## First synthesis and characterization of isolable thioselenenic acid, triptycene-9-thioselenenic acid<sup>†</sup>

Akihiko Ishii,\* Takeshi Takahashi, Akira Tawata, Aki Furukawa, Hideaki Oshida and Juzo Nakayama\*

Department of Chemistry, Faculty of Science, Saitama University, Saitama, Saitama 338-8570, Japan. E-mail: ishiiaki@chem.saitama-u.ac.jp; Fax: +81 48 858 3700; Tel: +81 48 858 3394

Received (in Cambridge, UK) 9th August 2002, Accepted 11th October 2002 First published as an Advance Article on the web 25th October 2002

Triptycene-9-thioselenenic acid was synthesized by hydrolysis of acetyl triptycene-9-thioselenenate, the structure of which was determined by spectroscopic data and X-ray diffraction analysis.

Hydrodichalcogenides R-X-Y-H, where X and Y are the same or different Group 16 elements, have drawn considerable interest from various points of view. While hydroperoxides (R-O–O–H) are compounds with a long history, the chemistry of sulfenic acids (R-S-O-H)1 and selenenic acids (R-Se-O-H)2,3 are of current interest. On the other hand, hydrodisulfides (R-S-S-H)4,5 and selenosulfenic acids (R-S-Se-H)6 have been claimed to be intermediates in enzymatic reactions of cystein and selenocystein with their lyases. A hydrodisulfide was found to be a key intermediate in thiol-mediated DNA-damage by leinamycin and its model compounds.7 Thioselenenic acids, R-Se–S–H, are a member of this group and have not been isolated yet. To our knowledge, there are only two reports on the observation of Cys–SeSH [1, Cys =  $(HO_2C)(H_2N)CHCH_2$ ] by UV-vis spectroscopy<sup>8</sup> and RSeSLi (R = Bu, Ph) in THF by <sup>77</sup>Se NMR spectroscopy.<sup>8b</sup> We report herein the isolation and structure determination of triptycene-9-thioselenenic acid (2).



Acetyl triptycene-9-thioselenenate (**3**) was prepared using a method analogous to that for the synthesis of acetyl alkyl disulfides (R–S–S–Ac).<sup>9</sup> Thus, di-9-triptycyl diselenide (**4**)<sup>3c</sup> was treated with MCPBA and then with thioacetic acid to give **3**<sup>10</sup> in 91% yield; it is not essential to isolate the intermediary formed selenoseleninate (Trip–Se(O)Se–Trip<sup>3c,11</sup>). The acetyl thioselenenate **3** was hydrolyzed with 60% perchloric acid in refluxing ethanol–dichloromethane to furnish the desired thioselenenic acid **2** in 63% yield. The sulfur atom of **2** was removed readily by treatment with triphenylphosphine to give the selenol **5**<sup>3c</sup> and triphenylphosphine sulfide almost quantitatively (Scheme 1).

The structure of thioselenenic acid **2** was determined by spectroscopic data and X-ray diffraction analysis.<sup>12</sup> An ORTEP drawing is depicted in Fig. 1 with relevant bond lengths and angles data. The Se1–S1 bond length was 2.1796(9) Å, indicating the single bond character, and the C1–Se1–S1 bond angle was 103.85(7)°. The SH proton appeared at  $\delta$  2.64 in the

† Electronic supplementary information (ESI) available: experimental section. See http://www.rsc.org/suppdata/cc/b2/b207810d/



Scheme 1 Reagents and conditions: i, MCPBA,  $CH_2Cl_2$ , 0 °C; ii,  $CH_3C(O)SH$ , 0 °C, and then rt; iii,  $HClO_4$ , EtOH,  $CH_2Cl_2$ , refl., 6 h; iv, PPh<sub>3</sub>, PhH, rt, 2 h.

<sup>1</sup>H NMR spectrum, and an absorption due to the S–H stretching vibration was observed at 2520 cm<sup>-1</sup> in the IR spectrum.

Thioselenenic acid **2** was stable under acidic conditions, as is evident from the preparation conditions, whereas it decomposed to selenol **5** and a small amount of diselenenyl sulfide **6**<sup>15</sup> by treatment with triethylamine followed by neutralization<sup>4</sup>*e*</sup> [Scheme 2, (a)]; when a solution of **2** was evaporated to dryness in the presence of triethylamine, diselenenyl sulfide **6** and diselenide **4** were formed [Scheme 2, (b)]. Treatment of **2** with triphenyltin chloride in the presence of triethylamine yielded stannyl thioselenenate **7**<sup>16</sup> and stannyl selenide **8**<sup>16</sup> [Scheme 2, (c)].

