Polyclonal antibody-catalysed hydrolysis of a β-lactam

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We report the first example of antibody-catalysed hydrolysis of a β -lactam where the antibodies were generated by a simple transition-state analogue; in this example the antibodies are polyclonal.

Since the first demonstration^{1,2} of catalytic antibodies, the field has grown rapidly with large numbers of catalytic antibodies being described^{3,4} for a range of chemical reactions. However, although there is a plethora of antibodies that catalyse the hydrolysis of esters and carbonates, only a few^{5–11} have been generated that catalyse the hydrolysis of amides. Consequently the production of catalytic antibodies for amide hydrolysis is the focus of considerable current effort^{3,4,12} and success promises a wealth of applications in medicine and biotechnology.

Here we report that antibodies that were designed to catalyse the hydrolysis of a carbonate have now been found to catalyse also the hydrolysis of a structurally related amide, a β-lactam. This is the first indication that it is possible to obtain catalytic antibodies with β-lactamase activity by immunisation with a simple transition-state analogue. We have shown previously 13 that polyclonal antibodies generated by the transition state analogue 1a catalyse the hydrolysis of the carbonate 2, and these catalytic antibodies have been the subject of extensive investigation. 14,15 We work with total IgG purified 13 from sheep antisera (*via* salt fractionation, and chromatography over protein G then Sephadex), and our most recent report 16 established that a polyclonal catalytic antibody preparation contained at most 8% catalytic antibodies, and probably less than 1%.

$$O_2N$$
 O_2N
 O_2N

Current work involves studies with a variety of 4-nitroanilides and 4-nitrophenyl esters and carbonates, with the aim of determining the minimum recognition features required for catalysis by these antibody preparations. One such compound is N-(4-nitrophenyl)azetidin-2-one 3.¹⁷ This β -lactam has the following advantages as a substrate for catalytic antibody investigations: (i) it is a reactive amide whose hydrolysis is activated by both ring strain and electronic effects, (ii) the product of hydrolysis, the amino acid carboxylate 4, contains a nitroaniline chromophore which facilitates kinetic measurements, (iii) base promoted hydrolysis, in the absence of catalyst, is well characterised, 12,18 (iv) the nitrophenyl group provides recognition and binding in antibody studies, (v)

product inhibition is unlikely because the ring-opened product is very different from both the substrate and the phosphate transition state analogue, and lastly (vi) it is intermediate in reactivity between a nitrophenyl ester and a nitroanilide. Given these advantages it is surprising that this structure has not been the subject of catalytic antibody investigations until now.

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We here report that hydrolysis of the β-lactam **3** is accelerated by the anti-nitrophenylphosphate antibodies at pH values between 8 and 10. This rate acceleration is completely inhibited by the phosphate transition state analogue **1b**, demonstrating that the observed activity is due to a binding site specific for the transition state analogue **1b** (preliminary experiments give a value for K_i of 1.5μM). For the antibody preparation 270-26,¹³ (2 μM IgG, pH 9, 37 °C) a plot of initial rate for the catalysed reaction against concentration of substrate **3** is shown in Fig. 1. At this pH the catalysed reaction has $K_M = 30 \,\mu\text{M}$ and $k_{\text{cat}} = 1.3 \times 10^{-5} \,\text{s}^{-1}$, on the assumption that the concentration of catalytic sites $= 2 \times [\text{IgG}]$. If the proportion of catalytic antibodies is assumed to be $8\%^{16}$ then $k_{\text{cat}} = 1.6 \times 10^{-4} \,\text{s}^{-1}$. This antibody activity is sustained for multiple turnovers (the catalytic reaction was followed for more than 6 turnovers)

Further evidence that the catalysed hydrolysis of the β -lactam is indeed antibody-mediated was provided by investigations of substrate specificity. The hydrolyses of N-(2-nitrophenyl)azetidin-2-one $\mathbf{5}^{17}$ (the 2-substituted analogue) and 4-nitrophenylformanilide $\mathbf{6}$ were not catalysed by the catalytic antibody preparation 270-26. It is of particular interest that the formanilide $\mathbf{6}$ is more labile towards hydrolysis than the substrate β -lactam $\mathbf{3}$. One possible explanation for this observation might

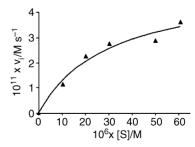


Fig. 1 Plot of initial rate against concentration of substrate 3. The points represent values obtained in initial rate experiments corrected for background hydrolysis ($k_{\rm non-cat}=6.4\times10^{-6}~{\rm s^{-1}}$), and the curve represents best fit values of $V_{\rm max}$ (5.11 ± 0.78 \times 10⁻¹¹ M s⁻¹) and $K_{\rm M}$ (30 ± 10 μ M) obtained by fitting the data to the Michaelis–Menten equation by non-linear regression using an unweighted least squares analysis ($r^2=0.974$).

be a different binding mode in the case of the formanilide such that catalysis does not occur. These data demonstrate that the catalysis is both specific for the 4-nitro substituent and is extremely limited in tolerance for alternative reaction centres, characteristics consistent with antibody catalysis.

