Double discrimination by binding and reactivity in fluorescent metal ion detection

Andriy Mokhir* and Roland Krämer

Received (in Cambridge, UK) 7th January 2005, Accepted 1st March 2005 First published as an Advance Article on the web 10th March 2005 DOI: 10.1039/b500232j

Metal ion detection using a fluorescent dye containing reactive ester groups allows discrimination on the basis of the hydrolytic reactivities of metal ions, which display the same qualitative fluorescence response on binding only.

Analysis of metal ions (M^{n+}) using fluorescent molecular sensors attracts a lot of attention, because this method is very sensitive, quick, does not require expensive instrumentation and can be performed in a high-throughput format.¹ The sensors usually combine a chelating ligand and a fluorophore, the emission of which is affected by metal ion coordination.² Responses depend on the thermodynamic stabilities of the corresponding M^{n+} complexes, which means that the effect of the metal ions is concentration dependent. For example, fluorescence emission of commercially available metal ion sensors, Phen Green SK, FL,³ Calcein⁴ and Fura-2,⁵ is quenched by Cu²⁺ at >µM concentrations. Analogous quenching effects can, however, be achieved by other paramagnetic metal ions, *e.g.* Ni²⁺, in more concentrated solutions, which means that samples of unknown composition cannot be unambiguously analyzed.

Here we present a dye, whose fluorescence emission is affected not only by metal ion binding, but additionally by a metal ion promoted reaction. If the former process is faster than the latter one, two parameters can be obtained from every analyte in a single experiment. These are a quick fluorescence change corresponding to the metal ion binding (F/F_0) and a slower change corresponding to the chemical reaction (dF/dt). In comparison, only the parameter F/F_0 is obtained from standard sensors.¹⁻⁴ This increased amount of information should allow discrimination of metal ions, which qualitatively effect the same response of the fluorophore (i.e. either fluorescence quenching or increase). In this paper we present **D1**, the first hybrid sensor⁶ of this type (Scheme 1). It is based on a Calcein framework, the iminodiacetate groups of which are esterified. A carboxylic group introduced at the 5(6)-position increases the solubility of D1 in water. The dye was synthesized from 2',7'-dichlorofluorescein-3'6'-diacetate-5(6)-carboxylate⁷ and diethyliminodiacetate using the protocol reported for similar derivatives.^{†8} After addition of a large excess of Cu²⁺, Ni²⁺, Fe³⁺ or Eu³⁺ to D1 its fluorescence is immediately quenched $(F/F_0 \sim 0)$, which indicates that the complexes formed in these solutions are not fluorescent in analogy with the complexes of the parent compound Calcein. In accordance with the structure of D1, mono and binuclear complexes can potentially be formed. However, in ESI mass spectra of these solutions no peaks corresponding to binuclear complexes $D1-(M^{n+})_2$ are present,



Scheme 1 Structure of the reactive sensor D1. Binding of M^{2+} to D1 and hydrolysis of the latter.

whereas peaks corresponding to mononuclear $D1-M^{n+}$ complexes can be detected. This suggests that $D1-M^{n+}$ is the only species in these solutions. Stability constants K_d of $D1-M^{n+}$ complexes have been determined by fluorimetry (Table 1). The affinity of D1 is considerably higher towards the Cu²⁺ ion than the other metal ions (Table 1). However, the metal ion discrimination based solely on the binding event could be masked by increased concentrations of the weaker binding M^{n+} ions. For example, Cu²⁺ (20 μ M) or 50 times more concentrated Ni²⁺, Fe³⁺ or Eu³⁺ produce similar immediate quenching (*F*/*F*₀) of the fluorescence of D1 (Fig. 1, Table 1).

^{*}andriy.mokhir@urz.uni-heidelberg.de

Table 1 Changes of fluorescence emission intensity of **D1** upon addition of different metal ions and thermodynamic parameters of some $D1-M^{n+}$ complexes

M^{n+}	$[M^{n+}]_{total}/mM$	F/F_0	$(\mathrm{d}F/\mathrm{d}t)_0$	$K_{\rm d}({\rm D1-M^{n+}})/10^6 {\rm M^{-1}}$
Cu ²⁺	0.02	0.42	-7.12	12 ± 3
Ni ²⁺	0.97	0.39	-0.42	601 ± 20
Fe ³⁺	0.97	0.39	-0.55	522 ± 90
Eu ³⁺	1.2	0.40	-2.54	497 ± 550^{a}
Zn^{2+}	1	1.07	0.30	
Ca ²⁺	1	0.94	~ 0	
Mg ²⁺	1	0.98	~ 0	
K^+	5	0.98	~ 0	—

^{*a*} The high standard deviation of the determined K_d may indicate formation of complexes other than D1–M^{*n*+} in solutions containing excess Eu³⁺.

