrin)–fullerene conjugate has been synthesised; highly soluble in cold water with formation of aggregates, it displays a negative solubility coefficient and has a partition coefficient between octanol and water ($\log P = 1.58$) in the suitable range for biological studies.

The study of the biological activity of fullerenes depends on the availability of water-soluble derivatives.¹ Such molecules have been obtained either by covalent addition of hydrophilic appendages or by complex formation with host molecules.² In both cases, it is advisable to use auxiliary compounds whose properties do not interfere with those of the fullerenes. Cyclodextrins (CD) (and their permethylated derivatives), known to be essentially non-toxic (at least β -CD),³ are particularly attractive in this respect, and in fact have been used both as hydrophilic appendage^{4,5} and as water-soluble carrier.^{6–9}

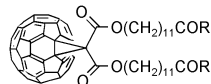
These two types of derivatives possess different properties: contrary to the non-covalent 2:1 γ -CD–C₆₀ complex,^{6–8} the UV-Vis spectra of water solution of the 2:1 β -CD–C₆₀ complex,⁹ or of a 1:1 β -CD–C₆₀ covalent conjugate⁴ are typical of the presence of aggregates, also revealed by direct physical measurements.¹⁰ Other 1:1 covalent conjugates (with α -, β - or γ -CD) have been prepared but are apparently less water-soluble.¹¹

In the case of the 2:1 γ -CD–fullerene complex, different equilibria, including (1) and (2) (Scheme 1), may take place in solution, so that if some other substrate with sufficient affinity for γ -CD were present, the fullerene could be displaced and possibly precipitate. A covalent binding between the fullerene and the γ -CD would probably impede this displacement.

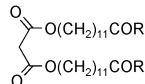
Scheme 1 Equilibria between γ -CD and C₆₀ in water.

Because of the formation of aggregates, the equilibria of the β -CD–fullerene 2:1 complex are apparently more complicated. However, here again, a covalent link could stabilise the complex, an interesting possibility because of the easier availability of the β -CD.

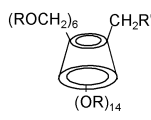
We have thus prepared the covalent conjugate **1a** in which the linker was expected to allow solvent-dependent equilibria between conformers such as **A**, **B** and **C** (Scheme 2). In water, **A** and **B** could form micelle-like aggregates, but if the CDs ensure sufficient hydrophilic protection, **C** could exist as a non-associated species.



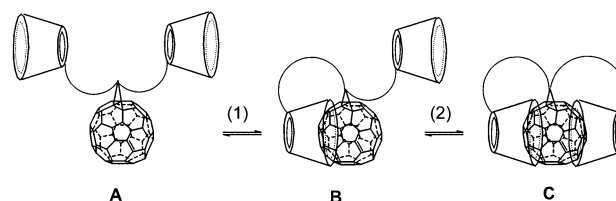
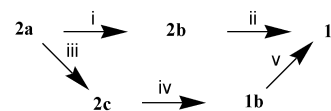
1a R = NH-PMCD
1b R = OBU^t



2a R = OH
2b R = NH-PMCD
2c R = OBU^t



3a R = H; R' = OTs
3b R = H; R' = N₃
3c R = Me; R' = N₃
3d R = Me; R' = NH₂

Scheme 2 Possible equilibria between conformers **A**, **B** and **C** of **1a**.

Scheme 3 Reagents and conditions: i, **3d**, 1-hydroxybenzotriazole (HOBt), CH₂Cl₂, DCC, rt, 48 h (89%); ii, C₆₀, toluene, CBr₄, DBU, rt, 14 h (30%); iii, (COCl₂, CH₂Cl₂, reflux, 36 h, solvent removal, then *t*-BuOH, Py (69%); iv, C₆₀, toluene, CBr₄, DBU, rt, 14 h (38%); v, TFA, CH₂Cl₂, rt, 4 h, solvent removal, then **3d**, DCC, HOBt, CH₂Cl₂, rt, 48 h (75%).

