

Elimination-Addition Mechanisms of Acyl Transfer Reactions

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I. Introduction

Acyl transfer reactions proceeding via addition-elimination mechanisms have been studied extensively, and, although this mechanism is not as thoroughly mapped as would be hoped, there exists a large body of work in the literature stemming in the main from the early '50's.^{1a} The elimination-addition mechanism for certain acyl-transfer reactions has been known since the early part of the century in contrast to the relatively late knowledge about the former mechanism, and it therefore seems somewhat paradoxical that until only recently have detailed mechanistic studies been reported for reactions in aqueous solution. This article reviews the recent and early work on the subject.

We shall be discussing compounds which can undergo a 1,2-elimination from the acyl function and the atom α to it (eq 1) and omit topics such as cyclizations (for example, the hy-



drolysis of 4-nitrophenyl hippurate)^{2a} and 1,4-eliminations. Reactions involving elimination of an electrophile other than the proton and compounds where the central acyl atom is other than carbon, phosphorus, or sulfur are not included. Metal ion complexes able to hydrolyze via an elimination-addition mechanism are reviewed elsewhere.^{2b}

A. Control of Mechanism

Mechanisms which are available for acyl transfer can be exemplified by transfer of a carbamoyl group from an acceptor to hydroxide and are illustrated in Figure 1. The mechanisms illustrated are limiting ones with respect to timing of bond making and bond breaking in a particular pathway, and the only concession to an intermediate mechanism is the E2 decomposition where leaving group and proton are abstracted simultaneously. In nonaqueous systems the process involving ionization (SN1) can prevail as, for example, in the Friedel-Crafts type reaction,^{3a,b} but we are concerned with aqueous media; the compounds involved do not necessitate possession of a proton on the α atom. Let us consider a free energy diagram (Figure 2) for A-E^{3c} attack of hydroxide ion on an ester and for the E1cB process for the same reaction; in the former an addition intermediate T⁻ is formed and in the latter, after ionization to yield conjugate base A⁻, elimination occurs to give the intermediate I which adds to give product. The E1cB profile is governed by the equilibrium constant between base and ester and the transition-state energy for intermediate formation which is presumably connected with the free energy of formation of I.⁴ The stability of the transition-state for intermediate formation is related among other things to steric constraints; for example, the formation of an endocyclic carbodiimide in a five-membered ring is very difficult

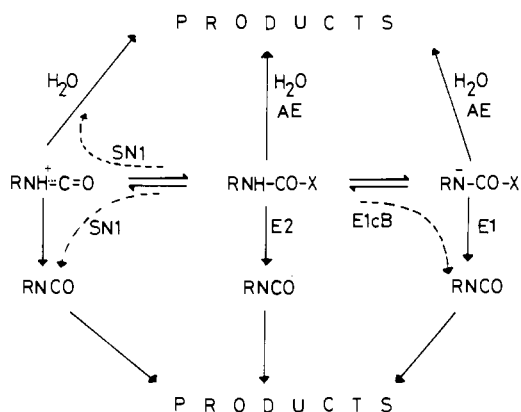


Figure 1. Mechanisms available for acyl transfer illustrated by the hydrolysis of a carbamate; the figure shows only limiting mechanisms with respect to the timing of bond formation and fission. The mechanistic symbolism used here and later is explained in ref 3c.

energetically and could force a reaction to take an A-E pathway.

Considerable discussion has centered on the question of concerted 1,2-eliminations as opposed to stepwise processes,⁵ and factors controlling the path taken, either E2 or E1cB, have been discussed theoretically.⁶ Recently a rule has been proposed which defines the pK_a limits of the base in which a concerted proton transfer is possible;^{7a} for example, the base pK_a must lie between that of the carbamate NH and the NH of RN^+HCO for an E2 mechanism to be possible in carbamoyl transfer.

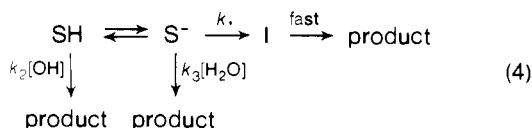
B. Effect of pH

Hydrolysis of an ester with a dissociable proton on the α atom possesses a sigmoid pH dependence for A-E, or E-A mechanisms (eq 2). The kinetic equation governing the hydrolysis is given by eq 3. The parameters are defined as: k_3 ,

$$k_{\text{obsd}} = k' / (1 + a_H / K_a) \quad (2)$$

$$= k_2 K_W / [K_a(1 + a_H / K_a)] + (k_1 + k_3) / (1 + a_H / K_a) \quad (3)$$

attack of water on conjugate base; k_1 , E1-elimination from conjugate base; k_2 , A-E attack of hydroxide on neutral ester; K_a , ionization constant of the " α proton"; k' , the observed plateau rate constant where k_{obsd} is independent of pH. Clearly the pH dependence is not an adequate tool for distinguishing between the various mechanisms (eq 4).



II. SN1 Mechanisms

All the evidence for SN1 mechanisms of aryl transfer is for acyl groups devoid of a labile " α proton". It is in any case unlikely that proton transfer from an electronegative α atom will be anything but diffusion controlled because the pK_a of the cationic intermediate should be less than that of the oxonium ion; for carbon α atoms this situation may not be true.

A. Carboxylic Acid Derivatives

Probably the first good evidence for an acylium ion was obtained by Treffers and Hammett in 1937^{7b} using cryoscopic measurements of mesitoic acid in concentrated sulfuric acid. Evidence for acylium ion formation in methyl mesitoate hy-

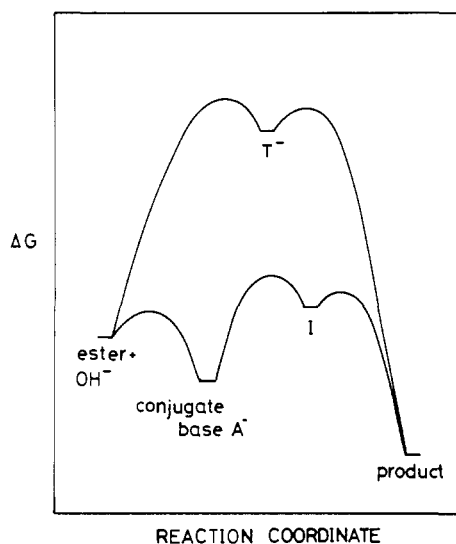
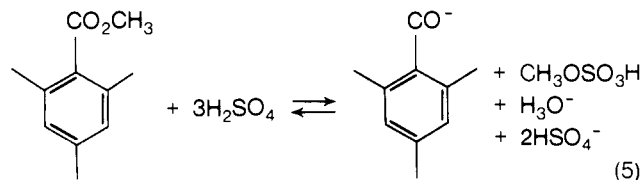
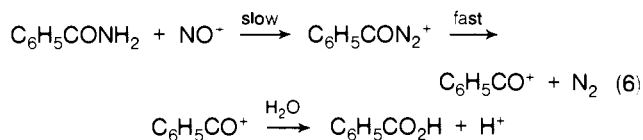


Figure 2. Free-energy diagram for hydrolysis of an ester proceeding via A-E attack of hydroxide on the acyl atom and via the E1cB pathway; symbols are described in the text.

drolyses comes from cryoscopic measurements using 100% sulfuric acid⁸ and dependence of the rate constant for hydrolysis on a Hammett acidity function in strong sulfuric acid solutions.^{9a} Positive entropies of activation close to zero and absence of exchange at carbonyl oxygen also indicate an SN1 mechanism.^{9b} The absence of exchange at carbonyl oxygen



in the diazotization of benzamide together with dependence of the rate constant on an H_R acidity¹⁰ was taken as consistent with acylium ion formation (eq 6). The arguments using



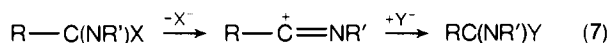
lack of exchange are strengthened by the observation that alkaline hydrolysis of methyl mesitoate has $k_{\text{hydrolysis}}/k_{\text{exchange}} = 6.8$.^{11a} Further evidence for an SN1 mechanism in acid-catalyzed hydrolysis is a Hammett ρ of -3.22 for hydrolysis of methyl 4-substituted 2,6-dimethylbenzoates.^{11b}

Benzoyl chlorides appear to hydrolyze via an A-E mechanism in solvents low in water content, but this shifts to SN1 as the water concentration increases.^{12-15a} Trapping experiments with weak nucleophiles^{15b} also suggest an acylium ion intermediate accounting for about 40% of the total reaction; this figure agrees with an estimate obtained by Hudson and Crunden^{15c} from solvolysis of benzoyl chloride in solvents of "equal ionizing power".^{15d} The powerful acceleration of the rate of hydrolysis of mesitoyl chloride by hydroxide and carbonyl oxygen exchange are not consistent with acylium ion intermediates under these conditions;^{16,17} the Hammett ρ values for neutral and acid- and base-catalyzed hydrolyses of 4-substituted 2,6-dimethylbenzoyl chlorides are, respectively -3.85 , -3.73 , and $+1.20$ in 99% acetonitrile-water. Absence of carbonyl oxygen exchange for acid and neutral hydrolysis is consistent with an SN1 mechanism for acid and neutral reactions.^{18a} Furthermore the neutral and acid Ham-

mett dependencies are on σ^+ . Ethyl chloroformate hydrolysis in water is 50-fold less reactive than benzoyl chloride,^{17b} if the acylium ion mechanism predominated, the reverse order would be expected.

Susz and his coworkers^{17c,d} found that in aprotic solvents hindered benzoyl chlorides (2,6-dimethyl- and mesityl) gave infrared spectra, when mixed with aluminum trichloride or titanium tetrachloride, indicative of a triple bond stretching frequency ($\sim 2200\text{ cm}^{-1}$). Nonhindered acid chlorides gave much lower frequencies. These results were considered consistent with a resonance hybrid possessing considerable acylium ion character. Recent work with antimony pentafluoride, fluorosulfonic acid, and sulfur dioxide media involved the demonstration of free oxocarbonium ion (CH_3CO^+)^{18b} via NMR techniques.

The transfer reactions of the imido acyl group have recently been shown to involve the E-A mechanism (eq 7).^{18b-i}



B. Carbamic Acid Derivatives

Hydrolysis of dimethylcarbamoyl chloride appears to be SN1 as judged by the small positive entropy of activation, lack of reactivity to hydroxide ion, and trapping by added nucleophiles which do not increase the rate yet yield trapped products. *N,N*-Dimethylcarbamoylpyridinium chloride, however, hydrolyzes probably via an A-E mechanism in water consistent with the absence of a mass law effect in pyridine buffers, large negative entropy of activation, a moderate solvent isotope effect, and a linear free-energy relationship with 4-nitrophenyl acetate reactivity with nucleophiles including water.^{19a} Similar evidence was marshalled^{19b} to show that diphenylcarbamoyl chloride and other carbamoyl chlorides^{19c} and the corresponding pyridinium derivative hydrolyzed via SN1 and A-E mechanisms, respectively.

The aminooxonium ion has been demonstrated as an entity in antimony pentafluoride-fluorosulfonic acid solutions.^{19d}

C. Phosphoramidates

N,N,N',N'-Tetramethyldiaminophosphorochloridate hydrolyzes in water faster than diisopropyl phosphorochloridate consistent with an SN1 mechanism. The rate of hydrolysis has a small negative entropy of activation and is unaffected by hydroxide ion, 3-cresolate ion, or pyrrolidine.^{20a} Probably the situation is better stated as an early cleavage of the P-Cl bond compared to bond formation between phosphorus and the nucleophile.^{20b}

D. Aminosulfonates

It was proposed that dimethylaminosulfonyl chloride hydrolyzed via an SN1 process because of the lack of effect of added nucleophiles.²¹ Mercuric chloride, a reagent known to promote SN1 reactions by electrophilic catalysis, increased the hydrolysis rates of dimethylcarbamoyl, dimethylaminosulfonyl, tetramethyldiaminophosphoryl, and benzoyl chlorides consistent with an acylium ion intermediate.²²

E. Phosphinates

Comparison of relative rates with the hydrolysis of *tert*-butyl chloride in different media indicates that diisopropylphosphinyl chloride probably hydrolyzes via an A-E mechanism, but di-*tert*-butylphosphinyl chloride, where an associative mechanism is suppressed by steric hindrance, goes via an SN1 pathway.²³ Cryoscopic studies with 100% sulfuric acid gave no indication of *stable* phosphinyl cation formation from phosphinic acids.

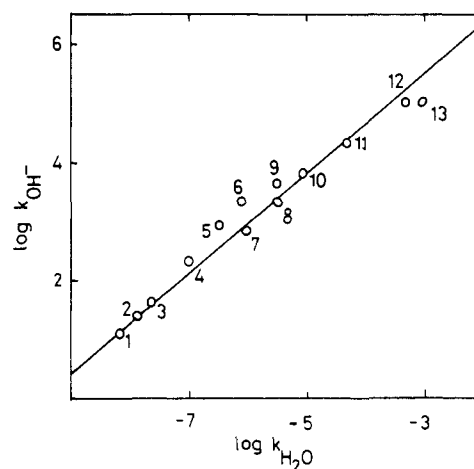


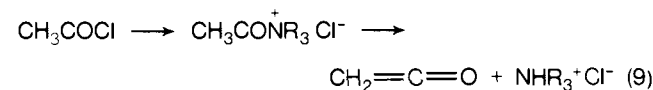
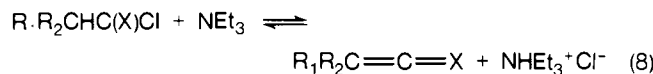
Figure 3. Plot of the logarithm of the second-order rate constants for reaction of hydroxide vs. water with esters: $\text{XCO}_2(2\text{-NO}_2\text{C}_6\text{H}_4)$, where X is (1) C_2H_5 , (2) CH_3 , (3) $\text{C}_6\text{H}_5\text{CH}_2$, (4) $\text{C}_2\text{H}_5\text{SCH}_2$, (5) $-\text{CO}_2^-$, (6) $(\text{CH}_3)_3\text{N}^+\text{CH}_2-$, (7) $\text{C}_6\text{H}_5\text{OCH}_2-$, (8) $\text{NCC}(\text{CH}_3)_2-$, (9) BrCH_2- , (10) ClCH_2- , (11) $\text{C}_5\text{H}_5\text{N}^+\text{CH}_2-$, (12) $\text{C}_2\text{H}_5\text{OCO}-$; (13) $\text{Cl}_2\text{CH}-$. The figure is redrawn from Figure 3 in B. Holmquist and T. C. Bruice, *J. Am. Chem. Soc.*, **91**, 2982 (1969).

III. E1cB Mechanisms

A. Carboxylic Acid Derivatives

1. Ketenes and Ketenimines

Ketene and ketenimine synthesis from acid chlorides²⁴⁻²⁷ and imine chlorides²⁸⁻³¹ with tertiary amines is a well-known preparative route (eq 8, $\text{X} = \text{O}, \text{NR}$) and presumably owes its success to the absence of protonic species in the solvent. Acylammonium ions could be involved (eq 9)²⁵ in the mecha-



nism, and the observation that ketene itself is not hydrolyzed exceptionally quickly in water^{31b} makes ketene intermediates in acyl transfers in water seem not unexpected (in retrospect).

2. Comparison with Addition-Elimination Reactions

The hydrolysis of 2-nitrophenyl esters of monosubstituted acetic acids in water was found to be related to the rate constant for hydroxide attack on these esters;³² hydroxide had 0.84 the selectivity to change in structure of the acid as the water rate (Figure 3) and 2-nitrophenyl acetates not falling on the regression line were judged to have the hydroxide reaction not proceeding via an A-E path because the line is defined for esters which must involve such mechanisms. Acetate itself fits the line and the hydroxide reaction is therefore A-E. The pK_a of the " α proton" of 2-nitrophenyl acetate is unknown but must lie between 14 and the pK_a of ethyl acetate (~ 25) so that preference for the A-E mechanism probably resides in the concentration of the anion being too low to support an E1 reaction competitive with the A-E path; the rate of proton removal is also most likely low for the acetate esters as well.

Enhancements in the hydroxide term occur with cyanoacetate (100-fold),³³ dimethylsulfonioacetate (10-fold),³³ ethyl malonate (8×10^3 -fold),^{34a} and ethyl α -methylmalonate (2×10^2 -fold)^{34a} where the mechanisms are proposed to be E-A, borderline, E-A, and E-A, respectively. The pK_a of the " α

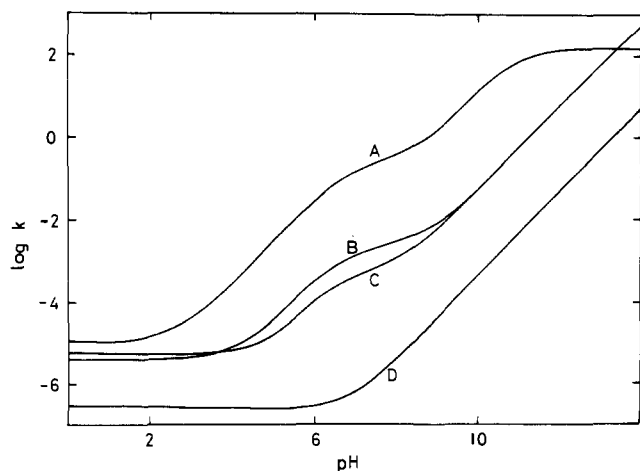


Figure 4. Plots of the pseudo-first-order rate constant for hydrolysis at zero buffer concentration of (A) 2-nitrophenyl ethyl malonate, (B) 2-nitrophenyl ethyl α -methylmalonate, (C) 4-nitrophenyl ethyl α -methylmalonate, (D) 2-nitrophenyl ethyl α,α -dimethylmalonate. The figure is redrawn from Figure 1 in B. Holmquist and T. C. Bruice, *J. Am. Chem. Soc.*, **91**, 2993 (1969).

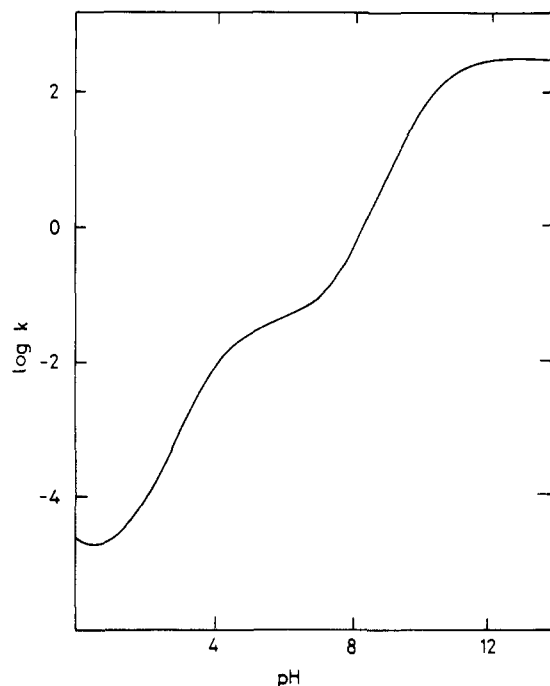
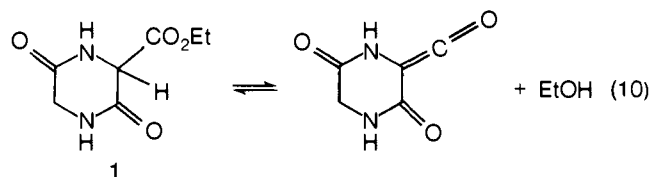


Figure 5. Plot of the pseudo-first-order rate constant for hydrolysis of 4-nitrophenyl acetoacetate at zero buffer concentration. The figure is redrawn from Figure 1 of R. F. Pratt and T. C. Bruice, *J. Am. Chem. Soc.*, **92**, 5956 (1970).

protons" in these esters is low; thus there is sufficient carbanion to sustain an E-A mechanism where this occurs. We shall see later, however, that an E1cB mechanism is not necessarily attendant on a low pK_a for the α proton. An ester with a labile " α proton" which undergoes rapid exchange is ethyl 2,5-piperazinedione-3-carboxylate,^{34b} and it is possible that this involves an E1cB process (eq 10).



The use of models which cannot react via an E-A mechanism is also exemplified by the arguments^{34c} against the E-A

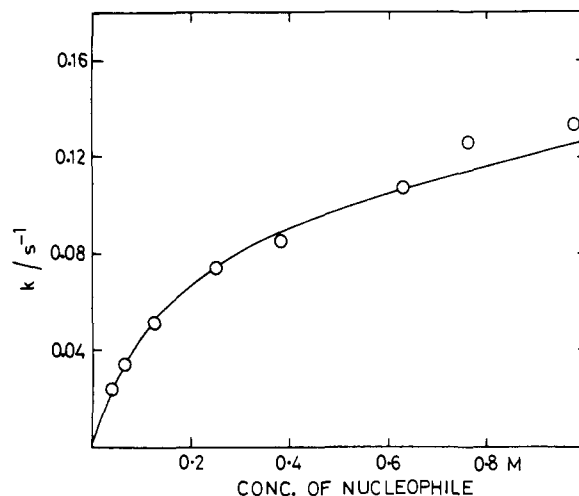
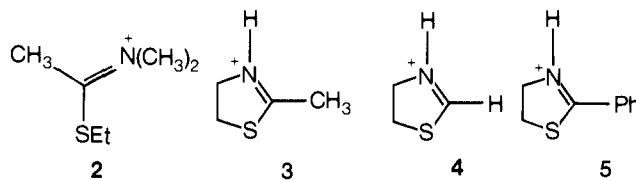


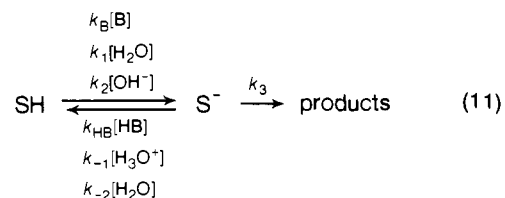
Figure 6. Effect of changing *N*-ethylmorpholine buffer concentration at pH 8.43 on the rate of release of 2-nitrophenol from 2-nitrophenyl ethyl α -methylmalonate; rates are corrected for background hydrolysis. An A-E mechanism involving attack of the amine on the acyl carbon contributes to the reaction. The figure is redrawn from Figure 3 in B. Holmquist and T. C. Bruice, *J. Am. Chem. Soc.*, **91**, 2993 (1969).

hydrolysis of *S*-ethyl *N,N*-dimethylthioacetimidate (2). The thiazolines (3, 4, and 5) hydrolyze in alkali with approximately similar rate constants^{34d} as expected for an A-E mechanism, and it is therefore unlikely that the acetimidate hydrolysis is E-A.