The reactive ester group of D1-Cu2+ is hydrolyzed with formation of the carboxylate complex of D2 (Scheme 1). It follows from the ESI mass spectral study that only a single ester group of D1 is hydrolyzed (Fig. 2) even after prolonged reaction time. Since the carboxylic group of **D2** is a better donor than the parent ester group, metal complexes of D2 are expected to be more stable than those of **D1**. This has been confirmed for Cu^{2+} : $K_d(D1 Cu^{2+}/K_d(D2-Cu^{2+}) = 3.2$. This effect is reflected in decreasing intensity of the overall fluorescence emission with time upon D1 hydrolysis in the presence of a metal ion concentration which does not saturate the chelating site (Fig. 1). The initial rate of D1 hydrolysis is proportional to the linear part of the dependence of fluorescence emission intensity vs. time $(dF/dt)_0$ (Fig. 1, Table 1). The latter values have been used for estimation of activities of metal ions in promoting D1 hydrolysis, assuming that metal ion affinity to the fluorophore is significantly increased by ester hydrolysis. It is well documented that metal ions have different abilities to activate ester groups toward hydrolysis.9 On the basis of $(dF/dt)_0$ values one can clearly distinguish Cu²⁺ (20 μ M), Ni²⁺, Fe^{3+} and Eu^{3+} (about 1 mM), which give very similar immediate fluorescence quenching F/F_0 . Hydrolytic activities of selected metal ions follow the given trend: $Cu^{2+} > Eu^{3+} > Fe^{3+} > Ni^{2+} > Zn^{2+} \gg$ Ca^{2+} , Mg^{2+} , K^+ . In the case of Zn^{2+} the fluorescence of D1 is



Fig. 1 Dependence of normalized fluorescence (F/F_0) on time after addition of Ni²⁺ (1 mM, trace 1) or Cu²⁺ (20 μ M, trace 2). [D1] = 1 μ M; MOPS 10 mM, pH 7; NaCl 50 mM; solvent: water with 1% DMSO.



Fig. 2 ESI mass spectra of solution of D1 ($20 \ \mu$ M), CuSO₄ ($20 \ equiv.$) in CH₃CN–H₂O (1 : 1), titrated with NEt₃ to pH 7: (1) immediately after CuSO₄ addition; (2) 25 min after CuSO₄ addition; (3) 60 min after CuSO₄ addition; (4) 90 min after CuSO₄ addition.

enhanced by both binding and ester cleavage by the metal ion, as expected in view of the 1.5 fold increase of Calcein fluorescence at saturation with Zn^{2+} .

Metal ion promoted reactions which affect the fluorescence of a reactant have been used earlier for Cu^{2+} detection by determination of end-point fluorescence intensity (chemodosimetry).¹⁰ Indirect analysis of the same metal ion by kinetic methods has been achieved by replacement/release of a catalytically active Pd²⁺ ion from a macrocyclic complex upon exposure to $Cu^{2+,11}$ Both methods are Cu^{2+} specific and do not allow a kinetic discrimination of different metal ions.

In conclusion, the fluorescence of a dye which is modified with a "reactive" chelating group is affected by both binding and subsequently in a time-dependent manner by the hydrolytic reactivity of a metal ion. The additional kinetic parameter dF/dt obtained with this sensor type allows to discriminate metal ions (Cu²⁺, Ni²⁺, Fe³⁺, Eu³⁺) which give the same immediate fluorescence response and are not readily distinguished by a single sensor that relies on metal ion binding only.

Andriy Mokhir* and Roland Krämer

Universität Heidelberg, Anorganisch-Chemisches Institut, Im Neuenheimer Feld, 270, Heidelberg, Germany. E-mail: andriy.mokhir@urz.uni-heidelberg.de; Fax: +49 6221 548439; Tel: +49 6221 548441

