

Efficient synthesis of a 4,5-epoxy-2-cyclohexen-1-one derivative bearing a spiro lactone *via* a Diels–Alder reaction with high π -facial selectivity: a synthetic study towards scyphostatin

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The Diels–Alder reaction of spiro lactones with cyclopentadiene afforded the adduct with high π -facial selectivity; a hydrophilic analogue of scyphostatin was synthesized from the Diels–Alder adduct.

Scyphostatin **1** (Fig. 1) was isolated from the mycelial extract of *Dasyascyphus mollissima* by Ogita and coworkers (Sankyo Co Ltd., Japan) in 1997 and was found to be a potent inhibitor of neutral sphingomyelinase (N-SMase).¹ Scyphostatin **1** consists of a hydrophobic side chain and a hydrophilic 4,5-epoxy-2-cyclohexen-1-one moiety. Our interest in such biphilic natural products² has inspired us to carry out synthetic studies on scyphostatin **1**. Other groups have also reported synthetic efforts.³ In this paper, the efficient synthesis of hydrophilic analogue **2** *via* a Diels–Alder reaction with high π -facial selectivity is described.

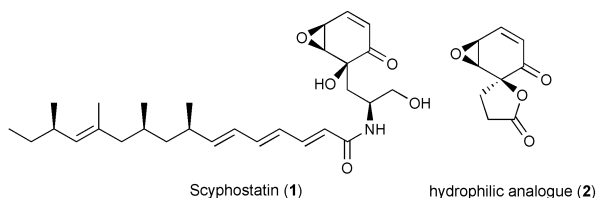
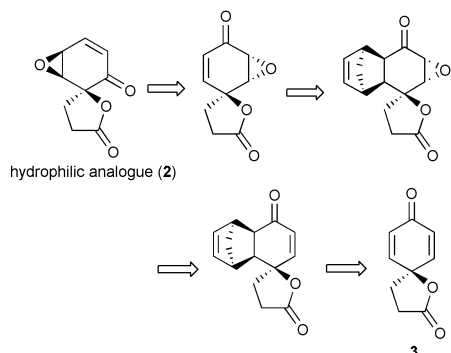


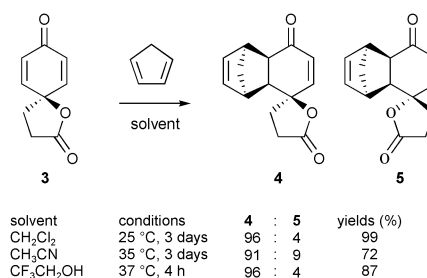
Fig. 1

Masked benzoquinones, obtained by the Diels–Alder reaction of cyclopentadiene and *p*-benzoquinones, have proven useful as precursors to highly functionalized cyclohexene derivatives.⁴ Especially noteworthy is the high π -facial selectivity in the Diels–Alder reaction of spiro lactone **3** with sterically hindered chiral bicyclic cyclopentadiene derivatives reported by Winterfeldt *et al.*⁵ Thus, we decided to look into the possibility using of this strategy by reacting spiro lactone with sterically undemanding cyclopentadienes for the preparation of the hydrophilic moiety as outlined in Scheme 1.

The spiro lactone **3** was synthesized by oxidative intramolecular lactonization with $\text{PhI}(\text{OAc})_2$.⁶ The Diels–Alder reaction of spiro lactone **3** with cyclopentadiene in CH_2Cl_2 and $\text{CF}_3\text{CH}_2\text{OH}$ preferentially afforded the adduct **4** (**4**:**5** = 96:4,



Scheme 1



Scheme 2

Scheme 2). The Diels–Alder reaction of spiro lactone **3** was accelerated in $\text{CF}_3\text{CH}_2\text{OH}$. When CH_3CN was used as a solvent, the π -facial selectivity of spiro lactone **3** with cyclopentadiene slightly decreased (**4**:**5** = 91:9). The mixtures of the adducts could be separated by HPLC. The stereochemistry of the adducts **4** and **5** was assigned by ¹H NMR NOE experiments (Fig. 2).⁷ Thus, we have found that even if the diene is sterically undemanding, the Diels–Alder reaction of spiro lactone **3** proceeds with high π -facial selectivity.

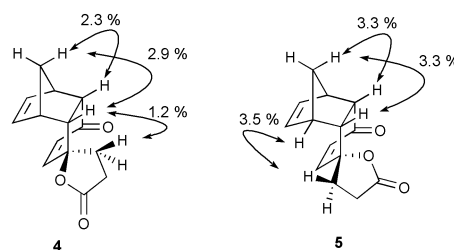
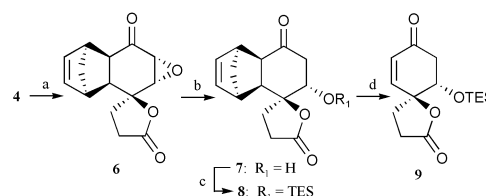


Fig. 2 Determination of the relative stereochemistry of adducts **4** and **5**.