Two different routes led to **1a**[†] (Scheme 3), both starting from **2a**.^{†12} 6^A-amino-6^A-deoxy-per-(O-methyl)- β -cyclodextrin (NH₂-PMCD) **3d** was prepared in 28% yield from β -CD by a combination of published methods.¹³ The methoxy groups were introduced in order not only to prevent side reactions but also to increase the solubility of the β -CD moiety, in spite of the negative solubility coefficient of the related permethylated β -CD;¹⁴ functionalisations of C₆₀ were effected by Hirsch-Bingel (HB) reactions.¹⁵

Soluble in toluene, methanol, ethanol, acetonitrile, dichloromethane and chloroform, **1a** may also be dissolved in water at 20 °C up to a limit of 90 mg mL⁻¹, one of the highest reported solubilities at pH 7.¹ Aggregates are present in water solutions: in dichloromethane, the expected UV-Vis spectrum of a methanofullerene is observed; in water, this spectrum is less resolved and a relative maximum at 430 nm is missing (Fig. 1), a sign of aggregate formation;^{16,17} similarly, the NMR peaks are much broader in water than in chloroform. No induced circular dichroism could be detected in water or in dichloromethane, thus excluding an appreciable population of conformer **C** and

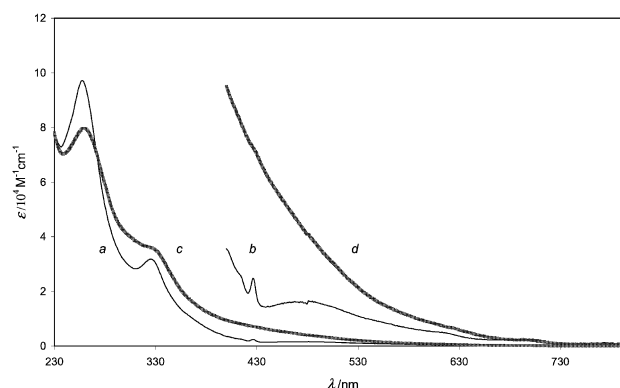
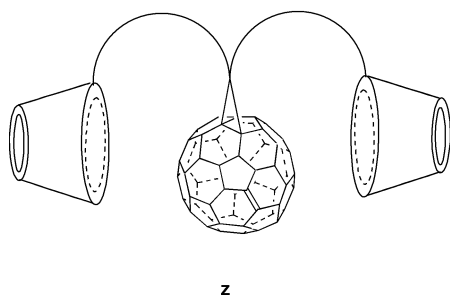


Fig. 1 Absorption spectra of **1a**: (a) 10⁻⁵ M in CH₂Cl₂; (b) same, $\times 10$; (c) 10⁻⁵ M in water; (d) same, $\times 10$.

suggesting a predominance of the extended form **A** in these solutions. Like the related permethyl- β -CD, **1a** has a negative solubility coefficient in water: when heated, clear 10^{-3} M and 2.5×10^{-4} M solutions became turbid at 30 and 42 °C, respectively and returned to their original state after cooling. A measure of the partition coefficient between octanol and water gave $\text{Log } P = 1.58$,¹⁸ in the range suitable for allowing penetration of cell membranes¹⁹ or oral absorption.²⁰

Thus, although both the high solubility in water at neutral pH and the convenient partition coefficient of **1a** make it well suited for biological studies, it may be desirable to eliminate the formation of aggregates in water. Since this phenomenon may be due to a poor steric fit or to a 'wrong' orientation of the β -CDs, it is possible that the γ -CD homologue of **1a** or β - or γ -CD conjugates such as **Z** be very soluble in water as well, but now without forming aggregates. Work along these lines is in progress.



We acknowledge financial support from the ENS, the CNRS and the European Union TMR contract BIOFULLERENES. We thank Dr René Bensasson for his interest in this work and Professor Pierre Sinaÿ and Bérengère du Roizel for a free and continuous exchange of information.