Notes and references

[†] Synthesis of **D1**: paraformaldehyde (0.48 g, 16 mmol) and diethyliminodiacetate (1.51 g, 8 mmol) were added to acetonitril (50 mL) and the suspension obtained was heated to reflux for 90 min. 2',7'-dichlorofluorescein-3'6'-diacetate-5(6)-carboxylate pyridinium salt (0.98 g, 1.6 mmol) in acetonitril-water (1 : 1, 50 mL) was added and the resulting solution was left stirring for 24 h. After cooling the reaction mixture CH₃CN was removed on a rotory evaporator and the aqueous solution was acidified with acetic acid to pH 4, cooled down to 4 °C and left standing at this temperature for 2 h. The precipitate formed was filtered, washed with cold water and dried at 0.01 mbar. The product was purified by silica gel chromatography using a CHCl₃-EtOH (9 : 1) mixture containing 0.1% AcOH. The resulting bright orange solid is a mixture of two isomers in ~4 : 1 molar ratio (6 : 5-isomers). Yield 0.62 g, 42%, $R_{\rm f} = 0.6$ in CHCl₃-EtOH (1: 5) mixture containing 0.1% AcOH. 1H NMR (δ , ppm, relative to TMS): 9.59 (broad s), 8.75 (s, 0.3H), 8.42 (dd, 0.3H, 3J = 8.2 Hz, 4J =1.4 Hz), 8.37 (dd, 1H, 3J = 8.0 Hz, 4J = 1.2 Hz), 7.90 (s, 1H), 8.13 (dd, 1H, 3J = 8.0 Hz, 5J = 0.6 Hz), 7.3 (dd, 0.3H, 3J = 8.0 Hz, 5J = 0.8 Hz), 6.62 (s, 0.6H), 6.60 (s, 2H), 4.42 (d, 2H, 2J = 14.3 Hz), 4.44 (d, 0.6 H, 2J = 14.3), 4.28 (m, 2.6H), 4.20 and 4.21 (two q, 10.4H, 3J = 7.2 Hz), 3.60, 3.61 (two s, 10.4H), 2.11 (s, 3.9H), 1.27 (t, 15.6H, 3J = 7.2 Hz). HR-ESI-MS, positive mode: found 847.1862, calcd. for $C_{39}H_{41}Cl_2N_2O_{15}$ [M + H]⁺ 847.1883. CHN analysis: found C-52.03%, H-4.91%, N-3.36%, calcd. for $C_{41}H_{48}Cl_2N_2O_{19}$ ([**D**1·CH₃CO₂H·(H₂O)₂]): C—52.2%, H—5.1%, N-3.0%.

Selected reviews: (a) P. Jiang and Z. Guo, Coord. Chem. Rev., 2004, 248, 205; (b) F. Pina, M. A. Bernardo and E. Garcia-Espana, Eur. J. Inorg. Chem., 2000, 2143; (c) L. Fabbrizzi, M. Licchelli, P. Pallavicini, L. Parodi and A. Taglietti, Perspect. Supramol. Chem., 1999, 5, 93; (d) R. Krämer, Angew. Chem., 1998, 110, 804; (e) A. P. de Silva, H. Q. N. Gunaratne, T. Gunnlaugsson, A. J. M. Huxley, C. P. McCoy, J. T. Rademacher and T. E. Rice, Chem. Rev., 1997, 97, 1515; (f) L. Fabbrizzi, M. Licchelli, P. Pallavicini, A. Perotti, A. Taglietti and D. Sacchi, Chem. Eur. J., 1996, 2, 75.

- 2 Selected recent reports on fluorescent Zn²⁺ and Cu²⁺ sensors: (a) T. Gunnlaugsson, J. P. Leonard, K. Senechal and A. J. Harte, Chem. Commun., 2004, 782; (b) R. Meallet-Renault, R. Pansu, S. Amigoni-Gerbier and C. Larpent, Chem. Commun., 2004, 2344; (c) C. C. Woodroofe, R. Masalha, K. R. Barnes, C. J. Frederickson and S. J. Lippard, Chem. Biol., 2004, 11, 12, 1659; (d) C. J. Chang, J. Jaworski, E. M. Nolan, M. Sheng and S. J. Lippard, Proc. Natl. Acad. Sci. USA, 2004, 101, 5, 1129.
- 3 (a) P. Chavez-Crooker, N. Garrido and G. A. Ahearn, J. Exp. Biol., 2001, 204, 1433; (b) http://www.probes.com.
- 4 W. Breuer, S. Epsztejn, P. Millgram and I. Z. Cabantchik, Am. J. Physiol., 1995, 268, 6 Pt 1, C1354.
- 5 K. A. McCall and C. A. Fierke, Anal. Biochem., 2000, 284, 307.
- 6 **D1** is not a typical sensor since metal ion detection occurs due to an irreversible reaction. Moreover, it does not belong to chemodosimeters as well since metal ions are detected *via* determination of the kinetic parameter d*F*/d*t* rather than *via* measurement of fluorescence after reaction completion. Therefore, we call **D1** a hybrid sensor.
- 7 The synthesis of 2',7'-dichlorofluorescein-3'6'-diacetate-5(6)-carboxylate was as reported for the 2',7'-difluoro analog: W. C. Sun, K. R. Gee, D. H. Klaubert and R. P. Haugland, J. Org. Chem., 1997, 62, 19, 6469.
- 8 C. C. Woodroofe and S. J. Lippard, J. Am. Chem. Soc., 2003, 125, 38, 11458.
- 9 T. H. Fife and T. J. Przystas, J. Am. Chem. Soc., 1985, 107, 4, 1041.
- 10 V. Dujols, F. Ford and A. W. Czarnik, J. Am. Chem. Soc., 1997, 119, 31, 7386.
- 11 Q. Wu and E. V. Anslyn, J. Am. Chem. Soc., 2004, 126, 45, 14682.