

Hammett analysis of a C–C hydrolase-catalysed reaction using synthetic 6-aryl-2-hydroxy-6-ketohexa-2,4-dienoic acid substrates

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A Hammett plot ($\rho = -0.71$) has been measured for C–C hydrolase enzyme BphD from *Pseudomonas LB400*, using six 6-aryl-2-hydroxy-6-ketohexa-2,4-dienoic acids synthesised by a Heck coupling strategy.

The bacterial degradation of aromatic compounds by soil bacteria such as *Pseudomonas spp.* commonly proceeds via the oxidative cleavage of catechol intermediates.¹ Extradiol catechol cleavage of 3-substituted catechols, catalysed by non-haem iron(II) dependent catechol 2,3-dioxygenase, yields 6-substituted 2-hydroxy-6-ketohexa-2,4-dienoic acids (see Fig. 1). NMR spectroscopic analysis has shown that 2-hydroxy-6-ketohexa-2,4-dienoic acid, the *meta* ring fission product on the phenylpropionate catabolic pathway of *Escherichia coli*, exists as the dienol tautomer in the *trans,transoid* conformation.²

This family of *meta*-ring fission products are substrates for a hydrolytic C–C cleavage reaction, catalysed by C–C hydrolases which belong to the α/β -hydrolase family (see Fig. 1).³ Crystallographic studies on C–C hydrolase BphD have revealed that this family of enzymes contain a serine–histidine–aspartate triad at their active site,⁴ yet mechanistic studies to date are not consistent with a nucleophilic mechanism, but instead favour a general base mechanism involving a *gem*-diol intermediate.^{5–7} Studies on this unusual family of enzymes are hampered by the lack of a synthetic route for the dienol substrates. In this paper we report the first total synthesis of these biological intermediates, and the kinetic evaluation via a Hammett plot of a series of synthetic substrates for C–C hydrolase BphD.

Synthesis of the diene portion of the target molecule was attempted using the palladium-catalysed Heck coupling⁸ of bromo-enol acetate **1**, prepared from ethyl 3-bromopyruvate.⁹ Palladium-catalysed couplings of **1** with phenyl vinyl ketone gave <10% yield of the coupled diene product. However, reaction of **1** with the corresponding ketal **2a** in the presence of palladium(II) acetate (0.1 equiv.), silver(I) acetate and triphenylphosphine at 80 °C cleanly gave the coupled ketal **3a** in 78% yield. Treatment with aqueous 2 M HCl gave the desired ketone **4a** in 93% yield. No other regioisomer was detected by NMR spectroscopy, thus reaction occurred selectively at the terminal β -carbon of alkene **2a**. Ketals **2b** (X = CH₃) and **2c** (X = OCH₃) were also coupled to give **4b** and **4c** in 76 and 68% yield, respectively, as shown in Scheme 1.

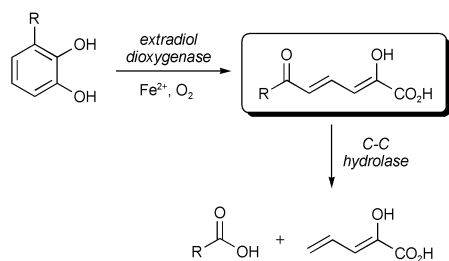


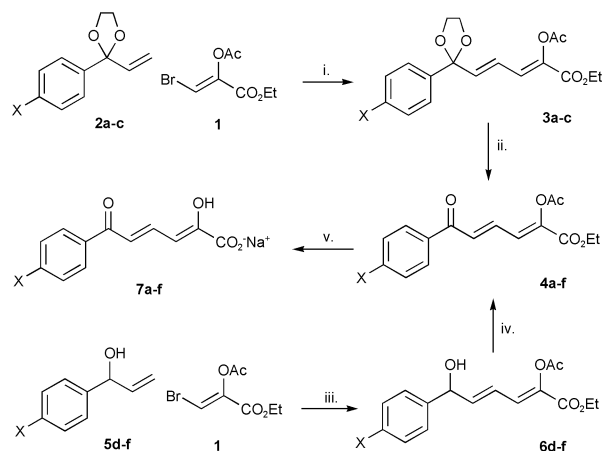
Fig. 1 Bacterial *meta*-cleavage pathway for degradation of 3-substituted catechols (R = H, CH₃, CH₂CH₂CO₂H, Ph), via extradiol catechol oxidative cleavage, to give a 6-substituted 2-hydroxy-6-ketohexa-2,4-dienoic acid, followed by C–C hydrolytic cleavage.