3. Proton Transfer

The pH dependencies for the pseudo-first-order rate constant for hydrolysis of a number of substituted phenyl esters possess inflections (Figures 4 and 5) which do *not* correspond to the pK_a 's of any carbon acid grouping in the substrate (2-nitrophenyl^{34a} and 4-nitrophenyl³⁵ ethylmalonate, 2- and 4-nitrophenyl ethyl α methylmalonate,^{34a} 4-nitrophenyl acetoacetate,³⁵ and 4-nitrophenyl *S*-ethyl thiomalonate³⁵). The hydrolyses are catalyzed by base,³⁴⁻³⁶ and the catalytic effect levels off as the buffer concentration increases (Figure 6). The pH dependencies and curved buffer catalyses can be accommodated by eq 11;³⁴ in the absence of buffer at low pH's



$k_{-1}[\text{H}_3\text{O}^+] > k_3$; thus $k_{\text{obsd}} = (k_1 k_3 / k_{-1}) [\text{H}_2\text{O}] / [\text{H}_3\text{O}^+]$, and the pH dependence has positive slope equal to unity. The k_1 term becomes rate limiting as the pH rises ($k_{\text{obsd}} = k_1 [\text{H}_2\text{O}]$) because $k_{-1}[\text{H}_3\text{O}^+]$ falls below k_3 ; the $k_2[\text{OH}^-]$ term becomes dominant as pH increases further, but when all substrate is converted to conjugate base, k_3 is rate limiting. "Curved" buffer catalysis can be explained by a change in rate-limiting step caused by $k_{\text{HB}}[\text{HB}]$ becoming greater than k_3 ³⁴ as seen in the *partial* kinetic equation (eq 12) omitting

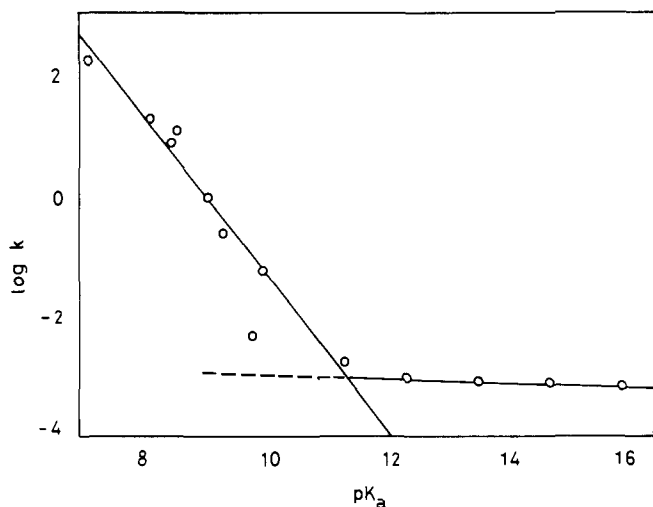
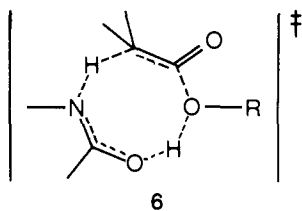


Figure 7. Effect of changing the leaving group on the hydrolysis of acetoacetate esters. Points above $pK_a = 10$ represent alkyl leaving groups; those below, phenol and oxime leaving groups. The parameter plotted ($\log k'$) is that defined in eq 3; the concave relationship indicates a change in mechanism. The figure is redrawn from Figure 3 in R. F. Pratt and T. C. Bruice, *J. Am. Chem. Soc.*, **92**, 5956 (1970).

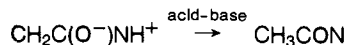
$$k_{\text{obsd}} = k_B[B]/(k_{\text{HB}}[HB]/k_3 + 1) \quad (12)$$

water and hydroxide as effective species. Linear catalysis at high concentration of buffer is consistent with a pathway involving attack at the ester carbonyl.³⁴

Concerted acid-base catalysis of acylation by ketene^{37a,b} has been suggested for the catalytic action of amides, and presumably a pathway exists for concerted acid-base catalysis via a ketene for ester exchange; the only real evidence which exists for this suggestion is the absence of catalysis by *N,N*-disubstituted amides. Simple acid-base catalysis of acyl-



ation by ketene could involve either formation of an intermediate reactive acyl compound^{37b} (CH_3COB^+) or facilitation of proton transfer in an addition-elimination type mechanism



4. Deuterium Oxide Solvent Isotope Effects

This effect has been utilized to attempt to distinguish between E-A and A-E mechanisms in hydroxide catalyzed reactions.³⁸ For the plateau region the E-A mechanism gives $k_{\text{obsd}} = k_1$ and the A-E process gives $k_{\text{obsd}} = k_2 K_w / K_a$ (see eq 3), and it has been assumed^{38b} that k_1 has little or no isotope effect. The method was tested for 5-nitrocoumarone where $k'(D/H) = 0.58$. The effect on k_2 , K_w , and K_a (D/H, 1.22, 0.15, H/D, 3.5, respectively) is available in the literature^{38a} or can be estimated,^{38b} thus $k'(D/H)$ is calculated to be 0.65 for the A-E mechanism in agreement with the observed value. Unfortunately, this criterion is not now as clear-cut as it was hoped because other bona fide E1 reactions have been studied and have $k_1(D/H)$ varying from unity to 0.69.³⁵ This result is not surprising since dispersal of charge from ground to transition state in E1 reactions would be expected to depend heavily on solvation.

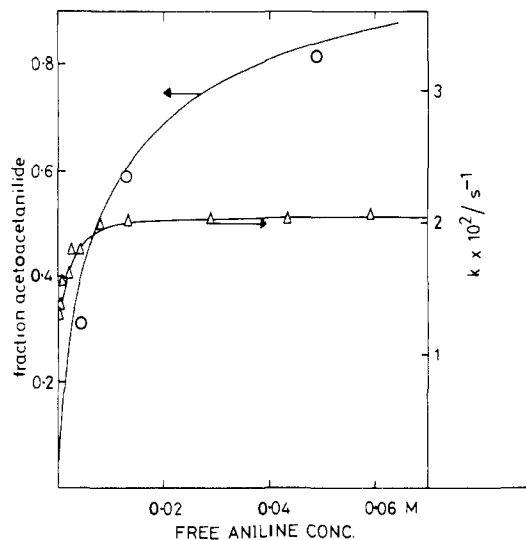
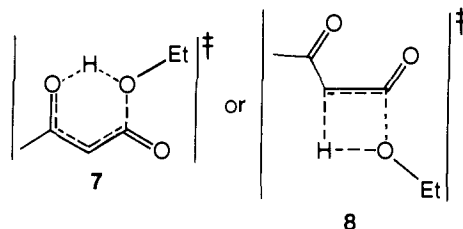


Figure 8. Trapping an intermediate in the reaction of 4-nitrophenyl acetoacetate with aniline and water at pH 4.6. Above 0.01 M aniline concentration the rate of 4-nitrophenol release is constant, but the fraction of acetoacetanilide in the product still increases considerably. The figure is redrawn from Figures 4 and 5 of R. F. Pratt and T. C. Bruice, *J. Am. Chem. Soc.*, **92**, 5956 (1970).

5. Leaving Group

The alkaline plateau rate constant for hydrolysis of substituted phenyl acetoacetates has a Hammett ρ value of +2.8, and the dependence is on σ^- , consistent with considerable C-O cleavage in the transition state of the rate-limiting step. A wider range of leaving groups can be included if the correlation is with the pK_a of the leaving hydroxyl function. Such a Bronsted-type plot (Figure 7) is meaningful provided the leaving atoms and their charge type are identical. The results were interpreted as a change in mechanism³⁵ (strongly indicated by the sharp break in the Bronsted plot from low to high slope as the pK_a is decreased) from E-A type for the phenyl esters to A-E type for the alkyl esters. The cause of this change probably resides in the greater sensitivity of E1 reactions to the leaving group than A-E reactions. Although alkyl acetoacetates undergo hydrolysis in alkali via the A-E mechanism, catalysis could still promote transfer via an E-A mechanism: cyclic transition states have been proposed for alcoholysis of alkyl acetoacetates leading to a ketene intermediate.³⁹ Evidence for this involves a rate constant independent of alcohol concentration and a small negative entropy of activation independent of solvent change. The formation of ket-



enes has been postulated in the use of malonates in providing thermally labile cross links for cotton modification.⁴⁰

6. Trapping

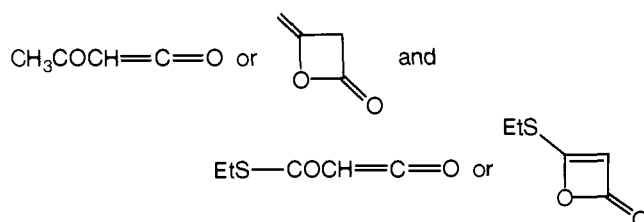
Formation of acetoacetanilide from 4-nitrophenyl acetoacetate in aniline buffers was found to increase significantly over a range of aniline concentration where the rate constant for 4-nitrophenol release was invariant (Figure 8). This result is consistent with a mechanism involving release of 4-nitrophenol prior to aniline attack. The absence of nucleophilic at-

TABLE I. Reactions where Elimination Intermediates Have Been Demonstrated in Basic Solution

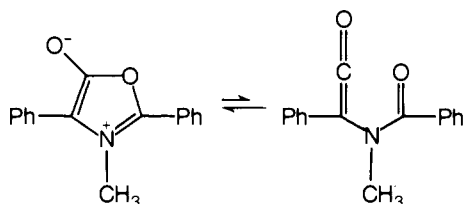
Reactants	Intermediates	Ref
NH ₂ C(NH)NH ₂	NH ₂ CN	46, 47
R'NHCON(NO)R	RCHN ₂ , R'NCO	<i>h</i> , 48, 49
NH(NO ₂)C(NH)N(NO)CH ₃	CH ₂ N ₂ , N(NO ₂)C(NH)	<i>e</i>
NH ₂ CSNHR	NCS ⁻ , RNCS	<i>c</i> , 50
RNHCON ₃	N ₃ ⁻ , RNCO	<i>g</i>
RNHC(NH)SR'	R'SH, RNHCN	51, 52
RNHCO ₂ R'	NCO ⁻ , R'SH	<i>d</i> , 53
RNHCS(S ⁻)	RNCS	<i>a</i> , 54
RNHCS(OR)	RNCS	<i>a</i> , 54
NH ₂ CS(OR)	NCS ⁻	55
RNHC(NH)NH ₂	NH ₂ CN ⁺	56
NH ₂ COOR	NCO ⁻	57
NH ₂ CONHR	NCO ⁻	<i>b</i> , <i>e</i> , <i>f</i> , 58, 59
NH ₂ CONHNO ₂	NCO ⁻ , NH ₂ NO ₂	<i>i</i>
PhCONHC(NH)(C ₂ H ₅ N ₃)	PhCONHCN	52

^a J. E. Hodgkins and M. G. Ettliger, *J. Org. Chem.*, **21**, 204 (1956); M. G. Ettliger and J. E. Hodgkins, *J. Am. Chem. Soc.*, **77**, 1833 (1955); **78**, 1952 (1956); A. Kjaer and R. Gmelin, *Acta Chem. Scand.*, **10**, 1193 (1956); **11**, 906 (1957); A. Kjaer, R. Gmelin, and R. B. Jensen, *ibid.*, **10**, 26,432 (1956); A. Kjaer and R. B. Jensen, *ibid.*, **10**, 141 (1956); D. L. Garmaise, R. Schwartz, and A. F. McKay, *J. Am. Chem. Soc.*, **80**, 3332 (1958); L. M. Dowling, and G. R. Stark, *Biochemistry*, **8**, 4728 (1969); J. von Braun, *Chem. Ber.*, **42**, 4568 (1909); **45**, 1563 (1912); J. von Braun and G. Manz, *Justus Liebigs Ann. Chem.*, **468**, 258 (1929); J. von Braun, W. May, and R. Michaelis, *ibid.*, **490**, 189 (1931); ^b W. H. R. Shaw and B. Grushkin, *J. Am. Chem. Soc.*, **82**, 1022 (1960); ^c W. H. R. Shaw and D. G. Walker, *ibid.*, **78**, 5769 (1956); **79**, 2681, 3683, 4329 (1957); **80**, 5337 (1958). ^d G. R. Stark, W. H. Stein, and S. Moore, *J. Biol. Chem.*, **235**, 3177 (1960); U. Weiss, *J. Am. Chem. Soc.*, **69**, 2684 (1947); G. Frerichs and P. Foerster, *Justus Liebigs Ann. Chem.*, **371**, 227 (1909); H. Bechurts and G. Frerichs, *J. Prakt. Chem.*, **66**, 172 (1902); J. D. Ravel, J. J. McCord, C. G. S. Skinner, and W. Shire, *J. Biol. Chem.*, **232**, 159 (1958). ^e G. D. Vogels, F. E. De Windt, and W. Bassie, *Recl. Trav. Chim. Pays-Bas*, **88**, 940 (1969); G. D. Vogels and C. Van Der Drift, *ibid.*, **88**, 951 (1969). ^f A. A. R. Sayigh, J. N. Tilley, and H. Ulrich, *J. Org. Chem.*, **29**, 3344 (1964). ^g T. Curtius and A. Burkhardt, *J. Prakt. Chem.*, **58**, 205 (1898); E. Oliveri-Mandala and F. Noto, *Gazz. Chim. Ital.*, **43**, 514 (1913); E. Oliveri-Mandala, *ibid.*, **44**, 662 (1914). ^h E. R. Garrett, S. Goto, and J. F. Stubbin, *J. Pharm. Sci.*, **54**, 119 (1965). ⁱ G. L. Boivin and P. A. Boivin, *Can. J. Chem.*, **29**, 478 (1951); T. L. Davis and K. C. Blanchard, *J. Am. Chem. Soc.*, **51**, 1797 (1929); B. Boopsingh and J. M. Briody, *J. Chem. Soc., Perkin Trans. 2*, 1487 (1972).

tack by aniline at the ester carbonyl is confirmed by the absence of an effect, on the rate constant for 4-nitrophenol release, of increasing the aniline concentration (see Figure 8). An intermediate was observed using ultraviolet spectroscopy (at 325 nm) and fast reaction equipment in the hydrolysis of 4-nitrophenyl *S*-ethyl malonate, but it is likely that this intermediate is the carbanion rather than the ketene. Although there seems to be little doubt about the existence of an intermediate after the carbanion in the transfer of some of these reactive esters, there is a shadow of doubt about the precise nature of these intermediates.^{41a}



An interesting example of intramolecular trapping was described recently^{41b} where the trapped ketene cannot be stabilized further by proton transfer:



7. Biological Aspects

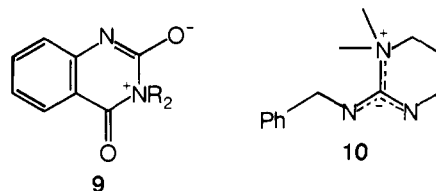
So far as we are aware there is no evidence for a ketene as an intermediate in biochemical pathways although there are many instances such as in the biosynthesis of lipids which could involve such species. Recently D amino acid oxidase has been found to be inhibited by 4-nitrophenyl glycinate but

not by ethyl glycinate.⁴² Although the free glycinate certainly ought to transfer via an A-E mechanism, the formation of a protonated imine could lower the pK_a of the "α proton" sufficiently to support an E-A mechanism for the 4-nitrophenyl ester. The poor leaving ability of the ethyl ester presumably ensures its unreactivity as an inhibitor.

B. Carbamic Acid Derivatives

1. Trapping

The intermediate in transfer reactions of the parent carbamate, isocyanic acid, is stabilized by proton transfer as the conjugate base and is therefore effectively trapped. Table I summarizes reactions where an intermediate has been demonstrated; carbodiimides and cyanamide from iminocarbamates are also usually sufficiently stable in water to have at least a transient existence. Intramolecular trapping of isocyanate (**9**)^{43a} and carbodiimide (**10**)^{43b} functions has been re-



ported; the absence of labile protons prevents stabilization via proton transfer.

2. Heavy Atom Isotope Effects

The deuterium oxide solvent isotope effect has been used to diagnose an E1cB mechanism for the alkaline hydrolysis of 4-nitrophenyl *N*-methylcarbamate.^{38c} The value for $k_{OH^-}(D/H) = 1.8$ is larger than expected for nucleophilic attack at the carbonyl carbon (1.22)^{44a} but is compatible with preequilibrium formation of an anion.^{44b} Primary heavy atom isotope effects are more definitive, but only a few relevant to this discussion have been reported. Hydrolysis of urea in acidic solution gave carbon primary isotope effects (¹²C/¹⁴C) = 1.101

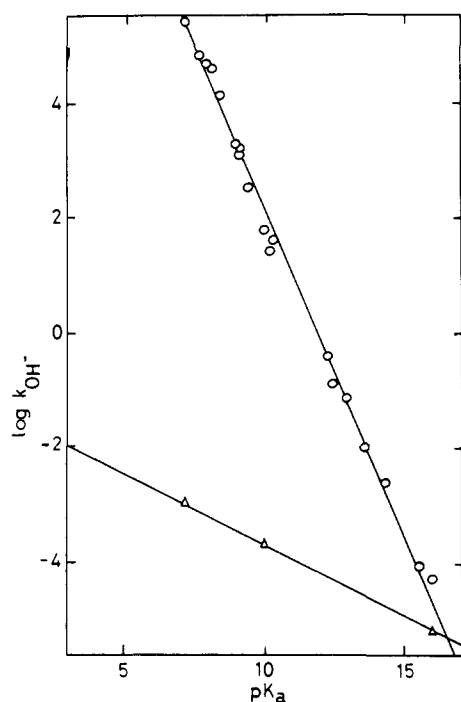


Figure 9. Dependence of k_{OH^-} on the pK_a of the leaving hydroxyl group for alkyl and phenyl esters of (Δ) *N*-methyl-*N*-phenylcarbamic acid and (O) *N*-phenylcarbamic acid. Alkyl esters are those with $pK_a > 10$; the figure is redrawn from Figure 1 of A. Williams, *J. Chem. Soc., Perkin Trans. 2*, 1244 (1973).

± 0.005 and $(^{12}C/^{13}C) = 1.055 \pm 0.003$.⁴⁵ These values indicate considerable N-C bond cleavage in the transition state of the rate-limiting step consistent with an E-A mechanism. Change in bond order from trigonal planar to tetrahedral in the N-C bond in an A-E mechanism could yield an isotope effect,^{45c} but this is likely in any case to be small. The bond order change effect has been considered by other workers for other systems to be relatively small.^{45d-g}

3. Leaving Group

Intermediate isolation is not useful for *N*-substituted carbamates, and linear free-energy relationships involving leaving group variation have been successfully employed as diagnostic tools.^{41a,60-63} The reaction of hydroxide with substituted phenyl *N*-phenylcarbamates possesses a high Hammett ρ (~ 3)^{41a,61} with σ^- dependence consistent with considerable C-O cleavage in the transition state of the rate-determining step. An A-E mechanism is characterized by low ρ and a σ dependence.⁶⁴ A Bronsted type relationship (Figure 9) holds for alkyl and phenyl esters of *N*-phenylcarbamic acid⁶⁰ suggesting an E-A mechanism over the whole range of pK_a up to the pK_a of ethanol. Probably a change in mechanism to A-E occurs for leaving groups slightly less acidic than ethanol. *N*-4-Nitrophenylcarbamates change mechanism when the pK_a of the leaving group is about 13.⁶¹ This result could be due to the lower thermodynamic stability of the intermediate 4-nitrophenyl isocyanate compared with the unsubstituted compound.

Small entropies of activation for the alkaline hydrolysis of phenyl *N*-phenylcarbamates argue for an E1cB mechanism,⁶⁵ but the ethyl esters have a relatively large negative value (-25.5 eu/mol) although the Bronsted-type plot suggests an E-A mechanism. The discrepancy is in part due to the fact that k_{OH^-} for a hydrolysis passing through an E1cB mechanism is composite and represents k_1K_2/K_w (see eq 2 and 3). The "Bronsted" data suggest that in any case a considerable proportion of the hydrolysis of the ethyl ester proceeds via the A-E pathway.

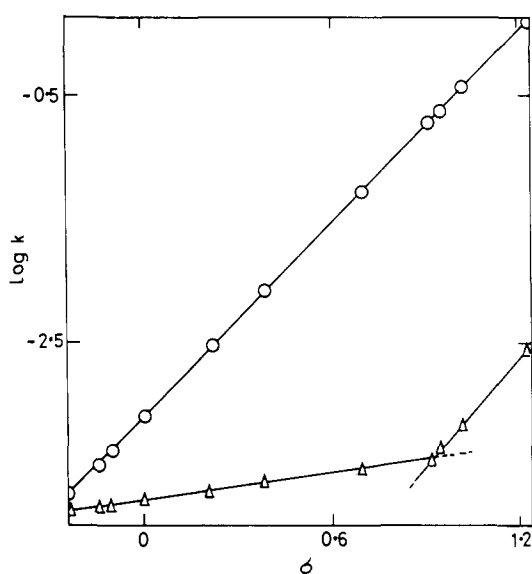
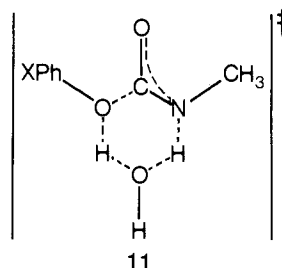


Figure 10. Effect of leaving group on the pseudo-first-order rate constants at pH 4 (Δ) and 8 (O) for the hydrolysis of aryl *N*-methylcarbamates. The circles represent the E1cB mechanism for alkaline hydrolysis; the triangles represent the neutral hydrolysis and a change in mechanism occurs as the electron withdrawing power of the substituent increases. The figure is redrawn from Figure 2 of T. Vontor and M. Večera, *Collect. Czech. Chem. Commun.*, **38**, 516 (1973).

The hydrolysis of substituted phenyl *N*-methylcarbamates involves a water term (plateau region of the pH profile) which has a Hammett dependence with a "break" diagnostic of a change in mechanism (Figure 10).^{66,67} Substituents with low σ values possess a low ρ value (0.36), whereas high σ substituents involve a high ρ value (3.04), and these results are indicative of a change in mechanism with increasing electron-withdrawing power of the aryl group. However, the authors^{66,67} equate a solvent deuterium oxide isotope effect (k_D/k_H) of 1.4, a high Hammett slope, and a large negative entropy of activation for the most reactive species with a cyclic transition state (**11**). The low slope, solvent deuterium oxide isotope effect of (D/H) 0.29, and lower negative entropy are associated with a simple A-E mechanism involving attack of water at the carbonyl in the least reactive series. These interpretations are clearly open to criticism, especially the allocation of Hammett slopes; before one can be satisfied as to the cause of the nonlinear Hammett plot, further work will be necessary.



4. Water Attack on Conjugate Base

The term k_1 of eq 4 for the hydrolysis of an acidic substrate refers to the decomposition of the conjugate base *prior* to water attack. The k_3 term refers to water attack on the conjugate base prior to release of the leaving group. The rate constants clearly are for limiting cases of a general mechanism: k_1 where C-O bond cleavage is complete; k_3 where C-O bond fission is incomplete when bond formation is advanced between water and carbon. Bearing these definitions

TABLE II. Rate Enhancements for Substrates with Blocked and Unblocked α Atoms^a

Ester ^b	Ratio $k_{\text{OH}}^{\text{H}}/k_{\text{OH}}^{\text{CH}_3}$	Ref
$\text{C}_6\text{H}_5\text{N}(\text{H})\text{COOpNp}$	7.3×10^4	41a, 61
$\text{CH}_3\text{N}(\text{H})\text{COOpNp}$	6.9×10^6 , 1.46×10^6	63, 68
$\text{N}(\text{H}_2)\text{COOpNp}$	6.25×10^8	63
$\text{C}_6\text{H}_5\text{N}(\text{H})\text{COOC}_6\text{H}_5$	1.05×10^6	61
$4\text{-NO}_2\text{C}_6\text{H}_4\text{N}(\text{H})\text{COOC}_6\text{H}_5$	7.99×10^4	61
$\text{C}_2\text{H}_5\text{OCOC}(\text{CH}_3)(\text{H})\text{COOpNp}$	6.77×10^3	34a

^a Other examples of this approach come in the appropriate sections for other acyl transfers. ^b Proton in parentheses is to be blocked by CH_3 ; pNp = 4-nitrophenyl; oNp = 2-nitrophenyl.

