

Unusual structure of the dimeric 4-bromocalcimycin–Zn²⁺ complexStéphane Vila,^a Isabelle Canet,^a Jacques Guyot,^a Georges Jeminet^{*a} and Loïc Toupet^b^a Université Blaise Pascal Clermont-Ferrand and CNRS, UMR 6504, Campus des Cézeaux, 63177 Aubière Cedex, France. E-mail: G. Jeminet@univ-bpclermont.fr; Fax: 33 047340 7717^b Université de Rennes I and CNRS, UMR 6626, Campus de Beaulieu, avenue du général Leclerc, 35042 Rennes Cedex, France

Received (in Cambridge, UK) 18th October 2002, Accepted 10th January 2003

First published as an Advance Article on the web 22nd January 2003

The X-ray structure of [Zn(4-bromocalcimycin)₂·H₂O] complex shows two highly different conformations of the ligand in the dimeric association, unusual in this ionophore family.

Since its discovery, calcimycin or A23187¹ **1** (Fig. 1, a carboxylic polyether antibiotic with calcium carrier properties, has attracted considerable attention in biology as a tool for the study of calcium second messenger in living systems.² Its non-fluorescent 4-bromo derivative **2** was subsequently described and found suitable for the same application in the presence of fluorescent probes.³

Erdahl and colleagues⁴ recently showed that **2** transports Zn²⁺ and Mn²⁺ with high selectivity over Ca²⁺ in phospholipid vesicles, and they made interesting findings concerning the stoichiometry of species involved in the transport. However, information on the structure of cation complexes with this specific ligand were lacking.

We recently undertook the preparation of crystalline adducts of **2** with various divalent cations suitable for X-ray analysis. Here we report the crystal structure obtained for a cation of topical biological interest, Zn²⁺, which reveals unusual features. To obtain the complex, we used a H₂O–CH₂Cl₂ biphasic system in which the free acid dissolved in the organic layer was stirred with an aqueous Zn(ClO₄)₂ solution at pH 10.[†]

The crystal structure determination by X-ray diffraction was consistent with the neutral dicarboxylate complex [Zn(4-bromocalcimycin)₂·H₂O] (Fig. 2).[‡] The zinc atom occupied the center of a slightly distorted octahedron (Table 1). Coordinations were provided by O(7) of a water ligand, O(1), N(2) and O(31), N(32) belonging respectively to the L1 and L2 benzoxazole-carboxylate moiety. Interestingly, as shown in Fig. 3 obtained from the crystal data, the conformation of L1 was such that O(6) of the ketopyrrole arm supplied the sixth liganding site, forming a complexing tripod. L1 and L2 were clearly not equivalent in the complex structure. The L2 benzoxazole ring is rotated by nearly 180° compared with L1, and the ketopyrrole arm is unfavourably positioned for participation in the scaffold (Fig. 2).

Octahedral arrangements of the same type were recently described for Zn(II) complexes containing more simple aromatic moieties with nitrogen and carboxylate coordinating sites, but no such unsymmetrically bound ligand with a bidentate/tridentate arrangement has been described.⁵

The specific conformation adopted by L1 and L2 did not permit the two head-to-tail intermolecular chelations observed in the well-known calcimycin dimeric complexes^{6–8} described

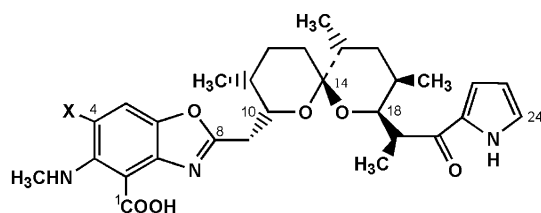
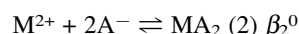
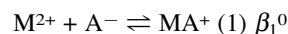


Fig. 1 X = H, calcimycin **1**; X = Br, 4-bromocalcimycin **2**.

for Ca²⁺, Mg²⁺ and Fe²⁺. One intermolecular hydrogen chelation remained for N(3)–H···O(31) and N(3)–H···O(32) (Table 1).