The high π -facial selectivity in Diels–Alder reaction of spiro lactone **3** with cyclopentadiene prompted us to use it for the synthesis of **2**, a model compound for the hydrophilic moiety of scyphostatin **1** (Scheme 3). Epoxide **6** was obtained by epoxidation with $\text{H}_2\text{O}_2/\text{LiOH}$, followed by reclosure to the lactone ring with 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide (WSCl) (66% over two steps). The epoxide ring of **6** was reductively cleaved with SmI_2 (89%) to give **7** as the only ring opened product.⁸ Protection of the secondary alcohol with

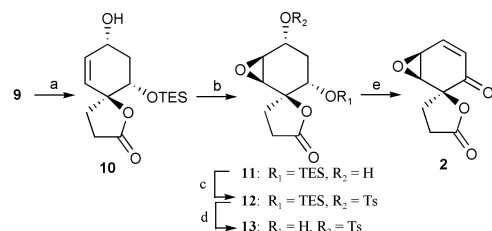


Scheme 3 Reagents and conditions: (a) 30% H_2O_2 , LiOH, THF, 0 °C, then WSCI, CH_2Cl_2 , 25 °C, 66% over two steps; (b) SmI_2 , MeOH, THF, –78 °C, 89%; (c) TESCl, imidazole, CH_2Cl_2 , 25 °C, 100%; (d) maleic anhydride, Ph_2O , 230 °C, 96%.

triethyl silyl group (TES) gave **8** (100%). Retro-Diels–Alder product **9** was obtained by heating (230 °C) **8** in the presence of maleic anhydride (96%).

With 5-hydroxy-2-cyclohexen-1-one bearing a spirolactone **9** in hand, we focused our attention on its conversion to 4,5-epoxy-2-cyclohexen-1-one (Scheme 4). Treatment of **9** with NaBH₄/CeCl₃ gave alcohol **10** as a single isomer (99%). The relative stereochemistry was assigned by coupling constants ($J_{H1-H2ax}$, $J_{H2ax-H3}$) and NOE experiments in ¹H NMR (Fig. 3). Contrary to our expectations, epoxide **11** was obtained as a single isomer upon epoxidation of **10** with 3-chloroperoxybenzoic acid (*m*CPBA) (95%).⁹ The relative stereochemistry was determined by ¹H NMR NOE experiments and coupling constants (Fig. 3). NOE enhancement between H5 and H7 was observed. The observed facial selectivity in the epoxidation reaction can be attributed to attack on the face opposite to the lactone CH₂ group to avoid repulsion, despite the presence of a hydroxy directing group (Fig. 4). Epoxide **11** was converted to epoxide **13** by tosylation, followed by removal of the TES group with TBAF (59% over two steps). When epoxide **13** was treated under Swern oxidation conditions, 4,5-epoxy-2-cyclohexen-1-one, bearing a spirolactone **2**, a hydrophilic analogue of scyphostatin **1**, was obtained with simultaneous extrusion of the TsO group (75%).¹⁰

In summary, we have developed a short and high efficient method of synthesizing of 4,5-epoxy-2-cyclohexen-1-one derivative **2**, a hydrophilic analogue of scyphostatin **1**. The method features a highly selective Diels–Alder reaction, reductive oxirane ring opening, 1,2-ene reduction, and



Scheme 4 Reagents and conditions: (a) NaBH₄, CeCl₃·7H₂O, THF, *i*PrOH, 0 °C, 99%; (b) *m*CPBA, CH₂Cl₂, 25 °C, 95%; (c) TsCl, DABCO, CH₂Cl₂, 25 °C, 79%; (d) TBAF, AcOH, THF, 25 °C, 75%; (e) (COCl)₂, DMSO, CH₂Cl₂, -60 °C, 75%.

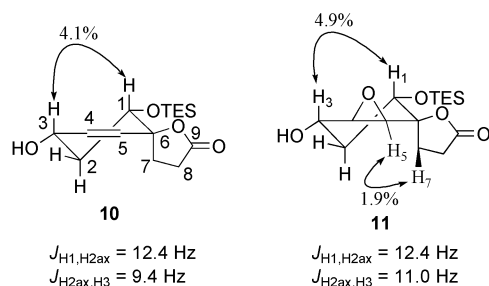


Fig. 3 Determination of the relative stereochemistry of **10** and **11**.