Polidoro and coworkers reported that treatment of  $(Cys-Se)_2$ with Na<sub>2</sub>S in an aqueous NaOH solution  $1(1 \times 10^{-3} \text{ M})$ generated Cys-Se-SH that exhibited an absorption maximum at 375 nm in the UV-vis spectrum.<sup>8</sup> Fig. 2 shows UV-Vis spectra of thioselenenic acid **2** and diselenenyl sulfide **6** in dichloro-



**Fig. 1** ORTEP drawing of triptycene-9-thioselenenic acid (2) (50% ellipsoidal probability). Relevant bond lengths (Å) and angles (deg) data: Se1–C1 1.975(2); Se1–S1 2.1796(9); C1–C14 1.536(3); C1–C2 1.537(3); C1–C15 1.538(3); S1–H1 1.24(6); C1–Se1–S1 103.85(7); C14–C1–C2 106.19(17); C14–C1–C15 105.15(17); C2–C1–C15 105.31(17); C14–C1–Se1 115.62(15); C2–C1–Se1 115.27(15); C15–C1–Se1 108.37(15).



**Scheme 2** Reagents and conditions: i,  $Et_3N$ , PhH, rt, 5 min; ii, aq. NH<sub>4</sub>Cl; iii, evaporation in the presence of  $Et_3N$ ; iv, Ph<sub>3</sub>SnCl,  $Et_3N$ , PhH, rt, 1 h. Yields are calculated from <sup>1</sup>H NMR integral ratios.



Fig. 2 UV-vis spectra of triptycene-9-thioselenenic acid (2) and di-(triptycene-9-selenenyl) sulfide (6).

methane. The absorption of **2** ceased approximately at 360 nm, while **6** has a broad absorption from 285 to 360 nm with the molecular absorption coefficient  $(\varepsilon/10^3 \text{ cm}^2 \text{ mol}^{-1}) \approx 1000$ . Anyway, we did not observe an explicit absorption maximum around 375 nm. This might be attributed to the contrasting character of the substituents: the more sterically demanding, hydropholic 9-tripycyl group and the much less sterically demanding, hydrophilic 2-amino-3-propionyl (Cys) group. We are therefore now investigating the preparation of thioselenenic acids having groups derived from the Cys group or alkyl substituents structurally simpler than the 9-triptycyl group.

## Notes and references

- A. Claiborne, J. I. Yeh, T. C. Mallett, T. J. Luba, E. J. Crane III, V. Charrier and D. Parsonage, *Biochemistry*, 1999, 38, 15407.
- 2 (a) H. E. Ganther, *Chem. Scr.*, 1975, **8A**, 79; (b) G. Mugesh and W.-W. du Mont, *Chem. Eur. J.*, 2001, **7**, 1365.
- 3 (a) K. Goto and R. Okazaki, *Liebgs Ann./Recueil*, 1997, 2393; (b) K. Goto, M. Nagahama, T. Mizushima, K. Shimada, T. Kawashima and R. Okazaki, *Org. Lett.*, 2001, 3569; (c) A. Ishii, S. Matsubayashi, T. Takahashi and J. Nakayama, *J. Org. Chem.*, 1999, **64**, 1084, and references cited therein.
- 4 (a) J. Tsurugi and T. Nakabayashi, J. Org. Chem., 1959, 24, 807; (b) T. Nakabayashi, J. Tsurugi and T. Yabuta, J. Org. Chem., 1964, 29, 1236; (c) S. Kawamura, T. Kitao, T. Nakabayashi, T. Horii and J. Tsurugi, J. Org. Chem., 1968, 33, 1179; (d) J. Tsurugi, Y. Abe and S. Kawamura, Bull. Chem. Soc. Jpn., 1970, 43, 1890; (e) J. Tsurugi, Y. Abe, T. Nakabayashi, S. Kawamura, T. Kitao and M. Niwa, J. Org. Chem., 1970, 35, 3263; (f) D. N. Harpp, D. K. Ash, T. G. Back and J. G. Gleason, Tetrahedron Lett., 1970, 3551; (g) S. Kawamura, T. Horii, T. Nakabayashi and M. Hamada, Bull. Chem. Soc. Jpn., 1975, 48, 2993; (h) N. E. Heimer, L. Field and R. A. Neal, J. Org. Chem., 1981, 46, 1374; (i) N. E. Heimer and L. Field, J. Org. Chem., 1985, 50, 4164; (k) G.