Also, the 3-aminomethyl substituent which was NH-chelated with the carboxylate group for calcimycin^{6–8} was moved out of the aromatic plane owing to the presence of the bulky bromine in the 4 position, and the existing intramolecular hydrogen bonding was thus suppressed. We show that this steric hindrance has repercussions on the complexing properties for divalent cations, the 1:1 and 1:2 thermodynamic stability constants β_1^0 and β_2^0 in MeOH (25 °C) :



(A[−], carboxylate form of **1** or **2**; M²⁺ divalent cation) being lowered as follows for Zn²⁺:⁹

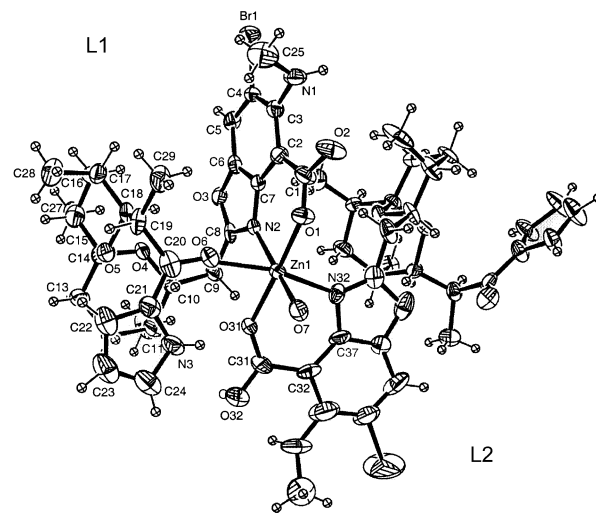


Fig. 2 ORTEP drawing of [Zn(4-bromocalcimycin)₂·H₂O] crystal structure with ligands L1 and L2 nomenclature.

Table 1 Selected bond lengths (Å) and angles (°)

Bond lengths		Angles	
Zn(1)–O(7)	2.103(6)	O(1)–Zn(1)–O(7)	88.3(2)
Zn(1)–O(31)	2.013(6)	O(7)–Zn(1)–N(2)	171.6(3)
Zn(1)–N(32)	2.147(8)	O(31)–Zn(1)–O(1)	174.2(3)
Zn(1)–O(1)	2.030(6)	N(32)–Zn(1)–O(6)	171.8(3)
Zn(1)–N(2)	2.147(7)	O(1)–Zn(1)–N(2)	85.5(3)
Zn(1)–O(6)	2.234(7)	O(1)–Zn(1)–O(6)	89.5(3)
		O(1)–Zn(1)–N(32)	98.5(3)
		O(31)–Zn(1)–N(32)	85.6(3)
		O(31)–Zn(1)–O(6)	86.3(3)

Intermolecular distances

N(3)–H···O(31)	2.123
N(3)–H···O(32)	2.451

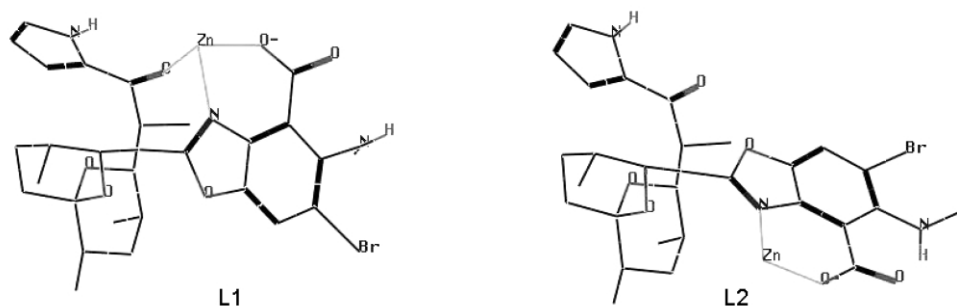


Fig. 3 A representation of L1 and L2 ligands structure coordinated with Zn²⁺ showing the specific benzoxazole rotation in L2.

calcimycin **1**: $\log \beta_1^0 = 8.24$, $\log \beta_2^0 = 18.34$

4-bromocalcimycin **2**: $\log \beta_1^0 = 6.72$, $\log \beta_2^0 = 13.40$

Interestingly, for both L1 and L2 the ketopyrrole arm adopted the preferential orientation already observed in the solid^{6–8} and liquid state¹⁰ for calcimycin with, in particular, the characteristic antiperiplanar position of H18 and H19.