TABLE III. Steric Effect of " α Substituent" on Rate^a

Substrate ^b	k_{OH} ($1/M^{-1} \text{min}^{-1}$)	Substrate	k_{OH} ($1/M^{-1} \text{min}^{-1}$)
RNHCOOpNp		$\text{R}_2\text{NCOOpNp}$	
CH_3	1.3×10^5	CH_3	2×10^{-2}
C_2H_5	1.6×10^5	C_2H_5	3.6×10^{-4}
<i>n</i> - C_3H_7	1.6×10^5	<i>n</i> - C_3H_7	1.7×10^{-4}
<i>i</i> - C_3H_7	1.9×10^5	<i>i</i> - C_3H_7	5.1×10^{-6}

^a Values are taken from ref 68; temperature 22°; water solvent. ^b pNp = 4-nitrophenyl.

in mind we can distinguish experimentally between the extremes for the hydrolysis of phenyl *N*-phenylcarbamate.^{41a} If all the reaction went via the k_3 pathway (see eq 4), the observed first-order rate constant $k_{\text{obsd}} = k_3 K_a / (K_a + a_{\text{H}})$; the phenyl ester has a $\text{p}K_a > 12$ and, since $a_{\text{H}} > K_a$ for the pH range studied ($\text{pH} < 12$), then $k_{\text{obsd}} = k_3 K_a [\text{OH}^-] / K_w$. Thus a hydroxide term k_{OH} can be experimentally determined by plotting k_{obsd} vs. $[\text{OH}^-]$ and $k_3 = k_{\text{OH}} K_w / K_a$. Since $k_{\text{OH}} = 54.2 M^{-1} \text{sec}^{-1}$, the term k_3 has a value in excess of 0.542sec^{-1} ($54.2 \times 10^{-14} / 10^{-12}$). If this is so, then a rate constant less than 0.542sec^{-1} should never be observed assuming that the neutral species (unionized carbamate) will react faster with water in an A-E mechanism than does the anion; at pH 9 a rate constant of 0.000548sec^{-1} was measured. Thus a mechanism involving considerable bond formation between water and the anion in the transition state (k_3) is not involved.

5. Comparison with Addition-Elimination Reactions

Blocking the labile " α proton" by methyl substitution should provide a rate constant for hydroxide attack approximating to that for the A-E mechanism in the unblocked substrate. For definitive conclusions any rate enhancement of the unblocked over blocked substrate should be large, but it is difficult and probably unwise to put an exact lower limit to this figure although the values given in Table II are probably definitive. Table II illustrates enhancements for some reported systems.

6. Substituents on the α Atom

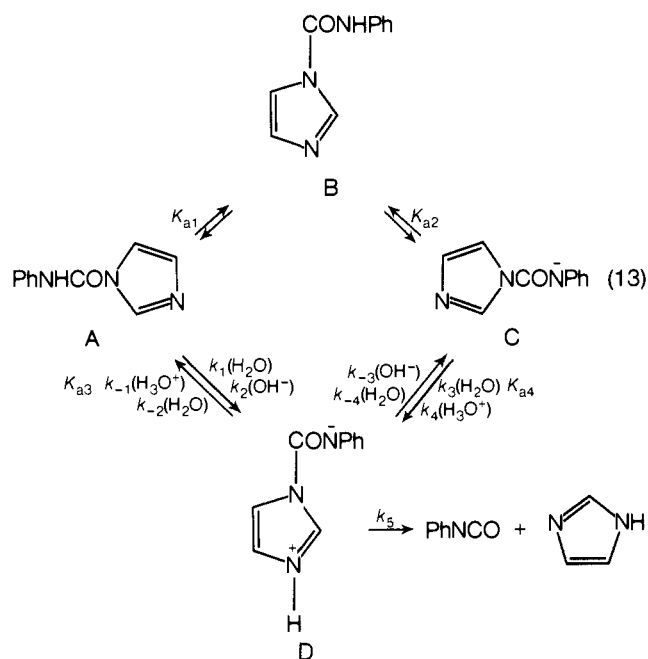
In principle the effect of substituents on the α atom on the rate constant can diagnose the mechanistic type in a hydrolytic process apart from the complete blocking already described. Table III shows that dependence of k_{OH} on the substituent does not vary substantially with increasing steric effect for a known E1cB reaction. A dramatic variation is observed with a bona fide A-E process, the alkaline hydrolysis of the *N,N*-disubstituted esters of carbamic acid. The E-A mechanism has less steric requirements for its rate-limiting steps (the ionization and the E1 processes) than the A-E mechanism involving the approach of a relatively bulky group to the acyl carbon. A better model would probably be the alkaline hydrolysis of substituted acetate esters because the di-

substitution in the present case over-accentuates steric effects.

Electronic effects of the *N*-substituent provide evidence for an E-A mechanism.⁶¹ The Hammett ρ for alkaline hydrolysis of phenyl (*N*-substituted)phenylcarbamates is +0.64; this value may be compared with the sensitivity of phenyl (*N,N*-methyl-substituted)phenylcarbamates to attack of the hydroxide ion (calculated from ref 61 to be +2.1).

7. Proton Transfer

The need for proton transfer in an E-A reaction with carbamates only gives rise to general acid-base catalysis under certain circumstances because proton transfer to or from electronegative atoms is usually diffusion controlled. Catalysis will only occur when proton transfer is necessary to or from an intermediate which has such a fast formation or decomposition step that the diffusion (proton transfer) step is rate limiting and the general base or acid is more efficient than bulk water. The hydrolysis of *N*-(*N'*-phenylcarbamoyl)imidazole has been shown to involve general acid-base catalysis⁶⁹ consistent with an E1cB mechanism (eq 13). The buffer catal-



ysis approaches zero as buffer concentration increases, consistent with a change in the rate-determining step. When the proton transfer steps are not rate limiting (at high buffer concentration) and the various species of eq 13 are in equilibrium with one another, a bell-shaped pH profile would be expected for the hydrolysis (dotted line in Figure 11); however, at zero buffer concentration two maxima are obtained. Assuming a steady-state concentration of the zwitterion (D), an equation relating k_{obsd} with a_{H} may be obtained (eq 14); values for

$$k_{\text{obsd}} = \frac{k_5(k_1 + k_2[\text{OH}^-])a_{\text{H}}/K_{a1} + k_5(k_3 + k_4 a_{\text{H}})K_{a2}/a_{\text{H}}}{k_{-1}a_{\text{H}} + k_{-2} + k_{-3}[\text{OH}^-] + k_{-4} + k_5} \times \frac{a_{\text{H}}K_{a1}}{a_{\text{H}} + a_{\text{H}}K_{a1} + K_{a1}/K_{a2}} \quad (14)$$

$\text{p}K_{a3}$ and $\text{p}K_{a4}$ may be calculated from the experimental data and come close to the expected values when k_{-1} and k_{-3} are taken as the diffusion rate constant ($10^{10} M^{-1} \text{sec}^{-1}$). As yet there are not enough data to construct an Eigen type plot^{70a} for the system.

The reverse of the hydrolysis of urea, namely synthesis via

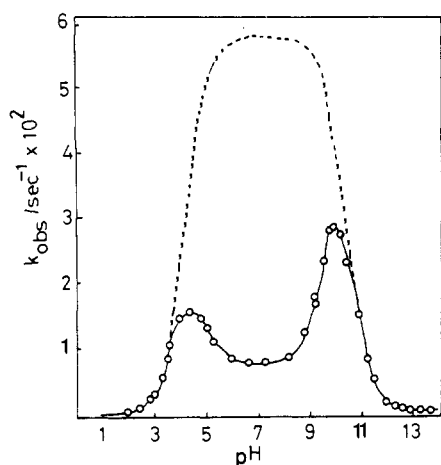
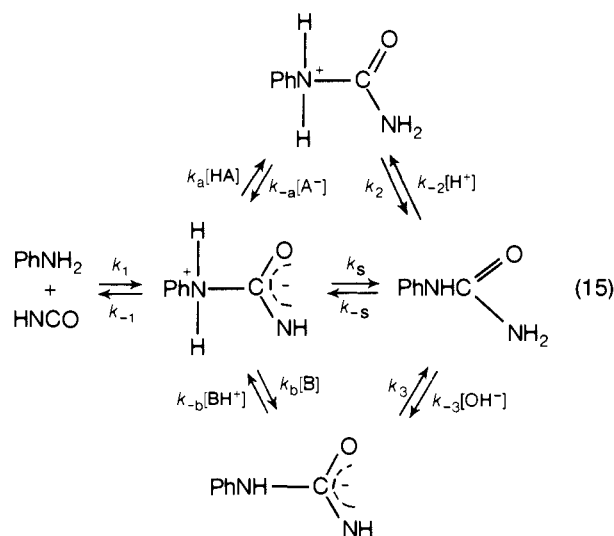


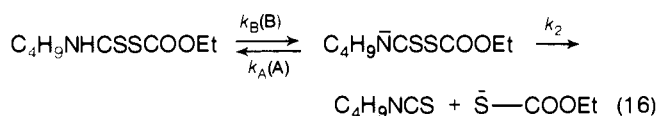
Figure 11. Effect of pH on the hydrolysis of *N*-(*N*-phenylcarbamoyl)imidazole (30°, 1 M ionic strength). The solid line represents the hydrolysis at zero buffer concentration; the dashed line is for higher buffer concentration. The figure is redrawn from Figure 1 of A. F. Hegarty, C. N. Hegarty, and F. L. Scott, *J. Chem. Soc., Perkin Trans. 2*, 1258 (1974).

isocyanic acid and amine, has been shown to involve^{71a,b} diffusion limiting proton transfer in the decomposition of the zwitterion intermediate (eq 15). Evidence for a change in rate-



limiting step from decomposition of the zwitterion to its formation is a nonlinear Bronsted type plot of amine reactivity vs. ammonium ion pK_a and the existence of a saturation effect in the buffer catalysis of the reaction of weakly basic amines with isocyanic acid. Strongly basic amines with k_1 rate limiting do not exhibit buffer catalysis. The Bronsted plots for acid and base catalysis are nonlinear as in the (now classical) Eigen plot^{70a} (e.g., Figure 12).

Another example of a change in rate-limiting step caused by increasing buffer concentration for reaction of a carbamate type compound has been reported by Hodgkins.^{54a} The catalysis of the formation of isothiocyanate from dithiocarbamates in pyridine buffers exhibits a saturation effect consistent with the mechanistic scheme of eq 16.



Catalysis by products in the reaction of isocyanates with amines and catalysis of this reaction by amides and acids has led to the suggestion that in nonaqueous solvents a concerted

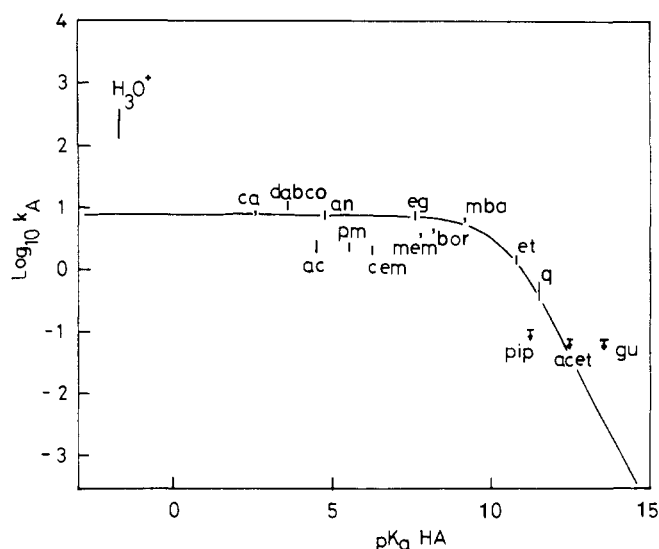
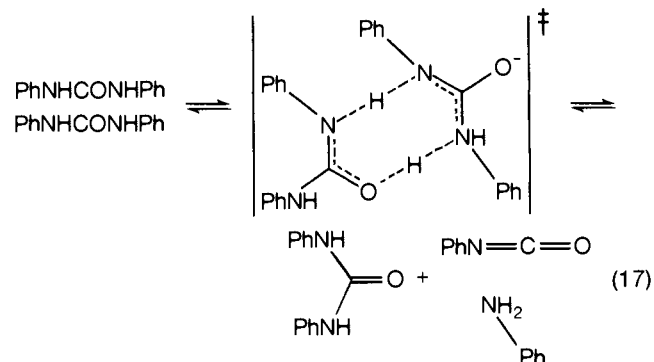
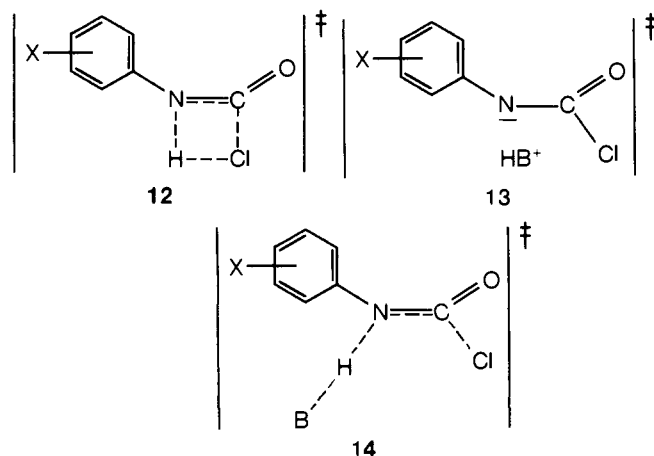


Figure 12. The Bronsted plot of k_A of the acid for the acid-catalyzed reaction of aniline with cyanic acid (25°, 1 M ionic strength). Legend: ca, chloroacetic acid; dabco, triethylenediamine dication; an, anilinium ion; pm, *N*-propargylmorpholinium ion; cem, *N*-2-chloroethylmorpholinium ion; mem, *N*-methylmorpholinium ion; eg, ethylglycine (protonated); bor, boric acid; mba, methyl β -alaninate (protonated); et, ethylammonium ion; q, quinuclidinium; pip, piperidinium; acet, acetamidinium; gu, guanidinium. The figure is redrawn from ref 71b.

catalytic path is involved (eq 17).^{71b-d} Consistent with this suggestion is the absence of catalysis by such amides as *N,N*-dimethylacetamide. In aqueous solutions the observation of catalysis in isocyanate formation from ureas or the reverse reaction is not usual owing to the swamping effect of water.



The decomposition of carbamoyl chlorides in weakly basic nonaqueous solvents is relatively insensitive to substituent on the *N*-phenyl group ($\rho = -0.50$)^{71a} consistent with a concerted mechanism (12). In the presence of a tertiary amine the



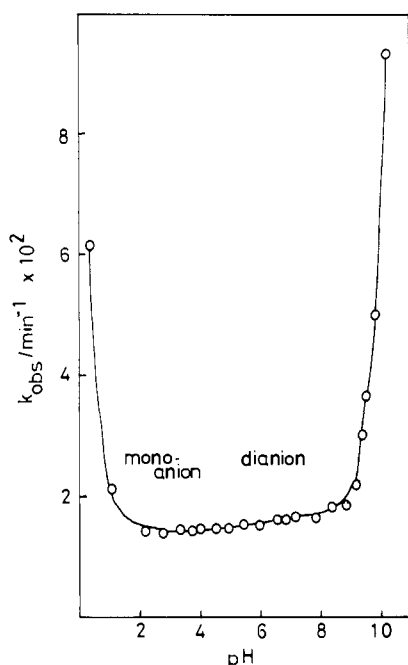


Figure 13. Hydrolysis of carbamoyl phosphate as a function of pH (37°, 0.6 M ionic strength). The figure is redrawn from Figure 2 of C. M. Allen and M. E. Jones, *Biochemistry*, **3**, 1238 (1964).

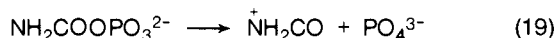
Hammett plot is biphasic with relatively high slope ($\rho = 0.9$) for electron-withdrawing substituents (where k_H/k_D is low); for electron-releasing substituents (with k_H/k_D of about 4) the Hammett slope is 0. These data were considered to be consistent with transition states **13** and **14**, respectively.

8. Carbamoyl Phosphate

The hydrolysis of carbamoyl phosphate has been the subject of a number of investigations,⁷² and interest in reactions of this ester have been stimulated by its important role in the urea cycle in primary metabolism, in pyrimidine biosynthesis, and by the possibility of its playing a role in the urease mechanism. Maximal oxygen-18 incorporation into the phosphate product follows the pH profile (Figure 13) for monoanion concentration indicating P–O cleavage for this species. Neutral and dianion hydrolysis and reaction of hydroxide with dianion gave C–O cleavage.^{72a} Trapping experiments with azide ion show that carbamoyl azide accumulates, but no rate enhancement occurs; small negative entropy of activation, lack of deuterium oxide solvent isotope effect, and lag in initial ammonia release are consistent with an E–A mechanism. Alternatively^{72b} the results were considered consistent



with an $\text{S}_{\text{N}}1$ mechanism (eq 19), and although later workers^{72a} ascribed trapping by azide to action on cyanic acid the fast deprotonation of the amino oxocarbonium ion could in effect be the trapping step. Both hydroxide attack on dianion



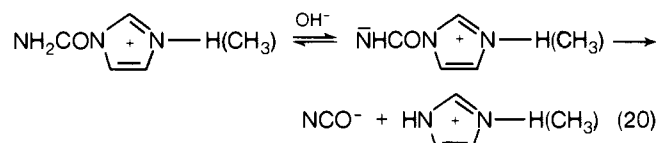
and on monoanion (effectively dianion hydrolysis in the pH region of dianion concentration) yield rate constants which fit a linear free energy relationship between k_{OH} and $\text{p}K_{\text{a}}$ of leaving alcohol constructed from ethyl and substituted phenyl carbamate data⁷³ consistent with an E1cB mechanism for these phosphate species and confirming the earlier conclusions. Azide trapping experiments indicate^{72c} the absence of isocyanate intermediates in the hydrolysis of *N*-4-nitrophenylcarbamoyl phosphate dianion showing that an E–A type mecha-

nism is not operative. P–O cleavage occurs with the dianion, but with the monoanion a small incursion of an E1cB mechanism is shown by the azide trapping experiments. However, the hydroxide rate constant for *N*-phenylcarbamoyl phosphate dianion hydrolysis ($2.73 \times 10^{-2} \text{ M}^{-1} \text{ sec}^{-1}$)^{72d} agrees with that ($8.56 \times 10^{-2} \text{ M}^{-1} \text{ sec}^{-1}$) calculated from the equation: $\log k_{\text{OH}} = 15.2 - 1.34\text{p}K_{\text{a}}$ ⁶⁰ for *N*-phenylcarbamates ($\text{p}K_{\text{a}}$ is the that of the leaving alcohol, and the $\text{p}K_{\text{a}}$ of the conjugate acid of the leaving trianion is taken as 12.3) consistent with an E1cB mechanism involving phenyl isocyanate. The value of k_{OH} for *N*-phenylcarbamoyl phosphate monoanion hydrolysis with hydroxide (calculated from the first-order rate constant for dianion hydrolysis, $\text{p}K_{\text{a}}$ for the phosphate, and K_{w}) comes to $3.17 \times 10^4 \text{ M}^{-1} \text{ sec}^{-1}$ close to the value ($7.42 \times 10^5 \text{ M}^{-1} \text{ sec}^{-1}$) calculated from the equation using as $\text{p}K_{\text{a}}$ for the conjugate acid of the leaving dianion 7.2. Evidence favoring at E1cB mechanism for the *N*-phenylcarbamoyl phosphate hydrolysis is that tris(hydroxymethylamino)methane does not accelerate the hydrolysis, but product analysis indicates significant fractions of the corresponding trapped urea.^{72d}

Carbamoyl phosphate is involved in a number of biological carbamoylation processes, but there is no definite evidence that carbamoylation proceeds via an E–A mechanism. The more important systems are: aspartate transcarbamoylase, ornithine transcarbamoylase (citrulline synthetase),⁷⁴ β -alanine transcarbamoylase,⁷⁵ oxamic acid transcarbamoylase,^{76a} and biotin biosynthesis.^{76b} Transfer of iminocarbamate occurs in argininosuccinate⁷⁷ and guanylic acid synthetase.⁷⁸

Heavy atom (carbon and anhydride oxygen of carbamoyl phosphate) effects suggest that the anhydride bond is unaffected on passage to the transition state, but that bonding to the carbon changes in the aspartate transcarbamoylase reaction;⁷⁹ these results are consistent with the formation of a tetrahedral intermediate rather than with an E1cB mechanism. The absence of ³²P incorporation into carbamoyl phosphate from inorganic phosphate except in the presence of aspartic acid and the absence of ¹⁸O uptake into carbamoyl aspartate or carbamoyl phosphate from enriched water are not consistent with a stepwise process involving a carbamoyl enzyme.⁸⁰ It was concluded that the enzymatic process involved simultaneous action of carbamoyl phosphate and aspartic acid in the synthesis of carbamoyl aspartate. Although the model work with carbamoyl phosphate involves the E–A mechanism of nucleophilic substitution, the later work on analogs indicates that A–E pathways may predominate under certain conditions, and it is likely that the more efficient intramolecular situation at the enzyme-active site promotes the A–E process.

Experiments with *N*-carbamoylimidazole⁷⁴ showed that it was not a good carbamoylating species and probably not involved in ornithine transcarbamoylase as an acyl-enzyme intermediate. The pH dependence for its hydrolysis is sigmoid,⁸¹ and it was proposed that the plateau region involved hydroxide and cation rather than the equivalent neutral species (eq 20); cyanate is the hydrolysis product.^{74,81} A test

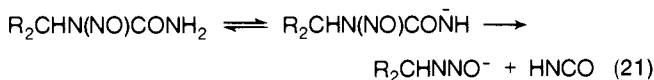


of this mechanism was to synthesize the *N*-carbamoyl-*N*-methylimidazolium ion and measure its hydroxide-catalyzed hydrolysis. Similarity of the two rate constants is judged to be evidence that eq 20 is correct.^{71b} Moreover the rate constant is only tenfold larger than that for the hydrolysis of 4-nitrophen-

nyl carbamate (imidazole and 4-nitrophenolate have conjugate acids with similar pK_a 's).