Aryl ketones containing electron-withdrawing substituents gave low yields of the corresponding ketal **2**. In these cases, Heck coupling was carried out using the corresponding allylic alcohols **5a** (X = H), **5d** (X = Cl), **5e** (X = NO₂) and **5f** (X = CN), synthesised by reaction of the *p*-substituted benzaldehyde with allyl magnesium bromide. Reaction of **5a** with bromide **1**, in the presence of Pd(OAc)₂/AgOAc/PPh₃, was found to give the desired alcohol **6a** as the major product, although other alkene by-products were also visible in the crude reaction product by ¹H NMR spectroscopy. The labile alcohol **6a** was oxidised to ketone **4a** using CrO₃/H₂SO₄, and isolated after chromatography in 69% overall yield. Alcohols **5d–f** were similarly coupled by this method, yielding the ketones **4d–f** in 70–80% yield.

Deprotection of di-esters **4a–f** was carried out by alkaline hydrolysis in aqueous sodium hydroxide for 2 h at room temperature, followed by neutralisation to pH 7 to give the products **7a–f** as their sodium salts in 80–90% yields. The ¹H NMR spectrum of **7b** showed signals for the diene portion at 7.07 (d, *J* 11 Hz, H-3), 7.47 (dd, *J* 15, 11 Hz, H-4) and 7.23 ppm, indicating that **7** exists in the *trans,transoid* (2-*Z*, 4-*E*) diene conformation, as found previously for an enzymatically-generated ring fission intermediate.²

C–C hydrolase BphD from the biphenyl degradation pathway of *Pseudomonas spp.* LB400^{10,11} was purified from an overexpressing strain of *E. coli*, to specific activity 3.6 u mg⁻¹. Treatment of a solution of **7a** in 50 mM potassium phosphate buffer pH 7.0 with an aliquot of BphD gave a linear decrease in absorbance at 434 nm, confirming the identity of **7a**.

Steady-state kinetic parameters were measured for the processing of synthetic substrates **7a–f** by hydrolase BphD. A plot of log(*k*_{cat}) vs. substituent parameter σ (Fig. 2) shows a decreasing rate for electron-withdrawing substituents, with reaction constant $\rho = -0.71 \pm 0.1$.[†] The two larger substituents



Scheme 1 Synthetic route for 6-aryl-2-hydroxy-6-ketohexa-2,4-dienoic acids. X = H (**a**), CH₃ (**b**), OCH₃ (**c**), Cl (**d**), CN (**e**), CF₃ (**f**). Reagents and conditions: i. Pd(OAc)₂ (0.1 equiv.), AgOAc, PPh₃, toluene, 80 °C; ii. 2 M HCl/H₂O; iii. Pd(OAc)₂ (0.1 equiv.), AgOAc, PPh₃, toluene, 80 °C; iv. CrO₃, H₂SO₄; v. NaOH, H₂O, 2 h. Yields described in text.

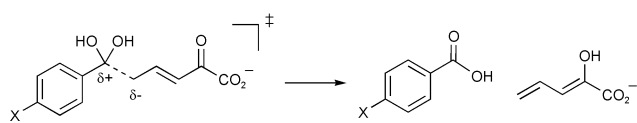
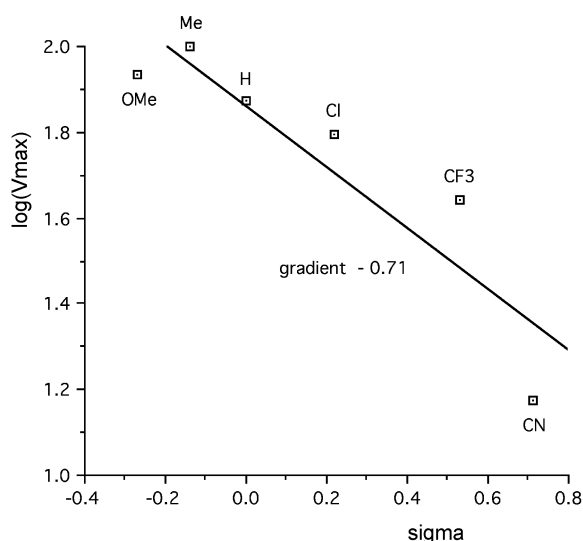