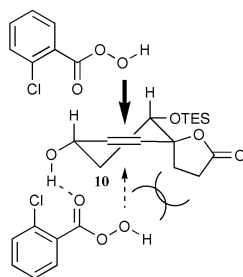


Fig. 4 Stereoselectivity in epoxidation of **10** with *m*CPBA.

subsequent epoxidation. Investigations towards the total synthesis of scyphostatin **1** based on this convenient strategy is now in progress.

The measurements of NMR and MS were made using JEOL JMN-LA500 and JEOL SX-102A, respectively, at the Instrument Center for Chemical Analysis, Hiroshima University. We thank Dr. Yoshikazu Hiraga for measurements of NMR (JEOL JMN-LA500) at the Hiroshima Prefectural Institute of Science and Technology. Financial supports of this work through Grant-in-Aid for Scientific Research (no.11740355 and no.14740349) provided by the Japan Society for the Promotion of Science and KANEKO-NARITA Research Bounty by Protein Research Foundation are heartily acknowledged. The authors thank F-TECH, Inc. for the gift of CF₃CH₂OH.

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- Spectral data for 4**: ¹H NMR (CDCl₃, 500 MHz) δ 1.35 (d, 1 H, *J* 8.7 Hz), 1.48 (d, 1 H, *J* 8.7 Hz), 2.15–2.27 (m, 2 H), 2.62 (ddd, 1 H, *J* 3.7, 8.5, 17.9 Hz), 2.78 (ddd, 1 H, *J* 9.2, 10.8, 17.9 Hz), 2.88 (ddd, 1 H, *J* 1.2, 3.3, 8.7 Hz), 3.04 (dd, 1 H, *J* 4.4, 8.7 Hz), 3.18 (m, 1 H), 3.39 (m, 1 H), 5.85 (dd, 1 H, *J* 1.1, 9.2 Hz), 5.87 (dd, 1 H, *J* 1.1, 10.0 Hz), 6.14 (dd, 1 H, *J* 2.5, 10.0 Hz), 6.52 (dd, 1 H, *J* 2.5, 9.2 Hz); ¹³C NMR (CDCl₃, 125 MHz) δ 27.5, 38.6, 45.2, 47.5, 47.4, 48.7, 50.8, 83.0, 130.7, 134.8, 135.4, 148.9, 175.9, 199.1; EI-HRMS *m/z* 230.0940 (M⁺, C₁₄H₁₄O₃ requires 230.0943). **5**: ¹H NMR (CDCl₃, 500 MHz) δ 1.47 (d, 1 H, *J* 8.6), 1.57 (dt, 1 H, *J* 1.6, 8.6 Hz), 2.17 (ddd, 1 H, *J* 4.6, 15.4, 13.1 Hz), 2.60 (ddd, 1 H, *J* 9.0, 10.4, 13.1 Hz), 2.71–2.78 (m, 2 H), 3.02 (ddd, 1 H, *J* 1.3, 3.2, 8.4 Hz), 3.19 (dd, 1 H, *J* 4.4, 8.4 Hz), 3.24 (s, 1 H), 3.44 (s, 1 H), 5.96–6.01 (m, 2 H), 6.07 (d, 1 H, *J* 10.2 Hz), 6.45 (dd, 1 H, *J* 1.3, 10.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 28.55, 31.88, 46.37, 46.47, 47.01, 49.38, 49.87, 82.23, 132.92, 134.10, 136.18, 143.02, 174.84, 199.11; FAB-HRMS *m/z* 231.1015 (M + H, C₁₄H₁₅O₃ requires 231.1021).
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- Spectral data for 2**: ¹H NMR (500 MHz, CDCl₃) δ 2.38 (dt, 1 H, *J* 9.1, 13.2 Hz), 2.70 (ddd, 1 H, *J* 3.8, 9.1, 17.5 Hz), 2.78 (ddd, 1 H, *J* 3.8, 9.2, 13.2 Hz), 2.94 (ddd, 1 H, *J* 9.1, 9.2, 17.5 Hz), 3.64 (dd, 1 H, *J* 3.4, 4.0 Hz), 3.79 (d, 1 H, *J* 3.4 Hz), 6.20 (d, 1 H, *J* 10.2 Hz), 7.26 (dd, 1 H, *J* 4.0, 10.2 Hz); ¹³C NMR (125 MHz, CDCl₃) δ 28.1, 28.2, 47.3, 58.7, 80.1, 130.8, 145.0, 175.6, 191.1; EI-HRMS *m/z* 180.0430 (M⁺, C₉H₈O₄ requires 180.0423).