9. Diazoalkane Synthesis

N-Nitroso-urea decomposition in alkoxide solutions was thought to proceed via an A-E mechanism because alkyl carbamates were isolated as product.⁸² However, the observation that lithium or potassium cyanates are obtained by reaction of *N*-nitroso-*N*-methylurea with butyllithium or potassium *tert*-butoxide provides evidence for an E1cB mechanism.^{49,83}

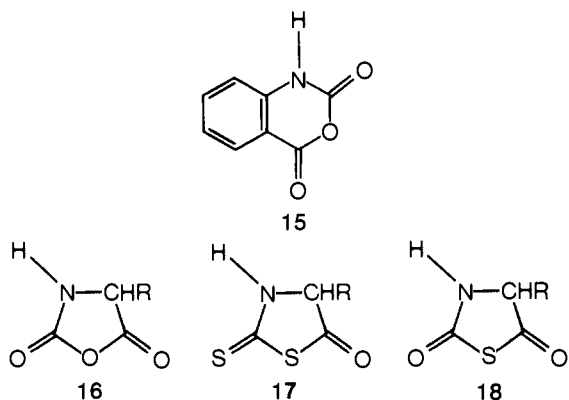


10. Carbamate Protecting Groups

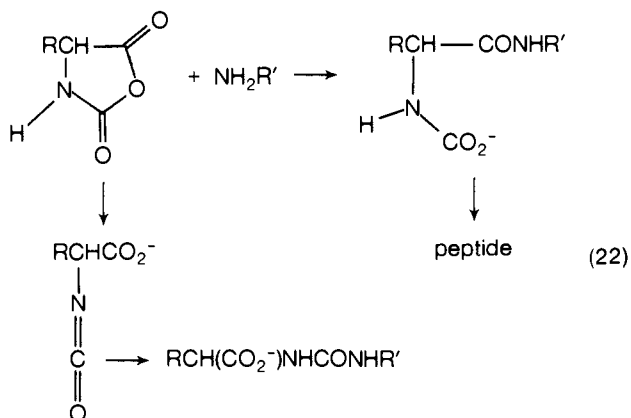
The extreme sensitivity of some carbamates to mild hydrolytic conditions has prompted the use of this group as a protecting agent. Peptide^{84a} and protein^{84b} thiols, sugars,^{84c,d} and protein tyrosines^{84e} have been protected in this way.

11. Cyclic Anhydrides of Carbamic Acid Derivatives

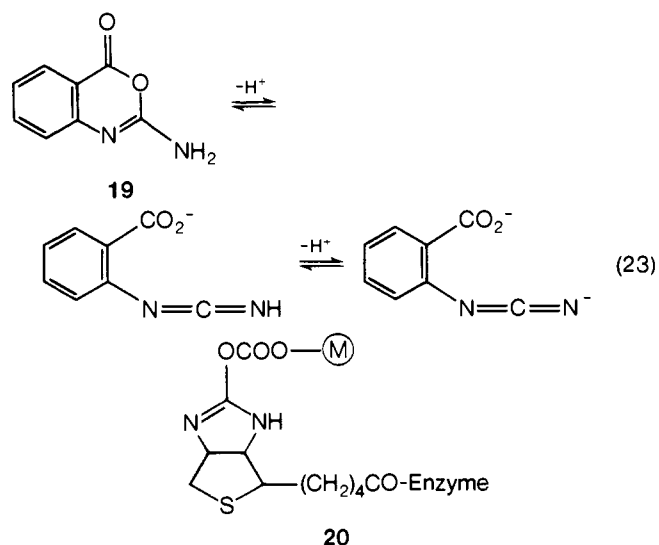
The following heterocyclic compounds could decompose via E-A mechanisms. Isatoic anhydride (**15**) forms ureido products with amines but *N*-methylation of the anhydride destroys this reaction, consistent with isocyanate formation.⁸⁵



Hydrolysis of *N*-carboxyamino acid anhydrides (**16**) involves an isocyanate intermediate according to infrared evidence, but it is uncertain whether this lies on the main hydrolytic pathway.⁸⁶ Peptide and polypeptide synthesis with *N*-carboxyamino acid anhydrides (**16**) and analogs (**17** and **18**)⁸⁷ could involve isocyanate formation as a competing pathway among other possibilities (see eq 22).



A carbodiimide intermediate has been demonstrated in the hydrolysis of 2-amino-4,5-benzo-6-oxo-1,3-oxazine (**19**)⁸⁸ but is probably a blind-alley intermediate. The substrate is a

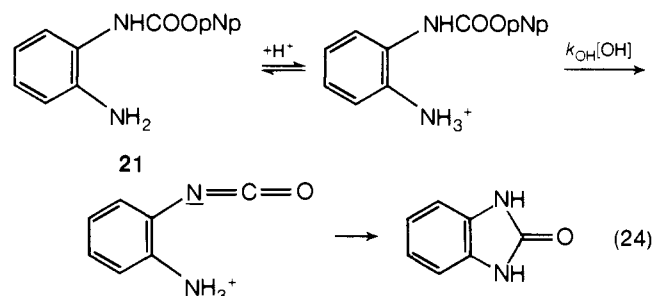


model for *O*-carboxybiotin complexes (**20**); steric requirements preclude a carbodiimide intermediate in the hydrolysis of this derivative; not only would the diimide be strained, say 60° from linearity, but the allene-like stereochemistry would be destroyed by forcing the intermediate to be planar.

12. Intramolecular Reactions

Reactions of carbamates where an intramolecular nucleophilic attack can occur are illustrated in Table IV; it is to be expected that the possibility of an intramolecular nucleophilic attack reduces the possibility of an E-A mechanism, and the results of the following studies show that control of mechanism is very sensitive to the nature of the intramolecular nucleophile.

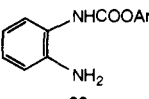
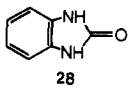
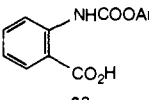
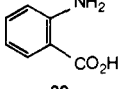
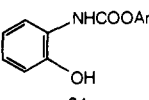
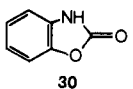
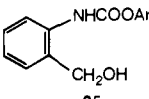
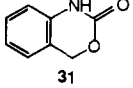
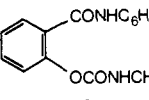
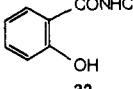
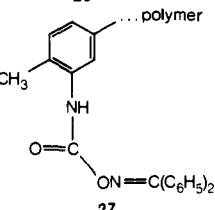
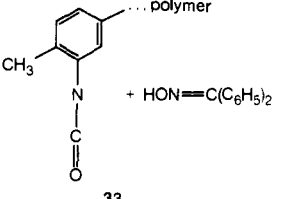
Cyclization of 4-nitrophenyl *N*-(2'-aminophenyl)carbamate (**21**)⁶¹ yields initially the isocyanate which is trapped by the amino function. This mechanism is consistent with unit slope of the pH profile for cyclization between pH 5 and 8 which indicates that attack of free amine is not a rate-determining step and with trapping experiments with 4-chloroaniline which do not enhance the rate yet give substantial quantities of the competing product urea. Trapping experiments from pH 5 upwards do not say anything about the mechanism involving a shoulder in the pH profile corresponding to a pK_a of 3.87: a mechanism giving rise to the shoulder could involve hydroxide attack (E-A) on the ammonium ion as in eq 24. The value of



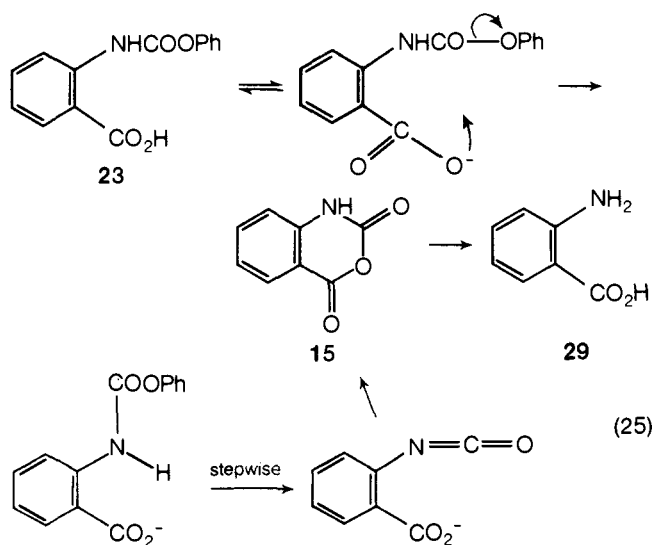
k_{OH} for this process ($3.34 \times 10^6 M^{-1} sec^{-1}$) may be calculated from the Hammett equation⁶¹ and $\sigma = 1.7$,⁹⁰ the apparent hydroxide ion rate constant ($k_2'K_{a1}'$ in ref 61) has a value $6.3 \times 10^6 M^{-1} sec^{-1}$, in good agreement with the calculated value assuming the isocyanate mechanism.

Reaction of phenyl (*N*-2-carboxyphenyl)carbamate (**23**) at neutral pH's to yield anthranilic acid (**29**) proceeds via isatoic anhydride.⁸⁹ The pH dependence of the decomposition is sigmoidal with a hydroxide term above pH 11 and inflection at the pH corresponding to carboxyl ionization. In the region of

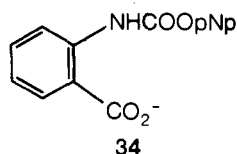
TABLE IV. Intramolecular Transfer of the Carbamate Group

Reactant	Product	Mechanism	Ref
		Hydroxide term E-A Neutral term E-A	61
		Hydroxide term, possibly E-A Anionic term A-E	89
		Hydroxide term, possibly E-A Anionic term A-E	91
		Hydroxide term A-E for R; E-A for Ar	92
		Possibly involves MeNCO	93
			94

pH 2–9 the *N*-methyl analog of **23** does not hydrolyze at a significantly different rate from that for the parent, and together with the deuterium oxide solvent isotope effect ($H/D = 1.2$) an A–E mechanism is probable.

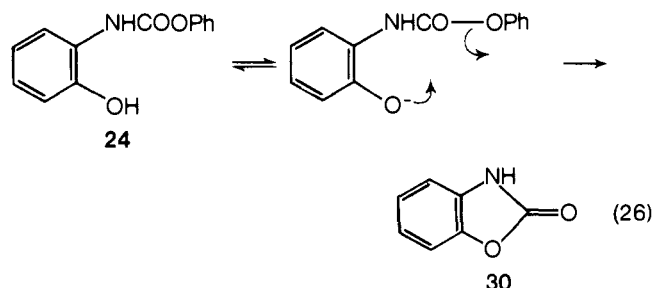


The value of σ for CO_2^- is uncertain⁹⁰ and could be zero or 0.31; we can calculate, using the Hammett relationship for 4-nitrophenyl (*N*-substituted)phenylcarbamates,⁶¹ the value of k_{OH} for isocyanate formation from **34**. Using the Hammett ρ selectivity of +2.86 for change of substituent on the leaving phenol group, we can then estimate the k_{OH} value for phenyl



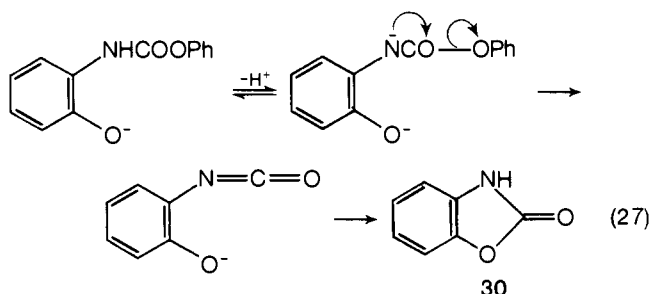
N-(2'-carboxyphenyl)carbamate (anion), and this is 63.4 or $100 \text{ M}^{-1} \text{ sec}^{-1}$ depending on the σ value employed. The observed value of $1.0 \text{ M}^{-1} \text{ sec}^{-1}$ ⁸⁹ is low, but the 100-fold difference does not rule out the intermediate formation of isocyanate in the alkaline range of pH; trapping experiments were not carried out in this pH range, and it is quite conceivable that a change in the mechanism should occur.

Intramolecular nucleophilic attack by the phenoxide group to yield 2-benzoxazolinone (**30**) from phenyl *N*-(2-hydroxyphenyl)carbamate (**24**) is consistent with intramolecular nucleophilic substitution.⁹¹ The pH dependence is sigmoid with an inflection at the $\text{p}K_{\text{a}}$ of the phenol and a hydroxide term predominating above pH 12. The plateau rate constant for the corresponding *N*-methyl analog is about tenfold greater, consistent with the A–E mechanism (eq 26); the slightly larger



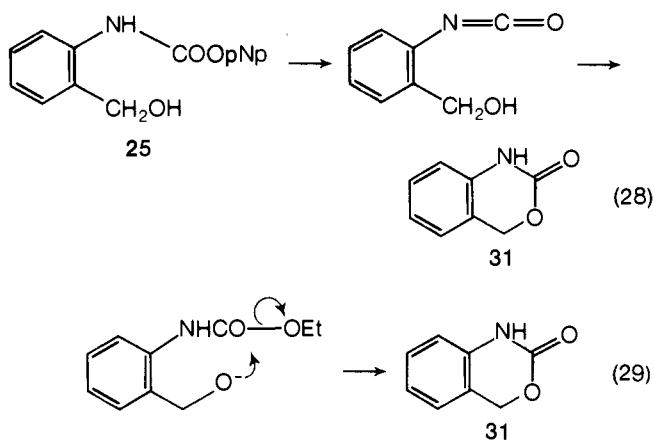
value for the *N*-methyl analog is possibly due to steric interactions between methyl and ortho hydrogens altering the conformation of the carbonyl group.⁹¹ The hydroxide term for the hydrolysis of the anionic species can be calculated assuming the mechanism involves isocyanate using the rate constant for phenyl *N*-phenylcarbamate under similar conditions ($54.2 \text{ M}^{-1} \text{ sec}^{-1}$) from ref 4 1a, the ρ value for substituents on the *N*-phenyl ring (+0.64),⁶¹ and a σ value of -1.00 ⁹⁰ for the oxyanion at the ortho position; the resulting value ($12.6 \text{ M}^{-1} \text{ sec}^{-1}$) is close to the observed value of k_{OH} (5.14 M^{-1}

sec^{-1}) consistent with an isocyanate intermediate in the alkaline region of pH (eq 27). As in the carboxylate participation,

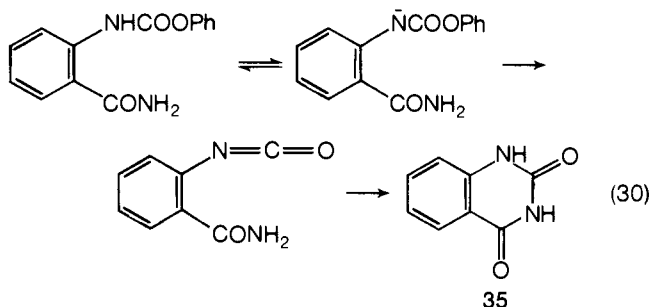


intermolecular hydroxide attack via the E-A mechanism competes successfully with the intramolecular A-E mechanism at high basicity.

Cyclization of esters of 2-hydroxymethylcarbanilic acid (**25**) and the *N*-methyl analog gives relatively stable benzoxazin-2-ones (**31**).⁹² The pH profile for phenol release shows hydroxide terms but no inflections, and the 4-nitrophenyl ester has a rate constant similar to the hydrolysis of 4-nitrophenyl *N*-phenylcarbamate but about 10^4 -fold larger than the cyclization of the *N*-methyl analog; the ethyl ester cyclizes some 10^6 -fold faster than the hydrolysis of ethyl *N*-phenylcarbamate (which is known to involve as rate-determining step the formation of isocyanate),⁶⁰ and the ethyl ester of the *N*-methyl analog (of the acid of **25**) has a cyclization rate constant similar to the parent. It was proposed⁹² that whereas the cyclization of the 4-nitrophenyl ester (**25**) involves an isocyanate intermediate (eq 28), the ethyl ester possesses an A-E mechanism (eq 29).



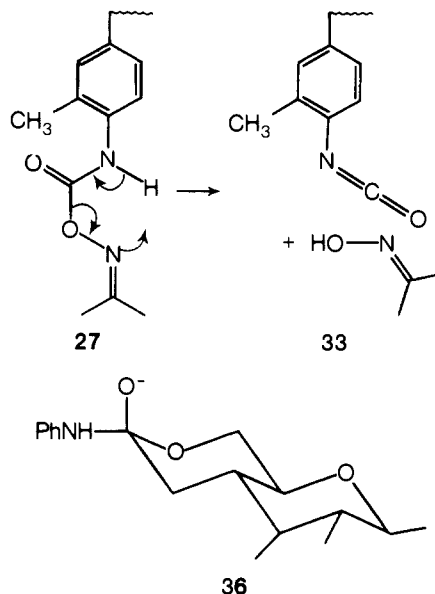
The hydrolysis of salicylanilide carbamates (**26**)⁹³ in alkali gave salicylanilide and in ethanol-water mixtures ethyl *N*-methylcarbamate; the formation of the latter strongly suggests methyl isocyanate as an intermediate, and the ratio of hydroxide ion rate constants for monomethyl- and dimethylcarbamoyl salicylanilides (see structure **26** in Table IV) in excess of 10^6 -fold is consistent with this conclusion. In a related paper^{93b} the formation of quinazolinones (**35**) pro-



ceeds via trapping of the isocyanate by the amide rather than via initial nucleophilic attack of amide on the carbamate. Distinction between the two mechanisms was made by comparing the hydroxide ion rate constants for the *N*-methyl carbamate and the parent—some 100-fold difference.

An interesting example of intramolecular catalysis involves the formation of isocyanates for cross-linking in polymers by using carbamate monomers possessing a basic function (**27** → **33**).⁹⁴

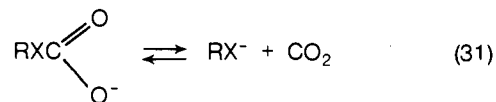
Migration from the 4- to the 6-hydroxyl in *N*-phenylcarbamoyl protected carbohydrates is postulated to involve an A-E mechanism possessing a tetrahedral adduct (**36**).^{84c}



C. Carbonic Acid Derivatives

1. Decarboxylation

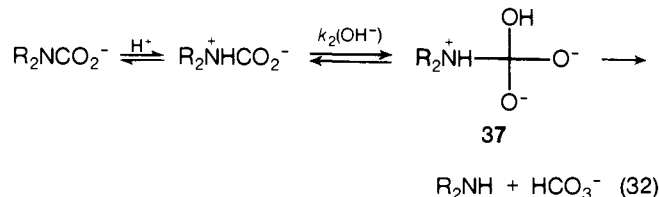
Carbonic acid derivatives of which a carboxylic acid is a special case decompose to yield carbon dioxide (or its analog), and the elimination mechanism (eq 31) has never been



seriously questioned. The subject of decarboxylation of carboxylic acids where RX^- is a stabilized carbanion comes under this heading; this topic has been the subject of extensive reviews⁹⁵ and will not be treated here.

2. Carbamic Acids

A recent kinetic study⁹⁶ has disposed of the A-E mechanism for *N*-arylcarbamate decomposition (eq 32); the value of

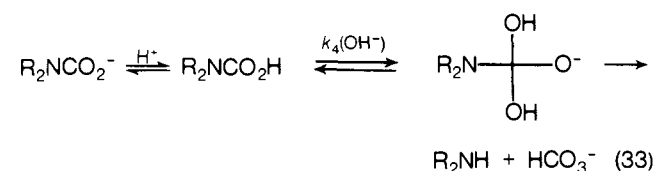


k_2 was estimated to be $6 \times 10^9 \text{ M}^{-1} \text{ sec}^{-1}$ assuming this mechanism for 2-ethoxyphenylcarbamic acid and is many orders of magnitude larger than the rate constant for attack of hydroxide ion on 4-nitrophenyl *N,N*-dimethylcarbamate ($3 \times 10^{-4} \text{ M}^{-1} \text{ sec}^{-1}$). Attack of hydroxide ion on the neutral carbamic acid (eq 33) can be similarly eliminated.

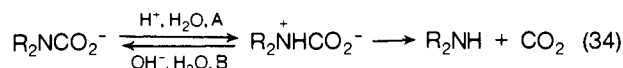
TABLE V. Decomposition of Alkyl Hydrogen Carbonates

ROH	pK _a	K _{eq} /M	k _{decomp} /min ⁻¹	Ref
Allyl	15.52	10 ^{1.61}	10 ^{-2.23}	102a
Glycol	14.77	10 ^{1.44}	10 ^{-2.2}	102b
2-Chloroethyl	14.31	10 ^{2.05}	10 ^{-1.5}	102b
Triethanolamine		10 ^{1.0}	10 ^{-2.14}	102g
2-Tetrahydrofurfuryl		10 ^{1.64}	10 ^{-2.13}	102h
Furfuryl	15.54 ^a	10 ^{1.93}	10 ^{-1.27}	102h
Methyl	15.54	10 ^{1.08}	10 ^{-2.62}	102a
Ethyl	16.00	10 ^{1.36}	10 ^{-2.99}	102c, 102j
Glycerol	14.4	10 ^{1.35}	10 ^{-2.89}	102k
Benzyl	15.08 ^a		10 ^{-1.85}	102a
Cyclohexyl			10 ^{-3.39}	102e
<i>n</i> -Butyl	16.06		10 ^{-3.05}	102i
Isobutyl	16.25 ^a		10 ^{-3.52}	102i
<i>sec</i> -Butyl			10 ^{-3.72}	102i
<i>n</i> -Propyl	16.00		10 ^{-3.14}	102f, 102d
Isopropyl	16.57 ^b		10 ^{-3.44}	102f

^a Calculated using pK_a = 15.9 - 1.42α* (P. Ballinger and F. A. Long, *J. Am. Chem. Soc.*, **82**, 795 (1960)). ^b Calculated by S. Takahashi, L. A. Cohen, H. K. Miller, and E. G. Peake, *J. Org. Chem.*, **36**, 1205 (1971).