Notes and references

† Selected analytical data **1a**: Calc. for $C_{211}H_{264}N_2O_{74} \cdot 7H_2O$: C 61.24, H 6.77, N 0.68; found: C 61.31, H 6.96, N: 0.57%. MS (FAB+, NaI) m/z : 4035 [M + Na]⁺ (40%), 1633. ¹H NMR (400 MHz, CDCl₃): δ 1.2–1.48 (m, 32H), 1.63 (br s, 4H), 1.85 (m, 4H), 2.18 (m, 4H), 3.16 (dd, $J = 3.3, 9.5$ Hz, 2H), 3.21 (m, 12 H), 3.38–3.46 (m, 40 H), 3.5–3.56 (m, 52 H), 3.58–3.70 (m, 70 H), 3.85 (m, 28 H), 4.5 (t, $J = 6.5$ Hz, 4H), 5.12 (t, $J = 3.0$ Hz, 4H), 5.16 (t, $J = 2.9$ Hz, 6H), 5.19 (br t, 4H), 6.02 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 25.84, 25.93, 25.53, 29.19, 29.36, 29.44, 29.55, 29.58, 36.84, 39.73, 58.26, 58.28, 58.31, 58.44, 58.56, 58.61, 58.91, 58.97, 58.98, 59.05, 59.32, 61.16, 61.24, 61.35, 61.39, 61.48, 61.59, 67.40, 69.83, 70.81, 70.92, 70.95, 71.07, 71.10, 71.15, 71.30, 71.47, 71.50, 71.61, 79.89, 79.94, 80.07, 80.23, 80.56, 80.70, 80.76, 81.32, 81.40, 81.52, 81.61, 81.72, 81.75, 81.92, 81.96, 82.06, 82.09, 98.55, 98.62, 98.72, 98.91, 99.00, 138.91, 140.89, 141.85, 142.14, 142.92, 142.96, 143.03, 143.82, 144.54, 144.59, 144.63, 144.82, 145.12, 145.20, 145.33, 163.66, 173.17.

1b: Calc. for $C_{95}H_{62}O_8$: C 85.69, H 4.69; found: C 84.11, H: 4.78%. ¹H NMR (200 MHz, CDCl₃): δ 1.10–1.62 (m 54 H), 2.15 (t, $J = 7.3$ Hz, 4H), 4.48 (t, $J = 6.3$ Hz, 4H). ¹³C NMR (50 MHz, CDCl₃): δ 25.0, 25.9, 28.04, 28.49, 29.02, 29.15, 29.24, 29.41, 29.47, 35.5, 53.0, 67.3, 72.0, 79.8,

138.89, 140.82, 141.78, 142.08, 142.88, 142.89, 142.96, 143.76, 144.48, 144.55, 144.75, 145.06, 145.13, 145.25, 163.6, 173.2.

2a: Mp 71–73 °C.

2b: Calc. for $C_{151}H_{266}N_2O_{74}$: C 55.06, H 8.30, N 0.85; found: C 54.50, H 8.14, N 1.08%. MS (FAB+, NaI), m/z : 3315 (100%) [M + Na]⁺. ¹H NMR (400 MHz, CDCl₃): δ 1.2–1.35 (m, 32H), 1.61 (m, 4H), 2.16 (dd, $J = 6.2, 6.4$ Hz, 4H), 3.14 (dd, $J = 3.3, 9.5$ Hz, 2H), 3.18 (m, 12H), 3.35 (s, 2H), 3.39–3.43 (m, 40H), 3.48–3.53 (m, 58H), 3.55–3.70 (m, 68H), 3.73–3.91 (m, 28H), 4.11 (t, $J = 6.7$ Hz, 4H), 5.08 (t, $J = 2.8$ Hz, 4H), 5.12 (t, $J = 3.5$ Hz, 6H), 5.16 (t, $J = 4$ Hz, 4H), 6.02 (br t, 2H). ¹³C NMR (100 MHz, CDCl₃): δ 25.63, 25.73, 28.29, 29.06, 29.24, 29.29, 29.34, 29.39, 29.41, 36.72, 40.05, 58.16, 58.18, 58.20, 58.34, 58.47, 58.51, 58.80, 58.87, 58.88, 58.92, 58.95, 59.22, 61.05, 61.14, 61.25, 61.29, 61.38, 61.39, 61.48, 65.51, 69.74, 70.71, 70.81, 70.86, 71.36, 70.98, 71.01, 71.05, 71.21, 71.39, 79.76, 79.84, 79.86, 79.96, 80.10, 80.20, 80.45, 81.21, 81.30, 81.42, 81.51, 81.62, 81.66, 81.67, 81.82, 81.86, 81.92, 81.96, 82.00, 98.47, 98.50, 98.67, 98.81, 98.88, 98.92, 166.7, 173.12.