We have demonstrated antibody catalysis of the hydrolysis of an N-aryl-β-lactam by antibodies raised to an immunogen that was not designed for this purpose. There have been only two other reports of antibody-catalysed hydrolysis of β -lactams. One of these antibodies was produced¹⁰ by an antiidiotypic approach, and the other¹¹ via reactive immunisation. Therefore, the example described above is the first report of antibodies with β-lactamase activity generated via immunisation with a simple transition-state analogue. The use of immunogens more closely related to the substrate structure (e.g. cyclic phosphonates) would be expected to produce antibodies with improved β-lactamase activity. The combination of stability ($t_{0.5} \sim 2$ weeks at pH 8) and susceptibility to antibody-catalysed hydrolysis makes N-aryl-β-lactams attractive targets for the development of prodrugs that can be activated by catalytic antibodies. For example, we are now investigating the β -lactam 7 as a potential nitrogen mustard prodrug for use in Antibody-Directed Abzyme Prodrug Therapy (ADAPT).¹⁹

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Notes and references

- A. Tramontano, K. D. Janda and R. A. Lerner, Science, 1986, 234, 1566–1570.
- 2 S. J. Pollack, J. W. Jacobs and P. G. Schultz, Science, 1986, 234, 1570–1573.
- 3 R. A. Lerner, S. J. Benkovic and P. G. Schultz, *Science*, 1991, 252, 659–667.
- 4 J. D. Stevenson and N. R. Thomas, Nat. Prod. Rep., 2000, 17, 535–577.
- 5 K. D. Janda, D. Schloeder, S. J. Benkovic and R. A. Lerner, *Science*, 1988, **241**, 1188–1191.
- 6 M. T. Martin, T. S. Angeles, R. Sugasawara, N. I. Aman, A. D. Napper, M. J. Darsley, R. I. Sanchez, P. Booth and R. C. Titmas, *J. Am. Chem. Soc.*, 1994, **116**, 6508–6512.
- 7 F. Benedetti, F. Berti, A. Colombatti, C. Ebert, P. Linda and F. Tonizzo, Chem. Commun., 1996, 1417–1418.
- 8 C. S. Gao, B. J. Lavey, C. H. L. Lo, A. Datta, P. Wentworth and K. D. Janda, J. Am. Chem. Soc., 1998, 120, 2211–2217.
- B. Gong, P. G. Schultz and S. S. Yoon, *J. Korean Chem. Soc.*, 1998, 42, 462–466.
- B. Avalle, D. Thomas and A. Friboulet, FASEB J., 1998, 12, 1055–1060.
- 11 F. Tanaka, H. Almer, R. A. Lerner and C. F. Barbas, *Tetrahedron Lett.*, 1999, **40**, 8063–8066.
- 12 G. M. Blackburn and S. X. Deng, *Biochem. Soc. Trans.*, 1993, 21, 1102–1107; O. Ersoy, R. Fleck, M. J. Blanco and S. Masamune, *Bioorg. Med. Chem.*, 1999, 7, 279; V. M. Yomtova, S. D. Kyurkchiev, N. N. Slavcheva and I. P. Ivanov, *Biocatal. Biotransform.*, 1998, 16, 307.
- 13 G. Gallacher, C. S. Jackson, M. Searcey, G. T. Badman, R. Goel, C. M. Topham, G. W. Mellor and K. Brocklehurst, *Biochem. J.*, 1991, 279, 871–881.
- 14 G. Gallacher, C. S. Jackson, M. Searcey, R. Goel, G. W. Mellor, C. Z. Smith and K. Brocklehurst, Eur. J. Biochem., 1993, 214, 197–207.
- 15 M. Resmini, R. Vigna, C. Simms, N. J. Barber, E. P. HagiPavli, A. B. Watts, C. Verma, G. Gallacher and K. Brocklehurst, *Biochem. J.*, 1997, 326, 279–287.
- 16 M. Resmini, S. Gul, S. Carter, S. Sonkaria, C. M. Topham, G. Gallacher and K. Brocklehurst, *Biochem. J.*, 2000, 346, 117–125.
- 17 Synthesised according to the method described in Hiroki Takahata, Yoshinori Ohnishi and Takao Yamazaki, *Heterocycles*, 1980, **14**(4), 467–469.
- 18 G. M. Blackburn and J. D. Plackett, *J. Chem. Soc.*, *Perkin Trans.* 2, 1973, 1366–1371; T. J. Broxton and L. W. Deady, *J. Org. Chem.*, 1974, 39, 2767; A. R. Butler, K. A. Freeman and D. E. Wright, *J. Chem. Soc.*, *Perkin Trans.* 2, 1977, 765–9.
- 19 P. Wentworth, A. Datta, D. Blakey, T. Boyle, L. J. Partridge and G. M. Blackburn, *Proc. Natl. Acad. Sci.*, 1996, 93, 799–803.