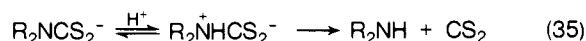


Decarboxylation of *N*-arylcabamates is oxonium ion catalyzed, and the pH dependence exhibits a plateau at low pH's; the rate constants possess a Bronsted type relationship^{96,97} with the pK_a of the ammonium ion which exhibits two linear portions consistent with a changeover in rate-determining step (Figure 14). Carbamates from weakly basic and strongly basic amines decarboxylate with general acid and specific acid catalysis, respectively,⁹⁶⁻⁹⁸ consistent with a mechanism where carbamates from weakly basic amines involve rate-limiting proton transfer while strongly basic amines involve rate-limiting decarboxylation (eq 34); urea synthesis from isocyanic acid and amines exhibits strikingly similar behavior.^{71a,b}

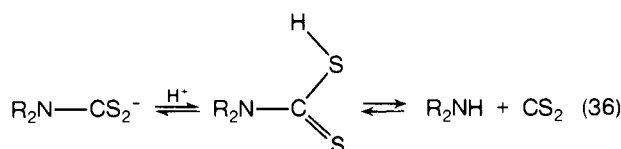


3. Dithiocarbamic Acids

Dithiocarbamic acid decomposition is acid catalyzed but becomes pH independent at low pH's,⁹⁹ consistent with the neutral or zwitterionic species being the reactant. Some workers prefer the zwitterion as the active species (eq 35),¹⁰⁰ while others^{101a} argue for an intramolecular proton



transfer in a neutral species (eq 36). If the mechanism is to



be distinguished from that of eq 35, the proton transfer must be included in some form of four-center transition state. The

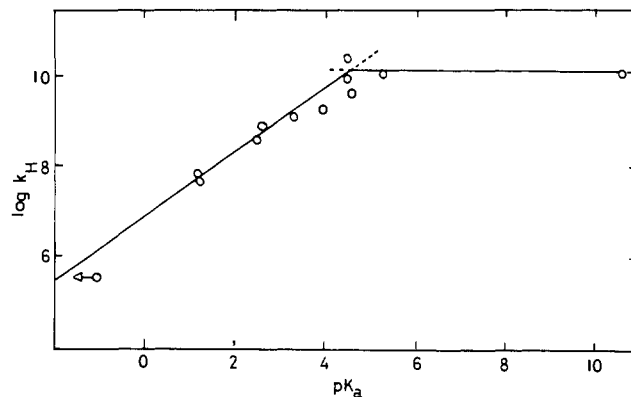
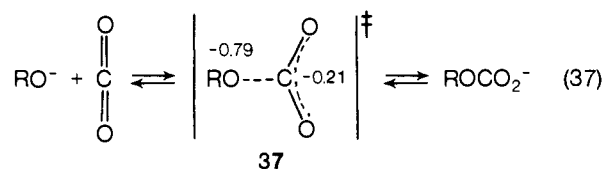


Figure 14. Effect of structure on the rate of hydronium ion catalyzed decarboxylation of *N*-substituted carbamic acids in water at 25°. All amines are aromatic except the one at high pK_a (cyclohexylamine) and at low pK_a (imidazoline). The figure is redrawn from Figure 5 of S. L. Johnson and D. L. Morrison, *J. Am. Chem. Soc.*, **94**, 1323 (1972).

stability of dithiocarbamic acids in nonpolar solvents tends to favor the zwitterionic mechanism.^{101b} An interesting review on dithiocarbamates has been published.^{101c}

4. Carbonic Acid Half Esters

Faurholt and his coworkers¹⁰² have studied in detail the decomposition of carbonate half esters; Table V collects some of the data which indicate that the unimolecular decomposition of the half ester is strongly related to the pK_a of the departing alcohol (a β_{lg} for the departure of the alcoholate may be calculated from Faurholt's data) consistent with a transition state with extensive C-O-alkyl fission. Although Faurholt's work yields data close to the presently accepted values, it was obtained over a period of about 30 years and apparently not with the view of obtaining Bronsted coefficients; more recent work with this aim (unpublished observations of Professor C. K. Sauers at Rutgers, 1973) provides better values for the Bronsted coefficients in the reaction of alkoxide ions with carbon dioxide (eq 37) (β_{nuc}, forward direction, 0.3; β_{lg}, reverse direction, -1.1; β, overall equilibrium, +1.4). The apparent charge on the various atoms in the transition state may be calculated from Sauers' values and from this follows a knowledge of the extent of bond fission (37).

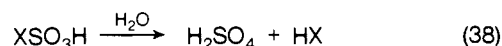


Mechanisms similar to eq 35 and eq 36 have been proposed for the decomposition of neutral *O*-alkyl dithiocarbamate esters.^{103b,c}

D. Sulfuric Acid Derivatives

1. Sulfates

The hydrolysis of monosulfate esters (X = OR in eq 38) is



part of a large class of reactions which includes the hydrolysis of Bunte salts (X = SR),¹⁰⁴ thiosulfate ion (X = S⁻),¹⁰⁵ halosulfonic acids (X = Hal), acyl sulfates (X = RCO-), sulfamic acids, hydroxylamine *N*- and *O*-sulfonates,¹⁰⁶ and the exchange of the oxygens of the sulfate ion in aqueous solution.¹⁰⁷ The pH dependencies of the hydrolyses, often com-

TABLE VI. Structure-Function Relationships for Reactions of Sulfate Derivatives

Substrate	Conditions	$T/^\circ\text{C}$	pK_a range	β_{lg}	Ref
Leaving Group Variation					
$\text{XC}_6\text{H}_4\text{OSO}_3\text{H}$	$N/24$ HCl	48.6	7.0-10.3	-0.25 ^c	121 ^b
		78.7	7.0-10.3	-0.21 ^c	121 ^b
	1.03 <i>M</i> HCl	45	4.0-8.4	-0.22	115
		25	4.0-8.4	-0.26	115
RSSO ₃ H	0.4 <i>M</i> HClO ₄	69.7	5.0-10.3	+0.12	104 ^b
$\text{XC}_6\text{H}_4\text{NHSO}_3\text{H}$	0.09 <i>M</i> HCl	45	3.8-5.4	-0.27 ^a	<i>e</i> ^b
		75	3.8-5.4	-0.36	<i>e</i> ^b
RNHSO ₃ H	1.41 <i>M</i> HCl	99.96	4.6-10.6	-0.57	<i>f</i> ^b
$\text{XC}_6\text{H}_4\text{OSO}_3^-$	Neutral	100	4.0-8.4	-1.2	115
	Cyclodextrin at pH 10	50.3	2 points	-1.5	132
$\text{C}_6\text{H}_5\text{SO}_2\text{OC}_6\text{H}_4\text{X}$	Alkaline	50	7.0-10.2	-0.93	<i>g</i> ^b
Nucleophile Variation					
4-Nitrophenyl- sulfate	Amines	35	3.0-12	β_{nuc} +0.13	108 ^e
	Thiophenols in DMF	90	6.0-7	+0.05 ^d	120 ^b

^a Imprecise, Brønsted scattered. ^b Calculated from data in the reference. ^c Ortho groups deviate. ^d Since these reactions were carried out in DMF at 90° the ionic states of the equimolar reactants used are unknown and the interpretation of β_{nuc} unclear. ^e F. L. Scott and W. J. Spillane, *Chem. Ind. (London)*, 1999 (1967). ^f J. D. Capps and M. D. Bentley, *J. Org. Chem.*, **33**, 1295 (1968). ^g R. V. Vizgert, *Zh. Obshch. Khim.*, **28**, 1873 (1958).

plex, usually exhibit portions due to a spontaneous reaction of the monoanion and an acid-catalyzed limb.¹⁰⁸ The spontaneous reaction is usually characterized by S-O cleavage, while the acid reactions are complicated by C-O fission.¹⁰⁸

Mechanistic insight into this area is important to the chemistry of both sulfonation and biological sulfate transfer. The former has been thoroughly reviewed by Gilbert¹⁰⁹ and examples are the use of sulfur trioxide complexes of organic bases (e.g., pyridine-*N*-sulfonate), halosulfonates, and sulfamic acid in the sulfonation of steroids.¹¹⁰ Sulfate esters play an important role in sulfate transfer,¹¹¹ in the biological reduction of sulfite,^{111d,112} in steroid isolation,¹¹³ and in carbohydrate chemistry.¹¹⁴

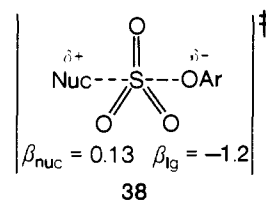
Monoanion Solvolysis. Solvolysis of the monoanion in neutral solution has been studied extensively only for aryl sulfate monoesters^{115,116} and is observable only for substrates with powerful leaving groups—a consequence of the high Brønsted exponent for leaving group variation ($\beta_{\text{lg}} = -1.2$, see Table VI). No spontaneous reaction for sulfamate ions has been observed since the leaving species would be R_2N^- , but the neutral solvolyses of tertiary amine-sulfur trioxide adducts have been extensively studied.¹¹⁷

The magnitude of β_{lg} is consistent with a markedly E1 transition state. The slow solvolysis of fluorosulfate ion,^{118a,b} due presumably to the strength of the S-F bond, is the basis of the fluoride inhibition of some sulfate reactions^{117c} and its use in trapping reactive sulfate intermediates;¹¹⁹ in addition, it indicates that factors other than the pK_a of the conjugate acid of the departing leaving group can control reactivity.

Nucleophilic attack of water on the monoanion has been excluded by specific electrolyte studies,¹¹⁵ but it has several times been noted that the difference between powerful aquation of a transition state and weak nucleophilic interaction of water with the monoanion may be semantic rather than real.^{115,116} Markedly negative entropies of activation and a positive deuterium oxide solvent isotope effect ($\text{H}/\text{D} = 1.26$) for solvolysis of the 4-nitrophenyl sulfate monoanion indicate considerable differential solvation of ground and transition states. The low Brønsted exponent for amine attack on the 4-nitrophenyl ester ($\beta_{\text{nuc}} = 0.13$; see Table VI) indicates weak interaction with nucleophile. Similar low dependence on base strength was observed for the attack of thiophenols.¹²⁰

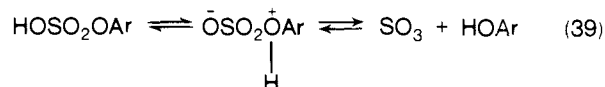
Catalysis. We have seen that the transition state for mono-

sulfate hydrolysis involves advanced bond fission but minimal interaction with the nucleophile; that is, the transition state approximates sulfur trioxide plus phenolate ion (38). Thus ca-

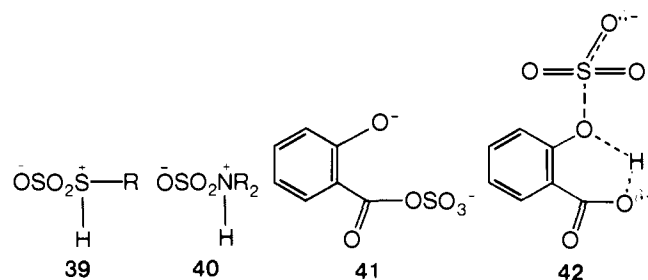


talysis may be expected from influences which stabilize the transition state (i.e., products) or improve the leaving group; little catalysis would be expected from activating the nucleophile.

Acid catalysis in the hydrolysis of aryl monosulfates has been explained by an A1 mechanism involving zwitterion formation.^{108b-d,115,121-125} This provides both an internal nucleophile in the form of an oxyanion and an activated leaving group. Such a scheme (eq 39) is supported by the small posi-



tive or zero entropies of activation,^{121,124} the inverse deuterium oxide solvent isotope effect,¹²⁵ and the steric effects of bulky ortho groups.^{121,124} The overall sensitivity to the basicity of the leaving group is low ($\beta_{\text{lg}} = -0.23$)^{121,125} since the Brønsted exponent is composite. Bunte salts¹⁰⁴ and sulfamic acids¹²⁶ may also hydrolyze in acid via zwitterions (39 and 40), and the latter hydrolyses have been recently reviewed.¹²⁶ Aryl selenates solvolyze via an A2 route.¹²⁷



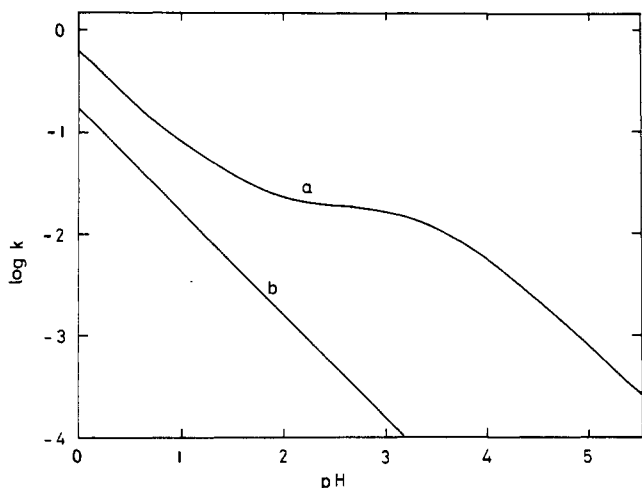
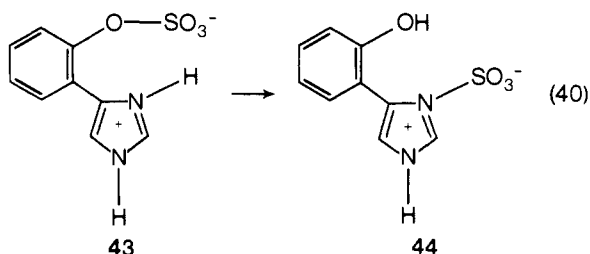
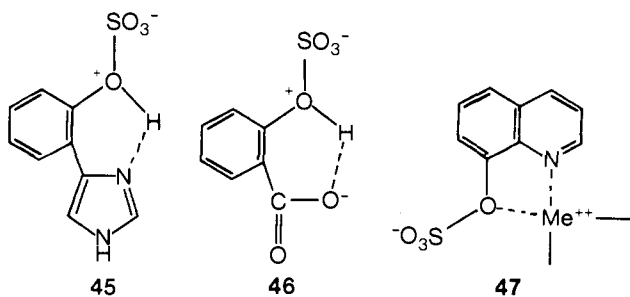


Figure 15. The pH-rate profiles (35°) for the hydrolysis of (a) salicyl sulfate and (b) 4-carboxyphenyl sulfate. The figure is redrawn from Figure 2 of S. J. Benkovic, *J. Am. Chem. Soc.*, **88**, 511 (1966).

Acid catalysis may also be provided intramolecularly: the carboxylic acid moiety of salicyl sulfate provides catalysis to expel the phenol and leave sulfur trioxide (Figure 15); the para isomer shows no such catalysis.¹²⁵ An acyl sulfate intermediate (41) was excluded using a hydroxylamine trapping experiment,¹²⁸ and the mechanism appears to involve intramolecular general acid catalyzed phenol expulsion (42). Comparison of trapping experiments with salicyl sulfate and sulfur trioxide in mixed alcoholic solvents was not definitive as to mechanism.¹²⁹ A similar case is the hydrolysis of 2-(4(5)-imidazolyl)phenyl sulfate.¹³⁰ A mechanism involving the *N*-sulfate (eq 40) is not absolutely ruled out because the rate constant for

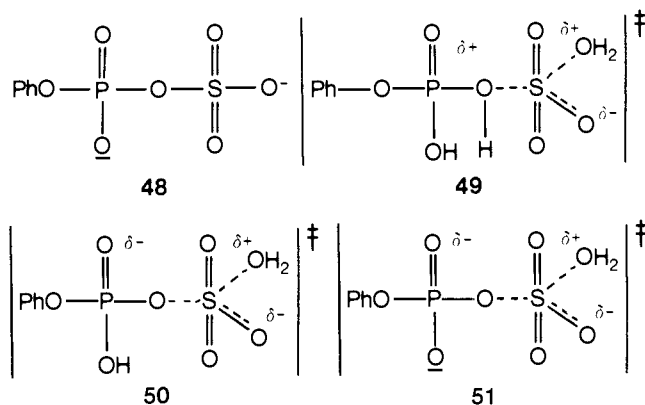


N-sulfate hydrolysis is larger than the hydrolysis rate for the starting ester (43). Fluoride ion does not trap any intermediate as fluorosulfonate in the hydrolyzing mixture but does trap the synthesized *N*-sulfate (44). Although fluoride ion is known to trap *N*- but not *O*-sulfate,^{117c} it has a low efficiency compared with water in trapping *N*-sulfonates. It is therefore only possible to place a limit on the amount of reaction proceeding via the *N*-sulfate (<50%). Catalysis of the hydrolysis of 8-hydroxyquinoline *O*-sulfate by intramolecular acid catalysis and by metal ions has been observed.¹²³ In none of the above cases is the nature of the electrophilic catalysis clear; the neighboring acid group may serve to aid proton transfer to the leaving group or to stabilize a zwitterionic transition state (45, 46, 47).



Other modes of catalysis investigated have been micellar^{117b,131} and cyclodextrin induced.¹³² The former case may be due to ground-state destabilization or transition-state stabilization depending on the conditions. It is interesting that the acid catalysis of sulfate hydrolysis becomes more pronounced in solvents of low polarity, especially ethers.^{108b,c} Moist dioxane increases the rate of hydrolysis of alkyl sulfates 10⁷-fold relative to water^{108c} possibly because of specific solvation of the zwitterionic transition state; sulfur trioxide, which resembles the transition state (see structure 38), is well known to give etherate complexes.¹³³

Acyl Sulfates. The sulfate donor in some biological transfers (e.g., formation of chondroitin sulfate, cerebrosulphatides, and phenolic sulfates)¹³⁴ has been shown to be 3'-phosphoadenosine 5'-phosphosulfate (PAPS). Model studies¹³⁵ on phenyl phosphosulfate (48) have indicated that the S-O fission process of the mono- and dianions and neutral species involve transition states with marked sulfur trioxide character (structures 49, 50, and 51) and that 48 acts to sulfylate[†] rather than phosphorylate only by a factor of 10.^{135a} Acetyl sulfate transfers the acetyl group as shown by the predominance of C-O cleavage; no sulfylation is observed with this ester.^{135a}



Biological Implications. The transition states for cyclodextrin catalyzed and aryl sulfate hydrolyses have been shown to approximate the normal solvolytic transition state with appreciable sulfur trioxide character.¹³² In addition, the rates of sulfylation of *Aspergillus* aryl sulfatase by aryl sulfates correlates with the corresponding acid catalysis rates, again pointing to a "sulfur trioxide" transition state.¹³⁶ Such observations indicate that implications drawn from aqueous sulfate chemistry may be a fair guide to sulfate chemistry at biological sites. The low Bronsted coefficient for nucleophile variation indicates that an active site may play little part in enzyme mediated sulfate transfer. The high dependence on leaving group hints that activation may be in this portion of the system.

The evidence indicates that the metabolism of selenium in the cell may share some of the enzymic paths of sulfur metabolism.¹³⁷ In view of this the observed A1 reaction of sulfates under acid conditions of catalysis^{108b-d,115,121-125} contrasts sharply with the A2 reactions of the corresponding selenates.¹²⁷

2. Amino Sulfonates

Until recently there was but little kinetic activity in this area and elimination-addition routes had been observed only for highly activated *N*-monosubstituted sulfamoyl chlorides in nonaqueous solutions. Thus, *N*-ethylsulfamoylamine (EtN=SO₂), produced by dehydrohalogenation of ethylsulfamoyl

[†] We use "sulfylate" here by analogy with the general term coined for all "acylations" involving phosphorus acids (namely "phosphylate"). The use of "yl" denotes the transfer of an acyl function.

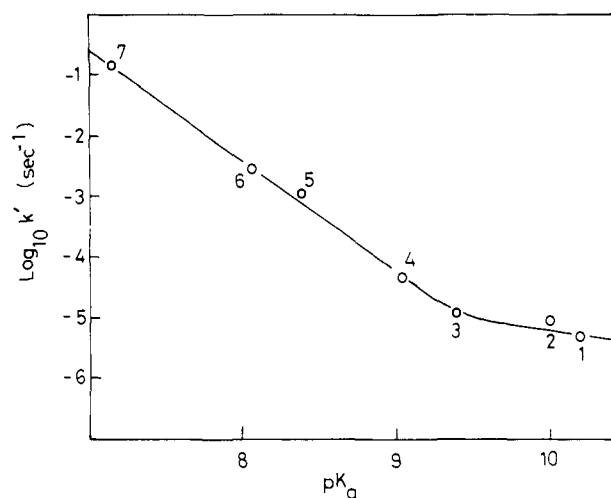
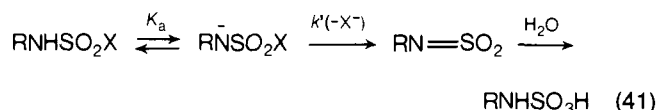


Figure 16. Dependence of k' (see eq 12) on pK_a of the phenol leaving group (60°, 50% ethanol-water). Legend: (1) 4-methoxyphenyl, (2) phenyl, (3) 4-chlorophenyl, (4) 3-chlorophenyl, (5) 3-nitrophenyl, (6) 4-acetylphenyl, (7) 4-nitrophenyl. The figure is redrawn from Figure 2 of ref 71b.

chloride, was trapped rapidly either by nucleophiles or by cycloaddition.¹³⁸ Other similar dehydrohalogenations have been reported^{139,140} and sulfamoyl chloride itself reacts with aniline in the presence of sodium hydroxide to form the sulfanilide.¹⁴¹ The pronounced acidity of the N-H group in monosubstituted aminosulfonates (RNHSO₂X) was indicated by salt formation (e.g., EtO₂NSO₂N⁺Et₃,¹³⁸ NH₂SO₂NSO₂N⁺Et₃,¹³⁹ and ArN-SO₂N₃R₃N⁺H¹⁴²) and opened the possibility of the E1cB route for hydrolysis (eq 41). The apparent pK_a 's of some sub-



stituted phenyl *N*-methylaminosulfonates in 50% aqueous ethanol are predictable from a Hammett relationship ($\rho = 2.54$);¹⁴³ the pK_a of the 4-nitrophenyl ester fits the value derived kinetically from eq 42. Since k' (the "alkaline" plateau

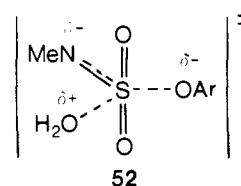
$$k_{\text{obsd}} = k_{\text{H}_2\text{O}} + k'/(1 + K_W K_a[\text{OH}]) \quad (42)$$

rate constant) is 1000-fold larger than $k_{\text{H}_2\text{O}}$, it is unlikely that k' reflects attack of water on the conjugate base of the ester.¹⁴³ The rates of hydroxide catalysis for aryl esters of benzene- and 4-phenylbenzenesulfonic acids correlate with Hammett σ values and possess ρ values of +2.75 and 2.56, respectively.¹⁴⁴ However, the alkaline plateau rate constants for aryl *N*-methylaminosulfonates show high correlation slopes with Hammett σ^- constants ($\rho_{k'} = +4.04$)¹⁴³ and the ρ value for the apparent reaction of hydroxide with the neutral esters ($k_{\text{OH}^-} = k'K_a/K_W$) is 6.58 (corresponding to a Bronsted β_{lg} of -2.3). Such high selectivity for departing species and σ^- dependence is typical of an E1cB pathway. The lower value of β_{lg} (~ -1.03) and approximately 10²-fold decreased reactivity toward hydroxide ion for aryl *N,N*-dimethylaminosulfonates supports this assignment. Absence of an effect of amine nucleophiles on the rate of release of phenol from the monomethyl derivative while yielding totally the amide product also supports the E1cB path.¹⁴³ The Bronsted exponent (β_{lg}) for aryl methylaminosulfonates changes sharply when the pK_a (lg) exceeds 9.4 (Figure 16), and this could be due to a changeover in mechanism to stronger participation of the nucleophilic water in the transition state for phenolate expulsion.⁵² Such an explanation is consistent with the phenols of higher pK_a needing extra driving force for their expulsion than the active phenols.^{143b}

TABLE VII. Rate Constants for Reaction of Hydroxide Ion with RNHSO₂N₃^a

R	$k_{\text{OH}^-}/(M^{-1} \text{sec}^{-1}) \times 10^{-6}$
Methyl	2.40
<i>n</i> -Propyl	2.63
Isopropyl	1.87
Isobutyl	2.20
<i>tert</i> -Butyl	4.80

^a Data taken from ref 142.