Derbesy, D. N. Harpp, B. Rather and G. Carroll, *Sulfur Lett.*, 1992, 35, 199.

- 5 (a) D. Cavallini, C. de Marco, B. Mondovì and B. G. Mori, Enzymologia, 1960, 22, 161; (b) J. Loiselet and F. Chatagner, Bull. Soc. Chim. Biol., 1966, 48, 595; (c) D. Cavallini, G. Federici, E. Barboni and M. Marcucci, FEBS Lett., 1970, 10, 125; (d) D. Cavallini, G. Federici and E. Barboni, Eur. J. Biochem., 1970, 14, 169; (e) A. Finazzi Agrò, G. Federici, C. Giovagnoli, C. Cannella and D. Cavallini, Eur. J. Biochem., 1972, 28, 89; (f) G. Federici, S. Duprè, R. M. Matarese, S. P. Solinas and D. Cavallini, Int. J. Pept. Protein Res., 1977, 10, 185; (g) T. Fujii, M. Maeda, H. Mihara, T. Kurihara, N. Esaki and Y. Hata, Biochemistry, 2000, 39, 1263.
- 6 (a) C. Cannella, L. Pecci, A. Finazzi Agrò, G. Federici, B. Pensa and D. Cavallini, *Eur. J. Biochem.*, 1975, **55**, 285; (b) G. M. Lacourciere and T. C. Stadtman, *J. Biol. Chem.*, 1998, **273**, 30921. See also ref. 5g.
- 7 (a) S. J. Behroozi, W. Kim and K. S. Gates, J. Org. Chem., 1995, 60, 3964; (b) K. Mitra, W. Kim, J. S. Daniels and K. S. Gates, J. Am. Chem. Soc., 1997, 119, 11691; (c) K. S. Gates, Chem. Res. Toxicol., 2000, 13, 953, and references cited therein.
- 8 (a) G. Polidoro, P. De Cicco, S. Di Luzio and G. Federici, *Boll. Soc. Ital. Biol. Sper.*, 1979, **55**, 2346; (b) C. Köllemann, D. Obendorf and F. Sladky, *Phosphorus, Sulfur Relat. Elem.*, 1988, **38**, 69.
- 9 F. Freeman, B.-G. Huang and R. I.-S. Lin, *Synthesis*, 1994, 699.
  10 3: mp 177–180 °C (CH<sub>2</sub>Cl<sub>2</sub>–hexane). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ
- 10 3: mp 17/-180 °C (CH<sub>2</sub>Cl<sub>2</sub>-nexane). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\sigma$ 2.50 (s, 3H), 5.37 (s, 1H), 6.98–7.05 (m, 6H), 7.35–7.40 (m, 3H), 7.47–7.51 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz)  $\delta$  29.8, 54.1, 64.9, 123.5, 123.7, 125.0, 125.9, 144.1, 145.2, 192.1; IR (KBr) 1700 (C=O) cm<sup>-1</sup>. Anal. Calc. for C<sub>22</sub>H<sub>16</sub>OSSe: C, 64.86; H, 3.96. Found: C, 64.80; H, 3.92%.
- 11 In this one-pot reaction, we have prepared  $(1-Ad)CH_2SeSAc$  and  $(1-Ad)CH(CH_3)SeSAc$  (1-Ad = 1-adamantyl).
- 12 2: pale yellow plates, mp 170-172 °C decomp (CH<sub>2</sub>Cl<sub>2</sub>-hexane). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ2.64 (s, 1H, SH), 5.38 (s, 1H), 7.00–7.08 (m, 6H), 7.36-7.42 (m, 3H), 7.49-7.54 (m, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz) δ 54.0, 61.1, 123.6, 123.8, 125.0, 125.7, 144.1, 145.8; IR (KBr) 2520 cm-1. Anal. Calc. for C20H14SSe: C, 65.75; H, 3.86. Found: C, 65.73; H, 3.89%. Crystal data: C<sub>20</sub>H<sub>14</sub>SSe, M<sub>w</sub> 365.36, pale yellow plate,  $0.24 \times 0.16 \times 0.08$  mm<sup>3</sup>, triclinic, space group  $P\overline{1}$ , a = 8.181(2),  $\dot{b} = 8.217(1), c = 13.175(3)$  Å,  $\alpha = 82.53(1), \beta = 72.58(1), \gamma = 10.175(3)$  $67.82(1)^{\circ}$ , V = 