All these observations suggest that in the complex formation, a 1 : 1 (L1–Zn⁺) association occurs followed by the approach of the second ligand L2. The latter is unable to displace the remaining water ligand by its ketone function to give a final 1 : 2 supramolecular association. The molecular arrangement is thus not well designed towards the solvent attack in the reverse step. This may be related to the transport experiments in vesicles⁴ where the authors state that Zn²⁺ is transported, in part, as a 1 : 1 complex in a pH-dependent stoichiometry.

In conclusion, we describe here what is to our knowledge the first solid state structure obtained with 4-bromocalcimycin. Furthermore, the Zn²⁺ complex studied shows an unusual supramolecular scaffold compared with known calcimycin Ca²⁺, Mg²⁺ and Fe²⁺ dimeric arrangements and with other described carboxylate zinc complexes.

As recently stated,¹¹ zinc homeostasis studies in higher animals and humans have proved difficult. This work may therefore help gain a better understanding of the mechanism of cellular transport of zinc through biological membranes.

This work was supported by the C.N.R.S. and the Ministère de l'Éducation Nationale, de la Recherche et de la Technologie (S.V. is grateful for a 'bourse de thèse M.E.N.R.T.').

Notes and references

† *Preparation*: A solution of 4-bromocalcimycin free acid, prepared from calcimycin as described¹² (100 mg, 0.17 mmol) in CH₂Cl₂ (17 mL) was stirred with 0.1 M aqueous Zn(ClO₄)₂ (50 mL). The pH was adjusted to ca. 10 by addition of tetrabutylammonium hydroxide. The mixture was stirred at 20 °C for 4 h under argon in the dark. The organic layer was then dried over anhydrous Na₂SO₄, filtered and evaporated. The solid residue was dissolved in EtOH (96.2 °C, Carlo Erba) and the solvent was left to evaporate at 20 °C for 1 week in the dark. Pale yellow crystals obtained proved suitable for X-ray analysis.

‡ *Structure analysis*: ZnBr₂N₆O₁₃C₅₈H₇₂·C₂H₅OH·H₂O, *M*_r = 1350.49, tetragonal, *P*₄₃₂₁, *a* = *b* = 15.4503(1), *c* = 52.1008(4) Å, *V* = 12437.07(15) Å³, *Z* = 8, *D*_x = 1.442 Mg m⁻³, λ(MoKα) = 0.71073 Å, μ = 17.50 cm⁻¹, *F*(000) = 5616, *T* = 110 K. The sample (0.35 × 0.22 × 0.20 mm) was studied on a NONIUS Kappa CCD with graphite monochromatized MoKα radiation. The cell parameters were obtained with Denzo and Scalepack¹³ with 10 frames (psi rotation: 1° per frame). The data collection¹⁴ (2θ_{max} = 60°, 1752 frames via 0.4° omega rotation and 28 s per frame, range *hkl*: *h* –19,19; *k* –13,13; *l* –52,66) gave 22439 reflections. The data reduction with Denzo and Scalepack¹³ led to 13040 independent reflections of which 7989 with *I* > 2.0σ(*I*). The structure was solved with SIR-97¹⁵ which revealed the non hydrogen atoms. After anisotropic refinement, many hydrogen atoms may be found with a Fourier Difference. The whole structure was refined with SHELXL97¹⁶ by the full-matrix least-

square techniques (use of *F* square magnitude; *x*, *y*, *z*, β_{ij} for Zn, Br, N, O and C atoms, *x*, *y*, *z* in riding mode for H atoms; 768 variables and 7989 observations with *I* > 2.0σ(*I*); calc. *w* = 1/[σ²(*F*_o)² + (0.129*P*)² + 59*P*] where *P* = (*F*_o² + 2*F*_c²)/3 with the resulting *R* = 0.097, *R*_w = 0.240 and *S*_w = 1.023 (residual around solvent molecules) Δρ < 2.5 e Å⁻³. The whole structure consists of the dimeric complex which crystallises with an ethanol molecule and a water molecule near a pyrrole ring. CCDC 195727. See <http://www.rsc.org/suppdata/cc/b2/b210280n/> for crystallographic files in CIF or other electronic format.