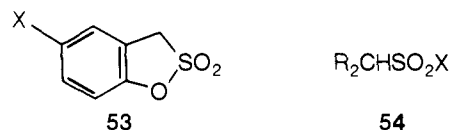


The base-catalyzed hydrolysis of some hypotensive sulfamoyl azides was suggested¹⁴² to involve an E1cB mechanism, and in view of the results of the work on the methylamino esters¹⁴³ this interpretation is probably correct. Results of the work on *N*-substituted azides¹⁴² can be interpreted¹⁴³ as evidence for the E-A process in hydrolysis: the absence of a substituent effect (Table VII) is not consistent with an A-E mechanism. The slight acceleration of the *tert*-butyl over the methyl substituent may reflect steric release in the E1cB transition state although differences in the solvation cannot be dismissed.

3. Alkanesulfonate Derivatives (R₂CHSO₂X)

In this section a mechanistic view will be presented of sulfonyl transfers of alkanesulfonate esters, halides, and sultones; brief mention will also be made of some analogous processes from other regions of sulfur chemistry. Reviews of arenesulfonate esters,¹⁴⁴ fluorosultones,¹⁴⁵ sultones,¹⁴⁶ and sulfenes¹⁴⁷ have appeared but the kinetic data available are not extensive. Reactions of alkyl esters of sulfonic acids will not be included since solvolysis is via C-O fission.¹⁴⁸

Sulfonyl derivatives command relatively important roles in bioorganic chemistry. Thus, α -toluenesulfonyl fluoride has been used to modify the active sites of serine proteases,¹⁴⁹ and 2-hydroxy-5-nitro- α -toluenesulfonic acid sultone (**53**, X = NO₂) sulfonylates several enzymes with 1:1 stoichiometry^{150,151} and is a useful active-site titrant.¹⁵² The large (>10⁵-fold) cyclic/acyclic rate ratio for alkaline hydrolysis of **53** (X = H) and phenyl α -toluenesulfonate¹⁵³ demands de-



tailed mechanistic consideration. When hydroxyl and amino groups cannot be sulfonylated with aromatic sulfonyl chlorides (e.g., in some oximes,¹⁵⁴ and in *N-tert*-butoxycarbonyl-2-(2-hydroxythiazolidine) in Woodward's cephalosporin-C synthesis¹⁵⁶), mild, rapid, introduction via sulfenes may be possible.

The marked lability of hydrogen atoms α to the sulfonyl group (e.g., in **53** and **54**) is fundamental to the chemistry of these compounds. Salts have been isolated (e.g., CH₃SO₂CH-SO₂N⁺Et₃,¹⁴⁷ CF₃CFSO₂C₅H₆N⁺¹⁴⁶) but only where the carbanion is highly stabilized. Many studies of exchangeability with solvent deuterons^{146,157-164} have been made and the C-H pK_a 's of **53** (X = H) and phenyl α -toluenesulfonate in dimethyl sulfoxide are 15.6 and 18.0, respectively.¹⁶⁵ Such

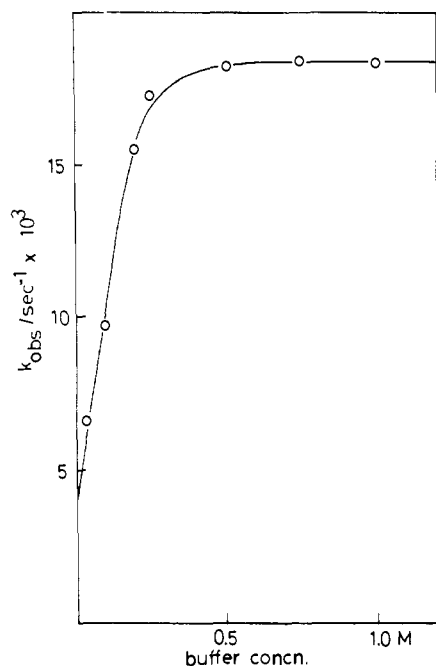
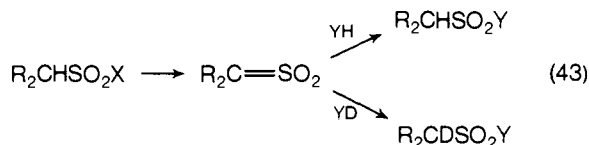


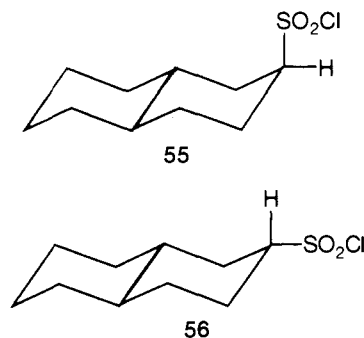
Figure 17. Rate constants for the release of 4-nitrophenol from 4-nitrophenyl toluene- α -sulfonate as a function of buffer concentration: buffer, diethylamine; fraction of base, 0.2; pH, 10.26; ionic strength, 1 M; 25°. The figure is redrawn from Figure 2 of ref 173.

work led to suggestions of elimination-addition routes via sulfenes (eq 43). Full discussions of the sulfene-mediated base



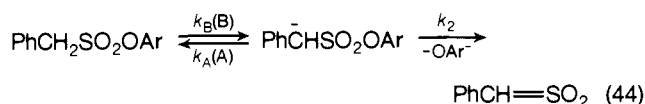
solvolysis of sulfonyl halides are already available (notably from the groups of King¹⁵⁹⁻¹⁶¹ and Truce^{162,163}) and only a brief summary follows. The most powerful argument for sulfene intermediates in these reactions is formation of mono- but no polydeuterated products in deuterio alcohols,¹⁶⁰⁻¹⁶³ in the absence of base, esterification of phenylmethanesulfonyl chloride proceeds slowly with no H-D exchange. Nucleophilic substitution at sulfur would yield undeuterated product, and, if a non-obligatory carbanion were formed, polydeuteration of the α position would be expected. The extent of exchange is strongly dependent on the nature of the exchanging carbon center (2-octanesulfonyl chloride exchanges less readily¹⁶⁶ than ethanesulfonyl chloride¹⁶⁰⁻¹⁶¹), the leaving group (2-octanesulfonyl azide shows no reaction under the conditions used for the chloride),¹⁶⁶ and the structure of the catalytic base.^{157,167} Thus, less bulky bases lead to monoexchange, but multiexchange becomes more pronounced as amine base size increases.¹⁵⁷

It is believed that E-A substitution with an E-2 step to sulfene occurs in the solvolyses of sulfonyl halides^{159,168} in view of the base size effect, variation of k_D/k_H with base, the dependence of the rates of pyridine-catalyzed chloride expulsion from substituted phenylmethanesulfonyl chlorides on σ^- Hammett constants ($\rho^- = 2.35$) and the observation that **55** reacts more slowly by a factor of 15 than **56** in the absence of base but more rapidly by a factor of 71 in the presence of triethylamine.¹⁵⁹ King¹⁵⁹ excludes the E1cB mechanism (as opposed to E2) since the rates of exchange and hydrolysis are incompatible on such a basis. Considering the discussions now taking place concerning the validity of the concerted elimination reaction in general,⁵ an E1cB mechanism for the



above solvolyses could adequately account for the experimental observations if the proton transfer step were essentially rate controlling. An E1cB mechanism has been proposed, however, for the hydrolysis of dichloromethanesulfonyl chloride since k_{exchange} is greater than $k_{\text{hydrolysis}}$.¹⁶⁹ Although distinction between E2, E1cB, or rate-controlling proton transfer in E1cB is not made, an E-A mechanism has been postulated to account for the higher base lability of α hydrogen containing fluoroalkanesulfonyl fluorides than their substituted analogs ($\text{CF}_3\text{CHFSO}_2\text{F}$ and $\text{CF}_3\text{CFCISO}_2\text{F}$).¹⁷⁰

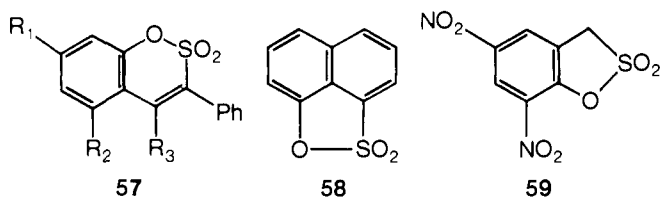
Vizgert has reported studies on aryl esters of substituted sulfonic acids, $\text{RSO}_2\text{OC}_6\text{H}_4\text{X}$,¹⁷¹ and ρ values for hydroxide hydrolysis are (at 50° in 70% v/v dioxane water): 2.53, 2.48, 2.56, and 2.46 for $\text{R} = \text{CH}_2=\text{CHCH}_2-$, $\text{CH}_2=\text{CMeCH}_2-$, CH_3 , and $n\text{-Pr}$, respectively. Since these values are close to those for aryl benzenesulfonates,¹⁴⁴ the E-A mechanism should remain hypothetical for these compounds although monodeuterated compounds are formed. Although the authors claimed a correlation with Hammett σ^- values, we could find no evidence of the use of substituents for which σ and σ^- values would differ significantly. Recent independent work from the laboratory of King¹⁷² and from this laboratory¹⁷³ has clarified the mechanism for aryl phenylmethanesulfonyl transfer reactions. The most convincing evidence for an E1cB mechanism involving an intermediate is the nonlinear dependence of rate on buffer concentration (Figure 17). At high buffer concentration the absence of a concentration effect is consistent with rate-controlling decomposition of the carbanion to sulfene, and the buffer dependence at low concentration corresponds to rate-controlling proton transfer (eq 44). Further evidence consistent with the E1cB mechanism



comes from the 1000-fold larger rate constant for hydroxide hydrolysis of the 4-nitrophenyl ester compared with the corresponding benzenesulfonate, trapping experiments with amines, a large primary deuterium isotope effect in the low buffer concentration region and a low effect in the high buffer concentration region,¹⁷⁴ and the high selectivity to Hammett substituents^{172,173} which are σ^- .¹⁷³ Further work from this laboratory¹⁷⁴ shows that as the leaving group ability of the phenol is increased a change in rate-determining step from the E1 reaction to the proton transfer occurs.

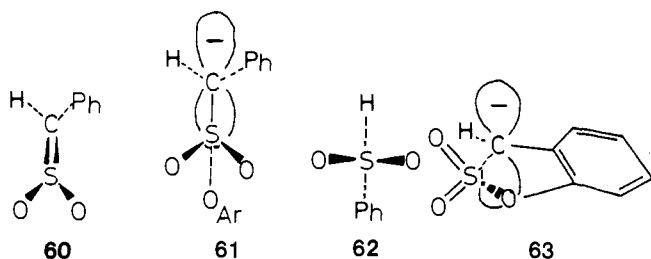
Although the chemistry of five- and six-membered sultones has been reviewed,¹⁴⁶ it is only recently that extensive kinetic studies have been undertaken.¹⁵¹ Alicyclic sultones generally react with nucleophiles via C-O fission,^{175,176} however, **53** ($\text{X} = \text{H}$) hydrolyzes in base with only S-O cleavage.¹⁷⁷ Pathways leading to S-O cleavage could involve either attack at sulfur in an A-E mechanism or a sulfene route with prior elimination of the leaving group. The latter possibility is made more real by the low reactivity of sultones of type **57** toward base cleavage,¹⁷⁸ the similarity of the $\text{p}K_a$ of **53** ($\text{X} = \text{H}$) to those

expected of carbamate esters ($pK_a > 14$) which follow an E1cB path, and the fact that the acyclic analog of **53** has a bona fide E1cB hydrolytic pathway. On the other hand, the large rate increase (10^5 – 10^6 -fold) shown by five-membered ring sultones relative to their acyclic analogs¹⁵³ need not be due to the incursion of an E1cB route since **58** shows a similar rate acceleration,¹⁷⁹ and comparable ring size effects are observed for cyclic sulfites¹⁸⁰ and sulfates.¹⁸¹ Moreover,

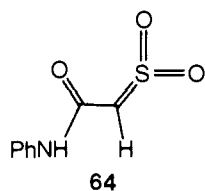


X-ray crystallographic studies have indicated considerable ring strain in five-membered sultones.¹⁸² Definitive evidence against the E1cB mechanism for the basic hydrolysis of esters of the type **53** was obtained in Kaiser's laboratory: the hydrolysis of **59** in deuterated base yields greater than 50% of the unlabeled sulfonic acid on recovery from the product mixture.¹⁸³ Further evidence was the reisolation of the calculated amount of unlabeled sultone (**53**, X = H) on hydrolysis (partial) in deuterated base.¹⁸³ The ρ value of +1.23 for a series of sultones (**53**)¹⁸⁴ is also very low for an E1cB process.

It remains to account for the apparent anomaly that esters of type **53** hydrolyze in base via an A–E mechanism whereas the acyclic analogs (eq 44) possess an E–A mechanism. Sulfenes probably have the planar configuration (**60**) as their most stable form so it is likely that the transition state for the E1 reaction will also be planar or very nearly so for those atoms constituting the sulfene intermediate (**61**). This is supported by recent theoretical calculations which indicate that the planar sulfene CH_2SO_2 is more stable by some 35 kcal/mol than the "perpendicular" form, the parent of **62**.¹⁸⁵ Although the benzylic proton of the sultone **53** is labile and could readily expel the phenoxide consideration of the transition state for such a mechanism, **63** shows that the incipient sulfene is in a "perpendicular" conformation and not the more stable planar form.¹⁷³



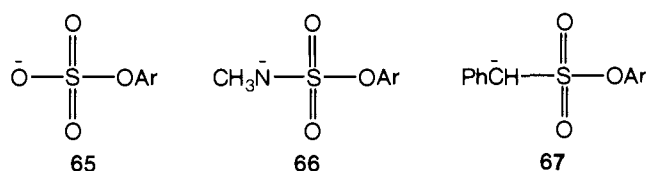
The unusual reactivity of the sulfonyl center in sulfoacetic acid derivatives has been explained by the intermediacy of a π -stabilized sulfene (**64**).¹⁸⁶ Iminosulfenes, $\text{R}_2\text{C}=\text{S}(\text{O})=\text{NX}$, have been proposed in the base-catalyzed alcoholysis of alkaneiminosulfonyl chlorides.¹⁸⁷



4. Role of the pK_a of the α Proton

We are now in a position to discuss the role of the pK_a of the α proton: in the series of anions **65**, **66**, and **67**, we are

fairly sure that weak participation by the nucleophile water is needed to expel the aryl oxide in the sulfate half ester, but this necessity only arises in the aminosulfonate case (**66**)

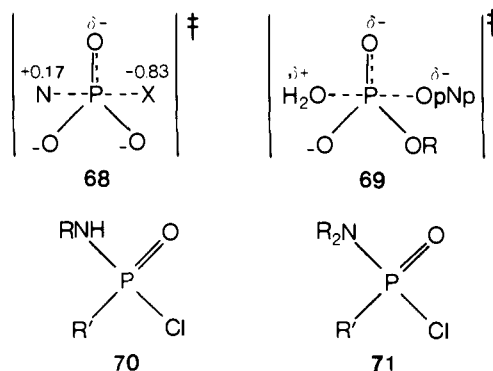


when the leaving group becomes weak, for example, with phenol.^{143b} With the carbanion no water assistance is necessary. The ratio of pK_a 's of the α protons is $65 < 66 < 67$. At high pK_a in a given substrate the concentration of anion may not be sufficient to support an E1cB process in competition with an A–E mechanism. At low pK_a , although the concentration of anion is relatively large, the driving force for expulsion of the leaving group may not be sufficient for the E1 process to compete. Since the E1 process is in a sense an internal nucleophilic displacement we shall speak of "internal nucleophilicity" when discussing the power of the anion to expel a leaving group. When the transition-state for the elimination is close to products the internal nucleophilicity is strongly dependent on the stability of the intermediate.

E. Phosphoric Acid Derivatives

1. Phosphates

The hydrolysis and transfer of monophosphate esters and derivatives in chemical and biochemical systems have been the subject of authoritative reviews,¹⁸⁸ and for this reason we only wish to mention the conclusions of these reviews and the most recent advances not incorporated. It has been concluded that extensive P–X cleavage occurs in the transition state **68** with little bond formation between nucleophile and phosphorus^{189a} although there is doubt as to the extent of bond formation in the hydrolysis of the dianion. In monoanion hydrolysis there is presumably proton transfer to the leaving group, and a stepwise process or a concerted cyclic mechanism involving water as a bridge has been postulated for this. Recent evidence involving the observation of an ^{18}O isotope effect (1.0204 ± 0.0044) in the hydrolysis of 2,4-dinitrophenyl phosphate at neutral pH's confirms the existence of a transition state (**68**) with considerable "metaphosphate" character. While the isotope effect does not rule out mechanisms which do not involve rate-determining P–O cleavage such as A–E processes, a preliminary study indicated no isotope effect in the hydrolysis of dibenzyl 2,4-dinitrophenyl phosphate.¹⁹⁰



In contrast with the dianions of monophosphates which hydrolyze via the above mechanism,^{190,191} phospho diesters with good leaving groups involve an A–E mechanism for monoanion hydrolysis¹⁹² with considerable bond formation in the transition state (**69**). The change in mechanism from E–A to

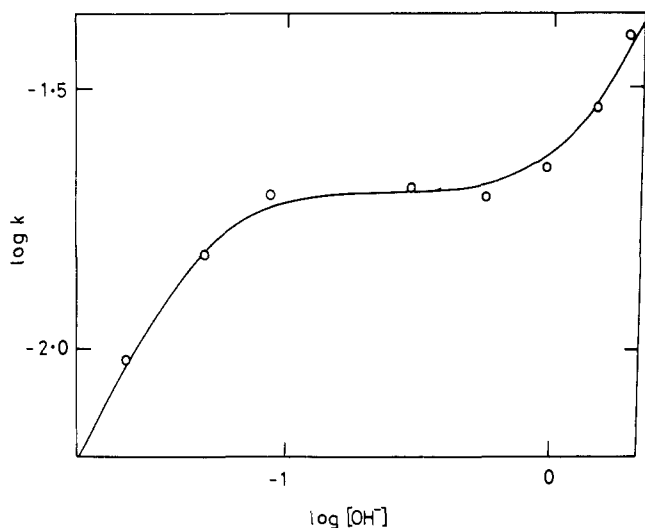


Figure 18. Dependence on hydroxide ion concentration of the rate constant for the hydrolysis of 4-nitrophenyl *N,N'*-diphenylphosphorodiamidate (58.3°, 50% ethanol-water). The figure is redrawn from Figure 5 of A. Williams and K. T. Douglas, *J. Chem. Soc., Perkin Trans. 2*, 318 (1973).

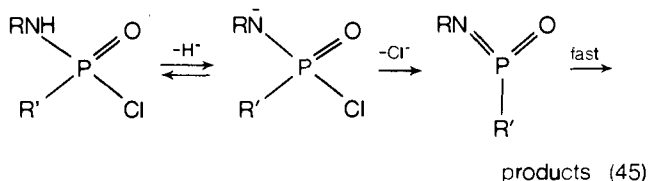
A-E as the phosphate is esterified is related to the lower "internal nucleophilicity" of the α atom in the diester case ($pK_a \approx 1$) compared with those in the monoester dianion (pK_a 's ≈ 1 and 6). A statistical factor is also involved in providing extra driving force for leaving group expulsion in the mono ester case.

2. Phosphoramidate

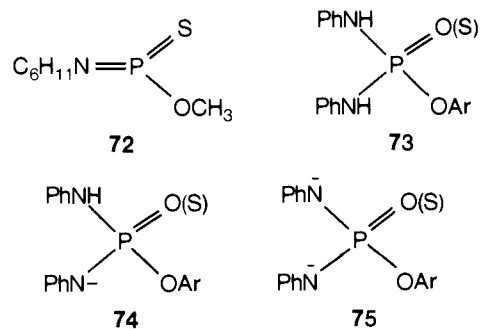
The large difference in alkaline rate constants between mono- and disubstituted nitrogen esters and halides was originally attributed to steric inhibition of the approach of hydroxide ion in an A-E mechanism. Later¹⁹³ it was attributed to hydrogen bonding of the hydroxide ion to the NH of the substrate. Westheimer¹⁹⁴ was the first to suggest the operation of an E-A mechanism of hydrolysis to account for the large rate constant differences. Further work showed that the reactions of nucleophiles other than hydroxide are not activated consistent with the proposed mechanism. Also, 2,6-lutidine catalyzes the hydrolysis of *N,N'*-diethylphosphoramidic chloride but not that of diethylphosphoryl chloride.¹⁹⁵

Hydrogen peroxide accelerates the rate of release of 4-nitrophenol from 4-nitrophenyl *N,N'*-diphenylphosphorodiamidate and the corresponding *N,N'*-dimethyl analog¹⁹⁶ in accord with an A-E mechanism for hydroperoxide attack and an E1cB mechanism for hydroxide attack. The changeover in mechanism is presumably caused by the powerful α -nucleophile effect of the hydroperoxide.

Stereochemical diagnosis of mechanism is possible with phosphoramidate hydrolyses; the alkaline hydrolysis of asymmetric methyl *N*-cyclohexylphosphorothioic chloride gives a

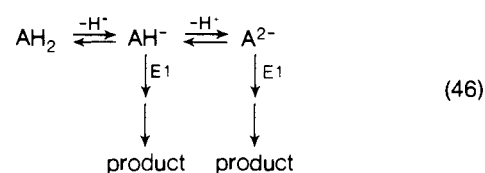


racemic product in accord with a planar intermediate, the metaphosphorimidate (72).¹⁹⁷ The products of alkaline hydrolysis of the 4-nitrophenyl ester are optically active. While the evidence for the planar intermediate is difficult to assign to any other process the evidence for the 4-nitrophenyl ester

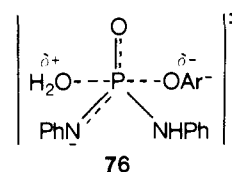


could fit either A-E or E-A mechanisms. A solvolysis study using methanol/water solvents for the chloride revealed a preference for reaction with methanol,¹⁹⁸ and it was concluded that hydrolysis did not involve free metaphosphorimidate.

Hydrolysis of substituted phenyl *N,N'*-diphenylphosphorodiamidates and the analogous thio ester revealed pH dependencies similar to Figure 18.¹⁹⁹ The observed rate law fitted a



scheme where both dianion and anion decomposed to metaphosphorimidate intermediates. The Hammett sensitivities for the apparent hydroxide rate constants are too large to accommodate a simple A-E mechanism.¹⁹⁹ None of the results so far obtained distinguish between the formation of free metaphosphorimidate and a transition state involving weak bond formation between nucleophile and phosphorus (76).