782.4(2) Å<sup>3</sup>, Z = 2,  $\rho_{calc} = 1.551$  g cm<sup>-3</sup>,  $\mu$ (CuK $\alpha$ ) = 4.42 mm<sup>-1</sup>. Mac Science MXC3KHF diffractometer with graphitemonochromated CuK $\alpha$  radiation ( $\lambda = 1.54178$  Å),  $\theta/2\theta$  scans method in the range  $3^{\circ} < 2\theta < 140^{\circ}$  ( $-9 \leq h \leq 9, 0 \leq k \leq 10, -15 \leq l \leq 16$ ), 2994 independent reflections. The structure was solved with direct methods (SIR9213), and refined with full-matrix least-squares (SHELXL-9714) using all independent reflections for 256 parameters. Absorption correction was done by a psi-scan method. The nonhydrogen atoms were refined anisotropically: R1 = 0.0303 ( $I > 2\sigma(I)$ , 2822 reflections), wR2 = 0.0860 (for all), GOF = 1.05; max/min residual density = 0.454/-0.452 e Å<sup>-3</sup>. CCDC reference number 191691. See http://www.rsc.org/suppdata/cc/b2/b207810d/ for crystallographic data in CIF or other electronic format.
- 13 A. Altomare, G. Cascarano, C. Giacovazzo and A. Guagliardi, J. Appl. Crystallogr., 1993, 26, 343.
- 14 G. M. Sheldrick, SHELXL-97, Program for Crystal Structure Refinement, Göttingen University, Germany, 1997.
- 15 6: pale yellow crystals, mp 345–346 °C decomp (hexane–CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz)  $\delta$  5.46 (s, 2H), 7.11 (pseudo d of quintet, J = 1.5, 7.4 Hz, 12H), 7.46 (dd, J = 1.5, 7 Hz, 6H), 7.76 (dd, J = 1.5, 8 Hz, 6H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz)  $\delta$  54.2, 65.2, 123.7, 124.4, 125.1, 125.9, 144.7, 145.6. Anal. Calc. for C<sub>40</sub>H<sub>22</sub>SSe<sub>2</sub>: C, 68.96; H, 3.76. Found: C, 68.31; H, 3.68%.
- 16 The structures of 7 and 8 were determined by X-ray crystallography. CCDC reference numbers 191692 (7) and 191693 (8). 7: yellow crystals, mp 176-178 °C decomp (Et<sub>2</sub>O-hexane). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 200 MHz)  $\delta 5.29$  (s, 1H), 6.74 (dt, J = 1, 8 Hz, 3H), 6.93 (dt, J = 1, 7 Hz, 3H), 7.30 (dd, J = 1, 7 Hz, 3H), 7.43–7.55 (m, 12H), 7.74–7.84 (m, 6H, accompanying satellite signals, J = 55 Hz, due to <sup>117</sup>Sn and <sup>119</sup>Sn); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 75.5 MHz) δ 54.0, 61.7, 123.1, 124.6, 124.8, 125.4, 129.1, 130.2, 137.0, 137.1, 144.4, 145.4. Anal. Calc. for C<sub>38</sub>H<sub>28</sub>SSeSn: C, 63.89; H, 3.95. Found: C, 63.71; H, 3.90. 8: colorless cubes, mp 253-255 °C (hexane-CH<sub>2</sub>Cl<sub>2</sub>). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 5.28 (s, 1H), 6.64 (dt, J = 1, 8 Hz, 3H), 6.87 (dt, J = 1, 8 Hz, 3H), 7.20-7.34 (m, 12H), 7.40 (dd, J = 1, 8 Hz, 6H, accompanying satellite signals, J= 55 Hz, due to <sup>117</sup>Sn and <sup>119</sup>Sn), 7.79 (d, J = 7.5 Hz, 3H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100.6 MHz) δ 54.1, 61.7, 122.6, 124.5, 125.36, 125.41, 128.6, 129.4, 136.8, 138.2, 144.8, 146.4. Anal. Calc. for C38H28SeSn: C, 66.89; H, 4.14. Found: C, 66.65; H, 4.04%.