Fig. 2 Hammett plot of $\log(k_{\text{cat}})$ vs. substituent coefficient σ for processing of substrates **7a–f** by C–C hydrolase BphD. Gradient of least squares fit line $\rho = -0.71 \pm 0.1$. The scheme shows likely transition state for C–C cleavage of a *gem*-diol reaction intermediate.

OCH₃ and CN both give relatively low values of k_{cat} , probably due to steric effects upon substrate binding, but comparison of isosteric CH₃ vs. CF₃ and OCH₃ vs. CN gives a consistent pattern. These data give some insight into the transition state for C–C bond cleavage during the enzymatic reaction, implying an accumulation of δ^+ charge adjacent to the aryl ring. This is consistent with heterolytic cleavage of the C–C bond, and delocalisation of the departing δ^- charge by the adjacent α,β -unsaturated ketone. This value is opposite in sign to the ρ values

of +1.8 and +0.9 reported for serine proteases α -chymotrypsin¹² and subtilisin¹³ respectively (using substituted phenyl acetates as substrates), which proceed *via* a nucleophilic mechanism. For comparison, ρ values for base-catalysed and acid-catalysed ester hydrolysis are +2.55 and -0.57 , respectively.¹⁴

The availability of a synthetic route to this class of biological intermediates has made possible the synthesis of a range of substrates for C–C hydrolase BphD, and could be used to synthesise ring fission intermediates found on other aromatic *meta*-cleavage pathways.

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Notes and references

† The k_{cat} kinetic parameter was used, since values of K_{m} for this enzyme are extremely low ($< 1 \mu\text{M}$),¹¹ generating more experimental error in $k_{\text{cat}}/K_{\text{m}}$ data.

- 1 T. D. H. Bugg and C. J. Winfield, *Nat. Prod. Rep.*, 1998, **15**, 513.
- 2 W. W. Y. Lam and T. D. H. Bugg, *J. Chem. Soc., Chem. Commun.*, 1994, 1163.
- 3 E. Diaz and K. N. Timmis, *J. Biol. Chem.*, 1995, **270**, 6403.
- 4 N. Nandhagopal, A. Yamada, T. Hatta, E. Masai, M. Fukuda, Y. Mitsui and T. Senda, *J. Mol. Biol.*, 2001, **309**, 1139.
- 5 W. W. Y. Lam and T. D. H. Bugg, *Biochemistry*, 1997, **36**, 12242.
- 6 I. M. J. Henderson and T. D. H. Bugg, *Biochemistry*, 1997, **36**, 12252.
- 7 S. M. Fleming, T. A. Robertson, G. J. Langley and T. D. H. Bugg, *Biochemistry*, 2000, **39**, 1522.
- 8 R. F. Heck, *Org. React.*, 1982, **27**, 345.
- 9 M. D. Fryzuk and B. Bosnich, *J. Am. Chem. Soc.*, 1979, **101**, 3043.
- 10 B. Hofer, S. Backhaus and K. N. Timmis, *Gene*, 1994, **144**, 9.
- 11 S. Y. K. Seah, G. Terracina, J. T. Bolin, P. Riebel, V. Snieckus and L. D. Eltis, *J. Biol. Chem.*, 1998, **273**, 22943.
- 12 M. L. Bender and K. Nakamura, *J. Am. Chem. Soc.*, 1962, **84**, 2577.
- 13 L. T. Kanerva and A. M. Klivanov, *J. Am. Chem. Soc.*, 1989, **111**, 6864.
- 14 N. S. Isaacs, *Physical Organic Chemistry*, Longman Press, Harlow, 1987, pp. 144–146.