Hydrolysis of phosphoramidates with expulsion of alkyl leaving groups could proceed via an E1cB mechanism.²⁰⁰

IV. Appendix. Elimination-Addition Mechanisms of Acyl Group Transfer

Recent work has included a novel application of solid-phase techniques to the problem of intermediate trapping. Rebek and his coworkers attach the acyl group donor to a polymer molecule and the acceptor or trapping nucleophile to another polymer. If the intermediate is sufficiently stable in free solution, it migrates to the acceptor where the product is detected by normal analytical methods. The technique, called the "three-phase test", is clearly a positive test: if no transfer is observed, then the intermediate could be nonexistent or too labile to exist long enough in solution to migrate. Addition-elimination must be very rare between polymer acceptors and donors. The method has been applied to carbamate,²⁰¹ β -ketoacyl,²⁰¹ and phosphoramidate²⁰² transfer and has provided the first convincing demonstration of the elusive metaphosphate (PO_3^{2-}) intermediate in monophosphate transfer.²⁰³

Sulfene chemistry has been the subject of an important review,²⁰⁴ and an E2 mechanism for hydrolysis of phenylmethanesulfonate esters of very acidic phenols has been demonstrated to have a carbanion-like transition state involving very little S-O bond cleavage.²⁰⁵

Trapping experiments with *N*-methylaniline acceptor have been used to infer the existence of monomeric metaphosphates in the pyrolysis of methyl 2-butenylphosphonate.²⁰⁶ Metaphosphorimidates were postulated as intermediates in the photolysis in methanol of *cis*- and *trans*-1-azido-2,2,3,4,4-pentamethylphosphetane 1-oxide to explain the identity of the ratio of *cis* and *trans* methyl esters.²⁰⁷ Recently three-coordinate five-valent phosphorus compounds ($R_2NP(NR)_2$ and $R_2NPS(NR)$) have been isolated from the corresponding azides; these unsaturated intermediates have stabilizing groups such as *tert*-butyl or trimethylsilyl attached to the nitrogen.²⁰⁸

V. References and Notes

- (1) (a) M. L. Bender, *Chem. Rev.*, **60**, 53 (1960); (b) T. C. Bruice and S. J. Benkovic, *Bioorg. Chem.*, **1**, 1 (1966); (c) W. P. Jencks, "Catalysis in Chemistry and Enzymology," McGraw-Hill, New York, N.Y., 1969; (d) S. L. Johnson, *Adv. Phys. Org. Chem.*, **5**, 237 (1967).
- (2) (a) J. de Jersey, P. Willadsen, and B. Zerner, *Biochemistry*, **8**, 1959 (1969); (b) M. L. Tobe, *Acc. Chem. Res.*, **3**, 377 (1970).
- (3) (a) D. P. N. Satchell, *Q. Rev. Chem. Soc.*, **17**, 160 (1963); (b) G. A. Olah, "Friedel-Crafts and Related Reactions," Vol. 1-4, Wiley-Interscience, New York, N.Y., 1963-1964. (c) We utilize the designation A-E to indicate an addition-elimination mechanism because the phenomenological symbols SN_2 have come to mean a particular mechanistic type involving concerted displacement of a group by a nucleophile. We shall use the symbols E-A to indicate an elimination-addition process in general and $E1cB$ for a specific type involving ionization of the α proton prior to elimination. We prefer not to use terms like BAC_2 in referring to these mechanisms as we believe that they are strictly phenomenological, and in some cases the observation "base catalysis, acyl oxygen cleavage, bimolecular" (BAC_2) changes to "uncatalyzed, acyl oxygen cleavage, unimolecular" with increase in base concentration although the mechanism does not change. SN_1 retains the same mechanistic meaning as in alkyl halide hydrolysis, namely, an ionization ($RX \rightarrow R^+ + X^-$) followed by addition of nucleophile.
- (4) G. S. Hammond, *J. Am. Chem. Soc.*, **77**, 334 (1955).
- (5) F. G. Bordwell, *Acc. Chem. Res.*, **3**, 281 (1970); **5**, 374 (1972).
- (6) R. A. More O'Ferrall, *J. Chem. Soc. B*, 274 (1970).
- (7) (a) W. P. Jencks, *Chem. Rev.*, **72**, 705 (1972); *J. Am. Chem. Soc.*, **94**, 4731 (1972); (b) H. P. Treffers and L. P. Hammett, *ibid.*, **59**, 1708 (1937).
- (8) (a) M. S. Newman, H. G. Kuivila, and A. B. Garrett, *J. Am. Chem. Soc.*, **67**, 704 (1945); (b) M. S. Newman, *ibid.*, **63**, 2431 (1941); (c) M. S. Newman, R. A. Craig, and A. B. Garrett, *ibid.*, **71**, 869 (1949).
- (9) (a) C. T. Chmiel and F. A. Long, *J. Am. Chem. Soc.*, **78**, 3326 (1956); (b) M. L. Bender, H. Ladenheim, and M. C. Chen, *ibid.*, **83**, 123 (1961).
- (10) H. Ladenheim and M. L. Bender, *J. Am. Chem. Soc.*, **82**, 1895 (1960).
- (11) (a) M. L. Bender and R. S. Dewey, *J. Am. Chem. Soc.*, **78**, 317 (1956); (b) M. L. Bender and M. C. Chen, *ibid.*, **85**, 37 (1963).
- (12) B. L. Archer and R. F. Hudson, *J. Chem. Soc.*, 3259 (1950).
- (13) D. A. Brown and R. F. Hudson, *J. Chem. Soc.*, 883 (1953).
- (14) B. L. Archer, R. F. Hudson, and J. E. Wardill, *J. Chem. Soc.*, 888 (1953).
- (15) (a) D. A. Brown and R. F. Hudson, *J. Chem. Soc.*, 3352 (1953); (b) V. Gold, J. Hilton, and E. G. Jefferson, *ibid.*, 2756 (1954); (c) R. F. Hudson and E. W. Crunden, *ibid.*, 501 (1956); (d) E. Grunwald and S. Winstein, *J. Am. Chem. Soc.*, **70**, 846 (1948).
- (16) C. A. Bunton and T. A. Lewis, *Chem. Ind. (London)*, 180 (1956).
- (17) (a) C. A. Bunton, T. A. Lewis, and D. R. Llewellyn, *Chem. Ind. (London)*, 1154 (1954); (b) H. K. Hall, *J. Am. Chem. Soc.*, **77**, 5993 (1955); (c) B. P. Susz and D. Casimatis, *Helv. Chim. Acta*, **44**, 395 (1961); (d) D. Casimatis, P. Gagnaux, and B. P. Susz, *ibid.*, **43**, 424 (1960).
- (18) (a) M. L. Bender and M. C. Chen, *J. Am. Chem. Soc.*, **85**, 30 (1963); (b) G. A. Olah, D. H. O'Brien, and A. M. White, *ibid.*, **89**, 5694 (1967); (c) I. Ugi, F. Beck, and V. Fetyer, *Chem. Ber.*, **95**, 126 (1962); (d) J. S. Clovis, A. Eckell, R. Huisgen, and R. S. Sutmann, *ibid.*, **100**, 60 (1967); (e) R. N. Butler and F. L. Scott, *Chem. Ind. (London)*, 1216 (1970); (f) J. Donovan, J. Cronin, F. L. Scott, and A. F. Hegarty, *J. Chem. Soc., Perkin Trans. 2*, 1050 (1972); (g) A. F. Hegarty, M. Cashman, and F. L. Scott, *J. Chem. Soc. B*, 1607 (1971); *Chem. Commun.*, 689 (1971); (h) A. F. Hegarty, M. P. Cashman, and F. L. Scott, *J. Chem. Soc., Perkin Trans. 2*, 44 (1972); (i) A. F. Hegarty, M. P. Cashman, J. B. Aylward, and F. L. Scott, *J. Chem. Soc. B*, 1879 (1971).
- (19) (a) S. L. Johnson and K. A. Rumon, *J. Am. Chem. Soc.*, **87**, 4782 (1965); (b) S. L. Johnson and H. M. Giron, *J. Org. Chem.*, **37**, 1383 (1972); (c) R. Bacaloglu, C. Daesca, and G. Ostrogovich, *J. Chem. Soc., Perkin Trans. 2*, 1011 (1972); (d) G. A. Olah, J. Nishimura, and P. Kreienbühl, *J. Am. Chem. Soc.*, **95**, 7672 (1973).
- (20) (a) H. K. Hall, *J. Org. Chem.*, **21**, 248 (1956); (b) P. S. Traylor and F. H. Westheimer, *J. Am. Chem. Soc.*, **87**, 553 (1965).
- (21) H. K. Hall, *J. Am. Chem. Soc.*, **78**, 1450 (1956).
- (22) H. K. Hall and C. H. Lueck, *J. Org. Chem.*, **28**, 2818 (1963).
- (23) P. Haake and P. S. Ossip, *Tetrahedron Lett.*, 4841 (1970).
- (24) W. E. Handford and J. C. Sauer, *Org. React.*, **3**, 124 (1947).
- (25) H. Böhme and G. Lérche, *Justus Liebig's Ann. Chem.*, **705**, 154 (1967).
- (26) F. Klages and E. Zange, *Justus Liebig's Ann. Chem.*, **607**, 35 (1957).
- (27) W. E. Truce and P. S. Bailey, *J. Org. Chem.*, **34**, 1341 (1969).
- (28) K. C. Brannock and R. D. Burpitt, *J. Org. Chem.*, **30**, 2564 (1965).
- (29) W. Reid and P. Junker, *Justus Liebig's Ann. Chem.*, **700**, 32 (1966).
- (30) (a) C. L. Stevens and M. E. Munk, *J. Am. Chem. Soc.*, **80**, 4065 (1958); (b) C. L. Stevens and J. C. French, *ibid.*, **76**, 4398 (1954).
- (31) (a) P. Dimroth, German Patent 1,166,771 (1964); *Chem. Abstr.*, **60**, 15774b (1964); (b) R. N. Lacey in "The Chemistry of Alkenes," S. Patai, Ed., Interscience, New York, N.Y., 1964, p. 1173.
- (32) B. Holmquist and T. C. Bruice, *J. Am. Chem. Soc.*, **91**, 2982 (1969).
- (33) T. C. Bruice and B. Holmquist, *J. Am. Chem. Soc.*, **90**, 7136 (1968).
- (34) (a) B. Holmquist and T. C. Bruice, *J. Am. Chem. Soc.*, **91**, 2993 (1969); (b) H. E. Zaugg, M. Freifelder, H. J. Glenn, H. W. Horrom, G. R. Stone, and M. R. Vernsten, *ibid.*, **78**, 2626 (1956); (c) G. E. Lienhard and T.-C. Wang, *ibid.*, **90**, 3781 (1968); (d) G. L. Schmir, *ibid.*, **87**, 2743 (1965).
- (35) R. F. Pratt and T. C. Bruice, *J. Am. Chem. Soc.*, **92**, 5956 (1970).
- (36) A. J. Kirby and G. J. Lloyd, *Chem. Commun.*, 1538 (1971).
- (37) (a) H. Pracejus and R. Samtleben, *Tetrahedron Lett.*, 2189 (1970); (b) P. J. Lillford and D. P. N. Satchell, *J. Chem. Soc. B*, 897 (1968); (c) C. D. Hurd, *J. Am. Chem. Soc.*, **61**, 476 (1939).
- (38) (a) R. P. Bell, "The Proton in Chemistry," Cornell University Press, Ithaca, N.Y., 1959, p. 188; (b) P. S. Tobias and F. J. Kézdy, *J. Am. Chem. Soc.*, **91**, 5171 (1969); (c) R. B. Homer and M. L. Bender, *J. Org. Chem.*, **30**, 3975 (1965).
- (39) D. S. Campbell and G. W. Laurie, *Chem. Commun.*, 355 (1971).
- (40) (a) S. P. Rowland, L. Z. Percy, C. H. Mack, and H. J. Janssen, *J. Am. Chem. Soc. B*, 404 (1968); (b) A. R. Bader and H. A. Vogel, *J. Am. Chem. Soc.*, **74**, 3993 (1952); (c) A. R. Bader, C. O. Cummings, and H. A. Vogel, *ibid.*, **73**, 4195 (1951).
- (41) (a) A. Williams, *J. Chem. Soc., Perkin Trans. 2*, 808 (1972); (b) E. Funke and R. Huisgen, *Chem. Ber.*, **104**, 3222 (1971).
- (42) R. A. Abeles, personal communication, 1973.
- (43) (a) E. Kuehle and R. Wegler, *Justus Liebig's Ann. Chem.*, **616**, 183 (1958); (b) J. C. Sheehan, P. A. Cruickshank, and G. L. Boshart, *J. Org. Chem.*, **26**, 2525 (1961).
- (44) (a) W. P. Jencks and J. Carriuolo, *J. Am. Chem. Soc.*, **82**, 675 (1960); (b) F. A. Long and J. Bigeleisen, *Trans. Faraday Soc.*, **55**, 2077 (1959).
- (45) (a) J. A. Schmitt and F. Daniels, *J. Am. Chem. Soc.*, **75**, 3564 (1953); (b) P. E. Yankwich and A. E. Veazie, *ibid.*, **80**, 1833 (1958); (c) A. C. Satterthwait and W. P. Jencks, *J. Am. Chem. Soc.*, **96**, 7018, 7031, 7045 (1974); (d) M. H. O'Leary and M. D. Kluetz, *ibid.*, **92**, 6089 (1970); (e) M. H. O'Leary, *ibid.*, **91**, 6886 (1969); (f) M. H. O'Leary and D. W. Hendrickson, *Fed. Proc., Fed. Am. Soc. Exp. Biol.*, **29**, 407 (1970); (g) M. H. O'Leary, D. T. Richards, and D. W. Hendrickson, *J. Am. Chem. Soc.*, **92**, 4435 (1970).
- (46) H. Krall, *J. Chem. Soc.*, 1396 (1915).
- (47) J. Bell, *J. Chem. Soc.*, 1216 (1926).
- (48) E. A. Werner, *J. Chem. Soc.*, 1093 (1919).
- (49) S. Hecht and J. W. Kozarich, *Tetrahedron Lett.*, 5147 (1972).
- (50) W. R. G. Atkins and E. A. Werner, *J. Chem. Soc.*, 1167 (1912).
- (51) A. Bernthsen and H. Klinger, *Ber.*, **11**, 492 (1878).
- (52) A. F. Hegarty, C. N. Hegarty, and F. L. Scott, *J. Chem. Soc., Perkin Trans. 2*, 2054 (1973).
- (53) G. R. Stark, *Biochemistry*, **4**, 588 (1965).
- (54) (a) J. E. Hodgkins and M. G. Ettinger, *J. Org. Chem.*, **21**, 404 (1956); (b) J. E. Hodgkins, W. P. Reeves, and Y.-T. G. Liu, *J. Am. Chem. Soc.*, **83**, 2532 (1961).
- (55) (a) L. A. Chugaev, *J. Russ. Phys.-Chem. Soc.*, **35**, 1116 (1903); *Chem. Ber.*, **35**, 2473 (1902); (b) O. Hermes, *J. Prakt. Chem.*, **97**, 465 (1866).
- (56) (a) E. Lieber and G. B. L. Smith, *Chem. Rev.*, **25**, 213 (1939); (b) J. Thiele, *Justus Liebig's Ann. Chem.*, **270**, 1 (1892); (c) E. Lieber and G. B. L. Smith, *J. Am. Chem. Soc.*, **59**, 2283 (1937); (d) J. Thiele, *Justus Liebig's Ann. Chem.*, **270**, 1 (1892); (e) D. D. Van Slyke, *J. Biol. Chem.*, **10**, 25 (1911-1912); (f) R. H. A. Plimmer, *Biochem. J.*, **10**, 115 (1916); (g) A. E. Koehler, *J. Biol. Chem.*, **42**, 267 (1920); (h) T. L. Davis and K. C. Blanchard, *J. Am. Chem. Soc.*, **51**, 1794 (1929); *Proc. Natl. Acad. Sci. U.S.A.*, **11**, 71 (1925); (i) R. Willstätter and A. Pfannenstiel, *Chem. Ber.*, **59**, 1870 (1926).
- (57) (a) J. Thiele and A. Lachmann, *Justus Liebig's Ann. Chem.*, **288**, 267 (1895); (b) E. A. Werner, *J. Chem. Soc.*, 622 (1918); (c) E. Melder, *Recl. Trav. Chim. Pays-Bas*, **6**, 169 (1888).
- (58) (a) E. A. Werner, *J. Chem. Soc.*, 84 (1918); (b) A. Scattergood, *Inorg. Synth.*, **2**, 87 (1946); (c) T. Walker and F. J. Hambly, *J. Chem. Soc.*, 746 (1895); (d) J. Walker and S. A. Kay, *ibid.*, 489 (1897); (e) J. Walker and J. K. Wood, *ibid.*, 21 (1900).
- (59) W. H. R. Shaw and T. J. Bordeaux, *J. Am. Chem. Soc.*, **77**, 4729 (1955).
- (60) A. Williams, *J. Chem. Soc.*, 1244 (1973).
- (61) A. F. Hegarty and L. N. Frost, *J. Chem. Soc.*, 1719 (1973); *J. Chem. Soc., Chem. Commun.*, 500 (1972).
- (62) Results of L. W. Dittert, "The Kinetics and Mechanism of Base-Catalyzed Hydrolysis of Organic Carbamates," Ph.D. Dissertation, University of Wisconsin, 1961; *Diss. Abstr.*, **22**, 1837 (1961). Discussed in ref 38c.
- (63) L. W. Dittert and T. Higuchi, *J. Pharm. Sci.*, **52**, 852 (1963).
- (64) (a) A. Williams and R. A. Naylor, *J. Chem. Soc. B*, 1967 (1971); (b) J. Ryan and A. A. Humffray, *ibid.*, 842 (1966); (c) T. C. Bruice and B. F. Mayahi, *J. Am. Chem. Soc.*, **82**, 3067 (1960); (d) E. Tommila and C. N. Hinshelwood, *J. Chem. Soc.*, 1801 (1938).
- (65) I. Christenson, *Acta Chem. Scand.*, **18**, 904 (1964).
- (66) T. Vontor and M. Vecera, *Collect. Czech. Chem. Commun.*, **38**, 516 (1973).
- (67) T. Vontor, J. Socha, and M. Vecera, *Collect. Czech. Chem. Commun.*, **37**, 2183 (1972).
- (68) J. E. Casida, K.-B. Augustinsson, and G. Jonsson, *J. Econ. Entomol.*, **53**, 205 (1960).
- (69) A. F. Hegarty, C. N. Hegarty, and F. L. Scott, *J. Chem. Soc., Perkin Trans. 2*, 1258 (1974).
- (70) (a) M. Eigen, *Angew. Chem., Int. Ed. Engl.*, **1**, 1 (1964); (b) A. Farkas