Atomic scattering factors were from International Tables for X-ray Crystallography¹⁷ and Ortep views realized with PLATON98.¹⁸

- M. O. Chaney, P. V. Demarco, N. D. Jones and J. L. Occolowitz, *J. Am. Chem. Soc.*, 1974, **96**, 1932.
- See for instance: G. A. Woolley, D. R. Pfeiffer and C. M. Deber, *Methods in Neurosciences*, Academic Press, Orlando, 1995, vol. 27, 52–68.
- C. M. Deber, J. T. Kun, E. Mack and S. Grinstein, *Anal. Biochem.*, 1985, **146**, 349.
- W. L. Erdahl, C. J. Chapman, E. Wang, R. W. Taylor and D. R. Pfeiffer, *Biochemistry*, 1996, **35**, 13817.
- K. Prout, A. Edwards, V. Mtetwa, J. Murray, J. F. Saunders and F. J. C. Rossotti, *Inorg. Chem.*, 1997, **36**, 2820; T. A. Zevaco, H. Görls and E. Dinjus, *Inorg. Chim. Acta*, 1998, **269**, 283; N. Okabe and N. Oya, *Acta Crystallogr., Sect. C*, 2000, **56**, 305; H. Necefoglu, W. Clegg and A. J. Scott, *Acta Crystallogr., Sect. E*, 2001, **57**, 465; J.-C. Daran, P. Lemoine and B. Viossat, *Acta Crystallogr., Sect. C*, 2002, **58**, 210; D. J. Darensbourg, J. R. Wildeson and J. C. Yarbrough, *Inorg. Chem.*, 2002, **41**, 973.
- G. D. Smith and W. L. Duax, *J. Am. Chem. Soc.*, 1976, **98**, 1578.
- M. Alléaume and Y. Barrans, *Can. J. Chem.*, 1985, **63**, 3482.
- E. Baker, E. N. Maslen, K. J. Watson and A. H. White, *J. Am. Chem. Soc.*, 1984, **106**, 2860.
- S. Vila, *thèse de doctorat*, Université Blaise Pascal de Clermont-Ferrand, N° d'Ordre: 1256, 2002. Values were determined by potentiometry. To be published in detail.
- A.-M. Albrecht-Gary, S. Blanc-Parasote, D. W. Boyd, G. Dauphin, G. Jeminet, J. Juillard, M. Prudhomme and C. Tissier, *J. Am. Chem. Soc.*, 1989, **111**, 8598.
- E. D. Harris, *Nutr. Rev.*, 2002, **60**, 121.
- M. Debono, R. M. Molloy, D. E. Dorman, J. W. Paschal, D. F. Babcock, F. Donner, C. M. Deber and D. R. Pfeiffer, *Biochemistry*, 1981, **20**, 6865.
- Z. Otwinowski and W. Minor, *Processing of X-ray Diffraction Data Collected in Oscillation Mode*, in *Methods in Enzymology, Macromolecular Crystallography, Part A*, ed. C. W. Carter and R. M. Sweet, Academic Press, London, 1997; vol. 276, pp. 307.
- Nonius (1999), *KappaCCD Software*, Nonius BV, Delft, The Netherlands.
- A. Altomare, M. C. Burla, M. Camalli, G. Cascarano, C. Giacovazzo, A. Guagliardi, A. G. G. Moliterni, G. L. Polidori and R. Spagna, *J. Appl. Crystallogr.*, 1999, **32**, 115–119.
- A. L. Sheldrick, (1997).SHELX97. Program for the Refinement of Crystal Structures, Univ. of Göttingen, Germany.
- International Tables for X-ray Crystallography*, (1992). vol. C. Edn A. J. C. (Kluwer Academic Publishers, Dordrecht).
- A. L. Spek, (1998) PLATON. A multipurpose crystallographic tool, Utrecht University, Utrecht, The Netherlands.