- 1 T. Da Ros and M. Prato, *Chem. Commun.*, 1999, 663–669.
- 2 T. Wharton, V. U. Kini, R. A. Mortis and L. J. Wilson, *Tetrahedron Lett.*, 2001, **42**, 5159–5162 and references therein.
- 3 K. Uekama, F. Hiramyama and T. Irie, *Chem. Rev.*, 1998, **98**, 2045–2076.
- 4 S. Samal and K. E. Geckeler, *Chem. Commun.*, 2000, 1101–1102.
- 5 S. Samal, B. J. Choi and K. E. Geckeler, *Chem. Commun.*, 2000, 1373–1374.
- 6 T. Andersson, K. Nilsson, M. Sundahl, G. Westman and O. Wennerstrom, *J. Chem. Soc., Chem. Commun.*, 1992, 604–606.
- 7 K. I. Priyadarsini, H. Mohan, A. K. Tyagi and J. P. Mittal, *J. Phys. Chem.*, 1994, **98**, 4756–4759.
- 8 T. Andersson, G. Westman, O. Wennerstrom and M. Sundahl, *J. Chem. Soc., Perkin Trans. 2*, 1994, 1097–1101.
- 9 C. N. Murthy and K. E. Geckeler, *Chem. Commun.*, 2001, 1194–1195.
- 10 S. Samal and K. E. Geckeler, *Chem. Commun.*, 2001, 2224–2225.
- 11 D. Q. Yuan, K. Koga, Y. Kourogi and K. Fujita, *Tetrahedron Lett.*, 2001, **42**, 6727–6729.
- 12 From 12-hydroxydodecanoic acid and malonyldichloride.
- 13 (a) **3a**: I. Baussanne, J. M. Benito, C. O. Mallet, J. M. Garcia Fernandez, H. Law and J. Defaye, *Chem. Commun.*, 2000, 1489–1490; (b) **3b**: H. Parrot-Lopez, H. Galons, S. Dupas, M. Miocque and G. Tsoucaris, *Bull. Soc. Chim. Fr.*, 1990, **127**, 568–571; (c) **3c, 3d**: M. T. Reetz and S. R. Waldvogel, *Angew. Chem., Int. Ed.*, 1997, **36**, 865–867.
- 14 E. B. Starikov, K. Brasicke, E. W. Knapp and W. Saenger, *Chem. Phys. Lett.*, 2001, **336**, 504–510, and references therein.
- 15 X. Camps and A. Hirsch, *J. Chem. Soc., Perkin Trans. 1*, 1997, 1595–1596.
- 16 R. V. Bensasson, E. Bienvenue, M. Dellinger, S. Leach and P. Seta, *J. Phys. Chem.*, 1994, **98**, 3492–3500.
- 17 R. V. Bensasson, E. Bienvenue, C. Fabre, J. M. Janot, E. J. Land, S. Leach, V. Leboulaire, A. Rassat, S. Roux and P. Seta, *Chem. Eur. J.*, 1998, **4**, 270–278.
- 18 The partition coefficient between octanol and water was determined at 21 °C: The UV-Vis spectrum of a solution of **1a** (12 mg) in water-saturated *n*-octanol (10 mL) was recorded. This solution was then shaken with water (10 mL) for 1 h. After one-hour rest, the UV-Vis spectrum of the separated octanol-phase was recorded, and the partition coefficient was calculated from the absorbance at 426 nm.
- 19 J. Sangster, *Octanol/Water Partition Coefficients Fundamental and Physical Chemistry*, John Wiley and Sons, Chichester, 1997.
- 20 A. W. Jensen, S. R. Wilson and D. I. Schuster, *Bioorg. Med. Chem.*, 1996, **4**, 767–779.