- and G. A. Mills, *Adv. Catal.*, **13**, 393 (1962); (c) J. M. Briody and D. Narinesingh, *Tetrahedron Lett.*, 4143 (1971); (d) S. A. Lammiman and R. S. Satchell, *J. Chem. Soc., Perkin Trans. 2*, 877 (1974); (e) R. Balcoglu and C. A. Bunton, *Tetrahedron*, **29**, 2721 (1973).
- (71) (a) A. Williams and W. P. Jencks, *J. Chem. Soc., Perkin Trans. 2*, 1753 (1974); (b) *ibid.*, 1760 (1974).
- (72) (a) C. M. Allen and M. E. Jones, *Biochemistry*, **3**, 1238 (1964); (b) M. Halmann, A. Lapidot, and D. Samuel, *J. Chem. Soc.*, 1944 (1962); (c) C. M. Allen and J. Jamieson, *J. Am. Chem. Soc.*, **93**, 1434 (1971); (d) S. Motola, Ph.D. Thesis, University of Wisconsin, 1968; *Diss. Abstr. B*, **29**, 559 (1968).
- (73) A. Williams, unpublished results.
- (74) J. M. Lowenstein, *J. Chem. Soc.*, 4667 (1956).
- (75) S. Grisolia and D. P. Wallach, *Biochim. Biophys. Acta*, **18**, 449 (1955).
- (76) (a) R. C. Valentine and R. S. Wolfe, *Biochem. Biophys. Res. Commun.*, **2**, 384 (1960); *Biochim. Biophys. Acta*, **45**, 389 (1960); (b) A. G. Lezius, E. Ringelman, and F. Lynen, *Biochem. Z.*, **336**, 510 (1963).
- (77) O. Roshovansky and S. Ratner, *J. Biol. Chem.*, **242**, 3839 (1967); **236**, 2254 (1961).
- (78) (a) R. Abrams and M. Bentley, *Arch. Biochem. Biophys.*, 7991 (1959); (b) U. Lagerkvist, *J. Biol. Chem.*, **233**, 143 (1958).
- (79) G. R. Stark, *J. Biol. Chem.*, **246**, 3064 (1971).
- (80) P. Reichard and G. Hanshoff, *Acta Chem. Scand.*, **10**, 548 (1956).
- (81) G. R. Stark, *Biochemistry*, **4**, 588 (1965).
- (82) D. E. Applequist and D. E. McGreer, *J. Am. Chem. Soc.*, **82**, 1965 (1960).
- (83) (a) W. M. Jones, D. L. Muck, and T. K. Tonday, *J. Am. Chem. Soc.*, **88**, 68 (1966); (b) W. M. Jones and D. L. Muck, *ibid.*, **88**, 3798 (1966); (c) S. Hecht and J. W. Kazarich, *J. Org. Chem.*, **38**, 1821 (1973); (d) A. F. McKay and G. F. Wright, *J. Am. Chem. Soc.*, **69**, 3028 (1947).
- (84) (a) K. Kawasaki, R. Camble, G. Dupuis, H. Romovacek, H. T. Storey, C. Yanaihara, and K. Hofmann, *J. Am. Chem. Soc.*, **95**, 6815 (1973); (b) H. Tschesche and H. Jering, *Angew. Chem., Int. Ed. Engl.*, **12**, 756 (1973); (c) H. O. Bouveng, *Acta Chem. Scand.*, **15**, 87 (1961); (d) P. J. Garegg, *ibid.*, **16**, 1849 (1962); (e) D. G. Smyth, *Acta Chim. Hung.*, **44**, 197 (1965).
- (85) (a) R. P. Steiger and E. B. Miller, *J. Org. Chem.*, **24**, 1214 (1959); (b) J. F. Bunnett and M. B. Naff, *J. Am. Chem. Soc.*, **88**, 4001 (1966).
- (86) (a) K. Koppie, *J. Am. Chem. Soc.*, **79**, 6442 (1957); (b) P. D. Bartlett and R. H. Jones, *ibid.*, **79**, 2153, 2159 (1957).
- (87) (a) J. D. Billimora and A. H. Cook, *J. Chem. Soc.*, 2323 (1949); (b) A. C. Davis and A. L. Levy, *ibid.*, 2419 (1951); (c) A. H. Cook and A. L. Levy, *ibid.*, 651 (1950); (d) R. Hirschmann, R. G. Strachan, H. Schwam, E. F. Schoenewaldt, H. Joshua, H. Barkemeyer, D. F. Veber, W. J. Paleveda, Jr., T. A. Jacob, T. E. Beesley, and R. G. Denkwalter, *J. Org. Chem.*, **32**, 3415 (1967); (e) J. P. Greenstein and M. Winitz, "Chemistry of the Amino Acids," Wiley, New York, N.Y., 1961, pp 860, 876; (f) R. Hirschmann, H. Schwam, R. G. Strachan, E. F. Schoenewaldt, H. Barkemeyer, S. M. Miller, J. B. Conn, V. Garskey, D. F. Verber, and R. G. Denkwalter, *J. Am. Chem. Soc.*, **93**, 2746 (1971); (g) M. Wilchek, S. Ariely, and A. Patchornik, *J. Org. Chem.*, **33**, 1258 (1968).
- (88) (a) A. F. Hegarty and T. C. Bruice, *J. Am. Chem. Soc.*, **92**, 6561, 6568, 6575 (1970); (b) A. F. Hegarty, R. F. Pratt, T. Giudici, and T. C. Bruice, *ibid.*, **93**, 1428 (1971).
- (89) (a) L. N. Frost and A. F. Hegarty, *J. Chem. Soc., Chem. Commun.*, 82 (1973); (b) A. F. Hegarty, L. N. Frost, and D. Cremin, *J. Chem. Soc., Perkin Trans. 2*, 1249 (1974).
- (90) G. B. Barlin and D. D. Perrin, *Q. Rev. Chem. Soc.*, 2075 (1966).
- (91) J. E. C. Hutchins and T. H. Fife, *J. Am. Chem. Soc.*, **95**, 2282 (1973).
- (92) J. E. C. Hutchins and T. H. Fife, *J. Am. Chem. Soc.*, **95**, 3786 (1973).
- (93) (a) R. S. P. Hsi, L. W. Brown, F. Kagan, A. R. Hanze, J. E. Stafford, and A. A. Forist, *J. Org. Chem.*, **37**, 3427 (1972); (b) A. F. Hegarty, L. N. Frost, and T. H. Coy, *ibid.*, **39**, 1089 (1974).
- (94) A. W. Levine and J. Fech, *J. Org. Chem.*, **37**, 1500, 2455 (1972).
- (95) (a) G. G. Smith and F. W. Kelly, *Prog. Phys. Org. Chem.*, **8**, 146 (1971); (b) L. W. Clark in "The Chemistry of Carboxylic Acids and Esters," S. Patai, Ed., Interscience, New York, N.Y., 1969, p 589.
- (96) S. L. Johnson and D. L. Morrison, *J. Am. Chem. Soc.*, **94**, 1323 (1972).
- (97) M. Caplow, *J. Am. Chem. Soc.*, **90**, 6795 (1968); **87**, 574 (1965).
- (98) M. Caplow and M. Yager, *J. Am. Chem. Soc.*, **89**, 4513 (1967).
- (99) (a) S. J. Joris, K. I. Aspila, and C. L. Chakrabarti, *Anal. Chem.*, **41**, 1441 (1969); (b) K. I. Aspila, S. J. Joris, and C. L. Chakrabarti, *J. Phys. Chem.*, **74**, 3625 (1970); (c) I. M. Bhatt, K. P. Soni, and A. M. Trivedi, *J. Indian Chem. Soc.*, **45**, 354 (1968); (d) A. Hulanicki, *Talanta*, **14**, 1371 (1967); (e) K. I. Aspila, U. S. Sastry, and C. L. Chakrabarti, *ibid.*, **16**, 1099 (1969); (f) D. J. Halls, *Mikrochim. Acta*, 62 (1969); (g) H. Bode, *Fresenius' Z. Anal. Chem.*, **142**, 414 (1954).
- (100) R. Zahradnik and P. Zuman, *Collect. Czech. Chem. Commun.*, **24**, 1132 (1959).
- (101) (a) D. M. Miller and R. A. Latimer, *Can. J. Chem.*, **40**, 246 (1962); (b) H. Bode and F. Newman, *Z. Anal. Chem.*, **169**, 410 (1959); (c) G. D. Thorn and R. A. Ludwig, "The Dithiocarbamates and Related Compounds," Elsevier, Amsterdam, 1962.
- (102) (a) A. Jensen, E. Jorgensen, and C. Faurholt, *Acta Chem. Scand.*, **6**, 1036 (1952); (b) I. Noring, A. Jensen, and C. Faurholt, *ibid.*, **6**, 404 (1952); (c) C. Faurholt, *Z. Phys. Chem. (Leipzig)*, **126**, 72, 85, 211, 227 (1927); (d) C. Faurholt and I. C. Jespersen, *ibid.*, **165**, 79 (1933); (e) H. Soling and C. Faurholt, *Dan. Tidsskr. Farm.*, **25**, 89 (1951); (f) C. Faurholt and J. C. Gjaldbaek, *ibid.*, **17**, 213 (1943); (g) E. Jorgensen and C. Faurholt, *Acta Chem. Scand.*, **8**, 1141 (1954); (h) M. B. Hansen and C. Faurholt, *ibid.*, **12**, 797 (1958); (i) V. Lund, J. C. Gjaldbaek, and C. Faurholt, *Dan. Tidsskr. Farm.*, **21**, 243 (1947); (j) C. Faurholt and J. C. Gjaldbaek, *ibid.*, **19**, 255 (1945); (k) C. Faurholt and K. P. Hansen, *ibid.*, **16**, 73 (1942); (l) C. Faurholt, K. Noiesen and F. Rath, *ibid.*, **11**, 267 (1937).
- (103) (a) D. M. Kern, *J. Chem. Educ.*, **37**, 14 (1960); (b) I. Iwasaki and G. R. B. Cooke, *J. Am. Chem. Soc.*, **80**, 285 (1958); *J. Phys. Chem.*, **68**, 2031 (1964); (c) E. Klein, J. K. Bosarge, and I. Norman, *ibid.*, **64**, 1666 (1960).
- (104) (a) J. L. Kice, J. M. Anderson, and N. E. Pawlewski, *J. Am. Chem. Soc.*, **88**, 5245 (1966); (b) J. L. Kice, *J. Org. Chem.*, **28**, 957 (1963).
- (105) W. A. Prior, *J. Am. Chem. Soc.*, **82**, 4794 (1960).
- (106) (a) J. P. Candler and R. G. Wilkins, *J. Am. Chem. Soc.*, **87**, 1490 (1965); (b) P. A. S. Smith, H. R. Alul, and R. L. Baumgarten, *ibid.*, **86**, 1139 (1964); (c) R. T. M. Fraser, *J. Chem. Soc.*, 1747 (1965).
- (107) A. J. Bottini and J. D. Roberts, *J. Am. Chem. Soc.*, **79**, 1462 (1957).
- (108) (a) R. L. Burwell, *J. Am. Chem. Soc.*, **74**, 1462 (1952); (b) S. Burstein and S. Lieberman, *ibid.*, **80**, 5235 (1958); (c) B. D. Batts, *J. Chem. Soc. B*, 551 (1966); (d) E. T. Kaiser, M. Panar, and F. H. Westheimer, *J. Am. Chem. Soc.*, **85**, 602 (1963); (e) S. J. Benkovic and P. A. Benkovic, *ibid.*, **88**, 5505 (1966).
- (109) E. V. Gilbert, "Sulfonylation and Related Reactions," Wiley, New York, N.Y., 1965.
- (110) (a) J. P. Joseph, J. P. Dusza, and S. Bernstein, *Steroids*, **7**, 577 (1966); (b) M. Higuchi, M. Takahashi, T. Suzuki, and Y. Sahashi, *J. Vitaminol.*, **11**, 261 (1965).
- (111) (a) K. S. Dodgson, B. Spencer, and K. Williams, *Biochem. J.*, **64**, 216 (1956); (b) J. D. Gregory and P. W. Robbins, *Annu. Rev. Biochem.*, **29**, 347 (1960); (c) F. Lipmann, *Science*, **128**, 575 (1958); (d) S. Black, *Annu. Rev. Biochem.*, **32**, 399 (1963); (e) R. K. Banerjee and A. B. Roy, *Biochem. Biophys. Acta*, **151**, 573 (1968); (f) A. B. Roy, *Adv. Enzymol.*, **22**, 205 (1960); (g) A. B. Roy, "The Enzymes," Vol. 5, P. D. Boyer, Ed., Academic Press, New York, N.Y., 1971, p 1.
- (112) H. Hilz and M. Kittler, *Biochem. Biophys. Res. Commun.*, **3**, 140 (1960).
- (113) P. A. Katzman, R. F. Straw, H. Buckler, and E. A. Doisy, *Recent Prog. Horm. Res.*, **9**, 45 (1954); (b) G. A. Grant and D. Beall, *ibid.*, **5**, 307 (1950); (c) A. B. Roy, *Biochem. J.*, **62**, 41 (1956).
- (114) J. R. Turvey, *Adv. Carbohydr. Chem.*, **20**, 183 (1965).
- (115) E. J. Fendler and J. H. Fendler, *J. Org. Chem.*, **33**, 3852 (1968).
- (116) S. J. Benkovic in "Comprehensive Chemical Kinetics," C. H. Bamford and C. F. H. Tipper, Ed., Elsevier, Amsterdam, 1972, Chapter 10.
- (117) (a) J. H. Kreuger and W. A. Johnson, *Inorg. Chem.*, **7**, 679 (1968); (b) M. D. Bentley, S. E. Bowie, and R. D. Limoges, *J. Phys. Chem.*, **75**, 1763 (1971); (c) B. E. Fleischfresser and I. Lauder, *Aust. J. Chem.*, **15**, 251 (1962); (d) I. G. Ryss and L. P. Bogdanova, *Zh. Neorg. Khim.*, **8**, 24 (1963); (e) I. G. Ryss, L. P. Bogdanova, S. L. Idels, and T. N. Kotlyar, *ibid.*, **14**, 2991 (1969); (f) I. G. Ryss, L. P. Bogdanova, S. L. Idels, and T. N. Kotlyar, *Kinet. Katal.*, **11**, 1057 (1970); (g) I. G. Ryss and T. N. Kotlyar, *ibid.*, **11**, 244 (1970).
- (118) (a) M. M. Jones and W. L. Lockhart, *J. Inorg. Nucl. Chem.*, **30**, 1237 (1968); (b) E. Buncel, *Mech. React. Sulfur Compd.*, **5**, 71 (1970).
- (119) Reference 116, p 48.
- (120) T. Kurusu, W. Tagaki, and S. Oae, *Bull. Chem. Soc. Jpn.*, **43**, 1553 (1970).
- (121) (a) G. N. Burkhardt, W. G. K. Ford, and E. Singleton, *J. Chem. Soc.*, 17 (1936); (b) G. N. Burkhardt, C. Horrex, and D. I. Jenkins, *ibid.*, 1649, 1654 (1936).
- (122) J. L. Kice and J. M. Anderson, *J. Am. Chem. Soc.*, **88**, 5242 (1966).
- (123) (a) R. W. Hay and J. A. G. Edmonds, *Chem. Commun.*, 969 (1967); (b) R. W. Hay, C. R. Clark, and J. A. G. Edmonds, *J. Chem. Soc., Dalton Trans.*, 9 (1974).
- (124) G. N. Burkhardt, A. G. Evans, and E. Warhurst, *J. Chem. Soc.*, 25 (1936).
- (125) S. J. Benkovic, *J. Am. Chem. Soc.*, **88**, 5511 (1966).
- (126) (a) L. P. Biceilli in "Sulfamic Acid and Its Electrometallurgical Properties," Symposium Proceedings, Milan, 1966; (b) R. B. Fischer and P. Haffley, *Proc. Indiana Acad. Sci.*, **72**, 123 (1962); (c) W. J. Spillane and F. L. Scott, *Mech. React. Sulfur Compd.*, **5**, 59 (1970).
- (127) C. A. Bunton and B. N. Hendy, *J. Chem. Soc.*, 3130 (1963).
- (128) Unpublished observations quoted in ref 130.
- (129) S. J. Benkovic and P. A. Benkovic, *J. Am. Chem. Soc.*, **90**, 2646 (1968).
- (130) S. J. Benkovic and L. K. Dunikowski, *Biochemistry*, **9**, 1390 (1970).
- (131) E. J. Fendler, R. R. Liechti, and J. H. Fendler, *J. Org. Chem.*, **35**, 1658 (1970).
- (132) W. I. Congdon and M. L. Bender, *Bioorg. Chem.*, **1**, 424 (1971).
- (133) (a) C. M. Suter, P. B. Evans, and J. M. Kiefer, *J. Am. Chem. Soc.*, **60**, 538 (1938); (b) C. M. Suter, "The Organic Chemistry of Sulfur," Wiley, New York, N.Y., 1944, pp 1-94.
- (134) (a) J. D. Gregory and F. Lipmann, *J. Biol. Chem.*, **229**, 1081 (1957); (b) E. Meezon and E. A. Davidson, *ibid.*, **242**, 1685 (1967); (c) A. S. Balasubramanian and B. K. Bachhawat, *Indian J. Exp. Biol.*, **1**, 179 (1963).
- (135) (a) S. J. Benkovic and R. Hevey, *J. Am. Chem. Soc.*, **92**, 4971 (1970); (b) W. Tagaki, T. Eiki, and I. Tanaka, *Bull. Chem. Soc. Jpn.*, **44**, 1139 (1971).
- (136) S. J. Benkovic, E. Vergara, and R. Hevey, *J. Biol. Chem.*, **246**, 4926 (1971).
- (137) T. C. Stadtman, *Science*, **183**, 915 (1974).
- (138) G. M. Atkins and E. M. Burgess, *J. Am. Chem. Soc.*, **94**, 6135 (1972).
- (139) G. M. Atkins and E. M. Burgess, *J. Am. Chem. Soc.*, **90**, 4744 (1968).
- (140) R. Appel and R. Heiwerth, *Angew. Chem., Int. Ed. Engl.*, **6**, 952 (1967).
- (141) R. Gray, *Chem. Ber.*, **92**, 509 (1959).
- (142) W. L. Matier, W. T. Comer, and D. Deitchman, *J. Med. Chem.*, **15**, 538 (1972).
- (143) (a) K. T. Douglas and A. Williams, *J. Chem. Soc., Chem. Commun.*, 356 (1973); (b) A. Williams and K. T. Douglas, *J. Chem. Soc., Perkin Trans. 2*, 1727 (1974).
- (144) R. V. Vizgert, *Usp. Khim.*, **32**, 1 (1963).
- (145) I. L. Knunyants and G. A. Sokolskii, *Angew. Chem., Int. Ed. Engl.*, **11**, 588 (1972).

- (146) (a) A. Mustafa, *Chem. Rev.*, **54**, 195 (1954); (b) D. S. Breslow and H. Skolnik, "Multisulfur and Sulfur and Oxygen Five- and Six-Membered Heterocycles," Interscience, New York, N.Y., 1966, Vol. 1, p 76; Vol. 2, p 774.
- (147) G. Opitz, *Angew. Chem., Int. Ed. Engl.*, **6**, 107 (1967).
- (148) (a) E. Tommila and M. Lindholm, *Acta Chem. Scand.*, **5**, 647 (1951); (b) C. A. Bunton and Y. F. Frei, *J. Chem. Soc.*, 1872 (1951).
- (149) (a) D. E. Fahrney and A. M. Gold, *J. Am. Chem. Soc.*, **85**, 997 (1963); (b) A. M. Gold and D. E. Fahrney, *Biochemistry*, **3**, 783 (1964); **4**, 897 (1965); **5**, 2911 (1966).
- (150) E. T. Kaiser, *Intra-Sci. Chem. Rep.*, **6**, 5 (1972).
- (151) E. T. Kaiser, *Acc. Chem. Res.*, **3**, 145 (1970).
- (152) F. J. Kézdy and E. T. Kaiser, *Methods Enzymol.*, **19**, 3 (1970).
- (153) O. R. Zaborosky and E. T. Kaiser, *J. Am. Chem. Soc.*, **88**, 3084 (1966).
- (154) W. E. Truce and A. R. Naik, *Can. J. Chem.*, **44**, 297 (1966).
- (155) G. Fodor quoted in ref 147.
- (156) R. B. Woodward, *Angew. Chem.*, **78**, 557 (1966).
- (157) J. F. King, E. A. Luinstra, and D. R. K. Harding, *J. Chem. Soc., Chem. Commun.*, 1313 (1972).
- (158) P. Müller, D. F. Mayers, O. R. Zaborosky, and E. T. Kaiser, *J. Am. Chem. Soc.*, **91**, 6732 (1969).
- (159) J. F. King and T. W. S. Lee, *J. Am. Chem. Soc.*, **91**, 6524 (1969).
- (160) J. F. King and T. Durst, *J. Am. Chem. Soc.*, **86**, 287 (1964).
- (161) J. F. King and T. Durst, *J. Am. Chem. Soc.*, **87**, 5684 (1965).
- (162) W. E. Truce, R. W. Campbell, and J. R. Norell, *J. Am. Chem. Soc.*, **86**, 288 (1964).
- (163) W. E. Truce and R. W. Campbell, *J. Am. Chem. Soc.*, **88**, 3599 (1966).
- (164) P. Müller and E. T. Kaiser, personal communication, 1973.
- (165) E. T. Kaiser and D. F. Mayers, personal communication, 1973.
- (166) D. J. Cram, R. D. Trepka, and P. S. Janiak, *J. Am. Chem. Soc.*, **88**, 2749 (1966).
- (167) W. E. Truce and J. R. Norell, *J. Am. Chem. Soc.*, **85**, 3231 (1963).
- (168) J. F. King and T. W. S. Lee, *Can. J. Chem.*, **49**, 3724 (1971).
- (169) R. Seifert, M. Zbirovsky, and M. Sauer, *Collect. Czech. Chem. Commun.*, **38**, 2477 (1973).
- (170) G. A. Sokolskii, L. J. Ragulin, and I. L. Knunyants, *Dokl. Akad. Nauk SSSR*, **196**, 124 (1971).
- (171) R. V. Vizgert and Y. G. Skrypnik, *Reakts. Sposobn. Org. Soedin.*, **32**, 413 (1972).
- (172) J. F. King and R. P. Beatson, 7th International Symposium on Sulfur Chemistry, Bangor, Wales, 1974, p B20; *Tetrahedron Lett.*, 973 (1975).
- (173) A. Williams, K. T. Douglas, and J. S. Loran, *J. Chem. Soc., Chem. Commun.*, 689 (1974).
- (174) A. Williams and K. T. Douglas, unpublished results.
- (175) (a) J. H. Helberger, J. R. Heyden, and H. Winter, *Justus Liebigs Ann. Chem.*, **586**, 147 (1954); (b) J. H. Helberger and H. Lautermann, *ibid.*, **586**, 158 (1954).
- (176) A. Mori, M. Nagayama, and H. Mandai, *Bull. Chem. Soc. Jpn.*, **44**, 1669 (1971).
- (177) O. R. Zaborosky, Ph.D. Thesis, University of Chicago, 1968.
- (178) E. M. Philbin, J. Clancey, and R. F. Timoney, *Chem. Ind. (London)*, 668 (1968).
- (179) E. T. Kaiser, K. Kudo, and O. R. Zaborosky, *J. Am. Chem. Soc.*, **89**, 1393 (1967).
- (180) J. G. Tillett, *Mech. React. Sulfur Compd.*, **4**, 129 (1969).
- (181) E. T. Kaiser, I. R. Katz, and T. F. Wulfers, *J. Am. Chem. Soc.*, **87**, 3781 (1965).
- (182) E. T. Kaiser, T. W. S. Lee, and F. P. Boer, *J. Am. Chem. Soc.*, **93**, 2351 (1971).
- (183) P. Müller, D. F. Mayers, O. R. Zaborosky, and E. T. Kaiser, *J. Am. Chem. Soc.*, **91**, 6732 (1969).
- (184) O. R. Zaborosky and E. T. Kaiser, *J. Am. Chem. Soc.*, **92**, 860 (1970).
- (185) K. N. Houk, R. W. Strozler, and J. A. Hall, *Tetrahedron Lett.*, 897 (1974).
- (186) B. E. Hoogenboom, M. S. El-Faghi, S. C. Fink, E. D. Hoganson, S. E. Lindberg, T. J. Lindell, C. J. Lima, D. J. Nelson, J. O. Olson, L. Remerfeld, and K. A. Wellington, *J. Org. Chem.*, **34**, 3414 (1969).
- (187) (a) C. R. Johnson and E. U. Jonsson, *J. Am. Chem. Soc.*, **92**, 3815 (1970); (b) K. Hovius and J. B.-F. N. Engberts, *Tetrahedron Lett.*, 181 (1972).
- (188) S. J. Benkovic, *Trans. N.Y. Acad. Sci.*, **32**, 330 (1970).
- (189) (a) S. J. Benkovic and K. J. Schray, "The Enzymes," 3rd ed. Ed., P. D. Boyer, Academic Press, New York, N.Y., 1972, p 209; (b) C. A. Bunton, *Acc. Chem. Res.*, **3**, 257 (1970); (c) J. R. Cox and O. B. Ramsay, *Chem. Rev.*, **64**, 317 (1964); (d) T. C. Bruice and S. J. Benkovic, "Bioorganic Mechanisms," Vol. 2, W. A. Benjamin, New York, N.Y., 1966, Chapters 5-7.
- (190) D. G. Gorenstein, *J. Am. Chem. Soc.*, **94**, 2523 (1972).
- (191) (a) G. DiSabato and W. P. Jencks, *J. Am. Chem. Soc.*, **83**, 4400 (1961); (b) A. J. Kirby and A. G. Varvoglis, *ibid.*, **89**, 415 (1967); **88**, 1823 (1966); (c) C. A. Bunton, E. J. Fendler, and J. H. Fendler, *ibid.*, **89**, 1221 (1967); (d) C. A. Bunton, E. J. Fendler, E. Humeres, and K.-U. Yang, *J. Org. Chem.*, **32**, 2806 (1967).
- (192) A. J. Kirby and M. Younas, *J. Chem. Soc. B*, 510 (1970).
- (193) D. F. Heath "Organo-phosphorus Poisons," Pergamon Press, Oxford, 1961, p 87.
- (194) F. H. Westheimer, *Chem. Soc., Spec. Publ.*, No. 8, 180 (1957).
- (195) E. W. Crunden and R. F. Hudson, *Chem. Ind. (London)*, 1478 (1958); *J. Chem. Soc.*, 3591 (1962).
- (196) D. B. Coult and M. Green, *J. Chem. Soc.*, 5478 (1964).
- (197) (a) A. F. Gerrard and N. K. Hamer, *J. Chem. Soc. B*, 539 (1968); (b) *ibid.*, 1122 (1967).
- (198) A. F. Gerrard and N. K. Hamer, *J. Chem. Soc. B*, 369 (1969).
- (199) (a) A. Williams and K. T. Douglas, *J. Chem. Soc., Perkin Trans. 2*, 1454 (1972); (b) *ibid.*, 318 (1973).
- (200) (a) M. A. H. Fahmy, A. Khasawinah, and T. R. Fukuto, *J. Org. Chem.*, **37**, 617 (1972); (b) I. Oney and M. Caplow, *J. Am. Chem. Soc.*, **89**, 6972 (1967).
- (201) J. Rebek, D. Brown, and S. Zimmerman, *J. Am. Chem. Soc.*, **97**, 454 (1975).
- (202) J. Rebek and F. Gaviña, *J. Am. Chem. Soc.*, **97**, 1591 (1975).
- (203) J. Rebek and F. Gaviña, *J. Am. Chem. Soc.*, **97**, 3453 (1975).
- (204) J. F. King, *Accounts Chem. Res.*, **8**, 10 (1975).
- (205) K. T. Douglas, A. Steltner, and A. Williams, *J. Chem. Soc., Chem. Commun.*, 353 (1975).
- (206) C. H. Clapp and F. H. Westheimer, *J. Am. Chem. Soc.*, **96**, 6710 (1974).
- (207) J. Wiseman and F. H. Westheimer, *J. Am. Chem. Soc.*, **96**, 4262 (1974).
- (208) (a) O. J. Scherer and N. Kuhn, *Chem. Ber.*, **107**, 2123 (1974); (b) E. Niecke and W. Flick, *Angew. Chem., Int. Ed. Engl.*, **13**, 134 (1974); (c) O. J. Scherer and N. Kuhn, *Angew. Chem., Int. Ed. Engl.*, **13**, 811 (1974).