Optically Active N-Protected α -Amino Aldehydes in Organic Synthesis

JANUSZ JURCZAK* and ADAM GOŁĘBIOWSKI

Institute of Organic Chemistry, Polish Academy of Sciences, 01-224 Warszawa, Poland

Received February 16, 1988 (Revised Manuscript Received July 12, 1988)

Contents

I.	Introduction	149
II.	Physical and Chemical Properties of	150
	N-Protected α-Amino Aldehydes	
III.	Preparation Methods of N-Protected α -Amino	150
	Aldehydes	
	A. Reductive Methods	150
	B. Oxidative Methods	151
	C. Miscellaneous Methods	152
IV.	Basic Transformations of N-Protected α -Amino	153
	Aldehydes into Useful Synthons	
	A. Additions of Metalloorganic Reagents	153
	B. Aldol Condensation and Related Reactions	153
	C. Diels-Alder Cycloaddition and Related	155
	Reactions	
	D. Wittig-Type Reactions	157
٧.	Total Synthesis of Natural Products	158
	A. Dipeptide Isosteres	158
	B. Unusual Amino Acids	159
	C. Amino Sugars	160
	D. Miscellaneous	161
VI.	Conclusions	162
VII.	References	162

I. Introduction

The synthesis of optically active organic compounds is one of the most important problems of contemporary chemistry. Pure enantiomers attain increasing commercial interest, especially in the field of pharmaceutical products. During recent years, asymmetric synthesis has greatly contributed to progress in highly controlled formation of new chiral centers. These processes still remain the basic problems in the total synthesis of natural products. Preparation of the latter in an optically pure form by application of chiral starting materials is very advantageous, enabling precise programming and efficient realization of synthetic pathways. Many monosaccharides and their readily available derivatives are versatile substrates for the synthesis of optically active target molecules.² α -Amino acids are the second important natural source of chiral substrates, useful in stereocontrolled organic synthe $sis.^{2c,3}$

Aldehydes are important and versatile compounds, widely used in organic synthesis. In recent years there has been a growing interest in chiral nonracemic aldehydes because of the development of new and effective methods for controlling stereochemistry of several basic organic reactions, such as metalloorganic addition to the carbonyl group,⁴ aldol condensation,⁵ [4 + 2] cycloaddition with carbonyl heterodienophiles,⁶ etc. Protected α -hydroxy (1) and α -amino (2) aldehydes are of special interest, owing to their ready availability in both enantiomeric forms from natural sources (sugars



Janusz Jurczak was born in 1941 in Poland. He received his M.Sc. from the Warsaw Technical University in 1964, his Ph.D. (Prof. A. Zamojski) in 1970, and his D.Sc. (habilitation) in 1979 from the Institute of Organic Chemistry of the Polish Academy of Sciences in Warsaw, where he is currently Professor of Chemistry. In 1971–1972 he was a postdoctoral fellow with Prof. V. Prelog at the Swiss Federal Institute of Technology in Zurich. His main scientific interests include methodology of organic synthesis, especially application of high-pressure techniques, stereocontrolled syntheses of natural products, and supramolecular chemistry.



Adam Golębiowski was born in 1959 in Warsaw. He received his M.Sc. from the Warsaw Technical University in 1983 and his Ph.D. in 1987 from the Institute of Organic Chemistry of the Polish Academy of Sciences under the direction of Prof. J. Jurczak. He is currently a research associate at this Institute. His research interests include methodology of organic synthesis and stereocontrolled syntheses of natural products.

and α -amino acids, respectively) and to pronounced versatility due to the presence of both the formyl group and suitably protected hydroxy or amino functionality in the molecule.

Recently, several extensive reviews on the application of α -hydroxy aldehydes in organic synthesis have been

SCHEME 2

published; 2e,5a,c but there is no general survey concerning α -amino aldehydes. On account of the increasing interest of chemists in α -amino aldehydes, reflected by an augmenting number of relevant publications, and in view of our belief that their further potential applications may be very important, we resolved to gather and present the actual knowledge concerning the use of optically pure N-protected α -amino aldehydes in stereocontrolled organic synthesis.

In the present review we shall focus attention mainly on the reactions using the carbonyl group of N-protected α -amino aldehydes to form a new chiral center. The main contents of the review are preceded by a presentation of methods for the preparation of N-protected α -amino aldehydes as well as their physical properties and chemical reactivity.

II. Physical and Chemical Properties of N-Protected α -Amino Aldehydes

N-Protected α -amino aldehydes are usually colorless crystals or oils, well soluble in typical organic solvents. They are relatively unstable both chemically and configurationally, particularly in solution. For this reason their elemental analysis and optical rotation measurements should be considered as only approximate. Therefore, it is recommended to use these compounds immediately after preparation; however, if purification is necessary, two methods are available: flash chromatography on silica gel, or formation of much more stable semicarbazone, followed by simple chromatography and subsequent decomposition to return the pure aldehyde.

The optical stability of some N-protected α -amino aldehydes during chromatography on silica gel was first studied by Ito et al. \(^{10}\) (Table 1, Schemes 1 and 2). As shown in Table 1, the order of the extent of racemization of Cbz-L- α -amino aldehydes on silica gel was as follows: Cbz-S-Bzl-L-cysteinal \gg Cbz-L-phenylalaninal > Cbz-L-leucinal \gg Cbz-N-g-nitro-L-argininal. The authors \(^{10}\) proposed a racemization mechanism for compounds 3 involving the protonated form 4 and enol 5 (Scheme 1). Aldehydes 3 with an enol-stabilizing R'group, e.g., Cbz-S-Bzl-cysteinal, racemize extremely quickly during contact with silica gel.

Limited racemization of Cbz-N^G-nitro-L-argininal (6) seems to be related to its cyclic carbinolamine structure 7 (Scheme 2), which probably prevents the nitroargininal derivative 6 from racemization due to keto-enol tautomerism.

Further studies on the optical stability of N-protected α -amino aldehydes were carried out by Evans and co-

TABLE 1. Optical Stability of Selected α -Amino Aldehydes on Silica Gel

	deş racemiz			
α -amino aldehyde	0 ha	6 h ^a	22 ha	
Cbz-NG-nitro-L-argininal	0	5	9	
Cbz-L-leucinal	0	32	65	
Cbz-L-phenylalaninal	0	53	85	
Cbz-S-Bzl-L-cysteinal	7	99	100	

TABLE 2. Optical Stability of Boc-L-Leucinal during Storage

storage time, day	storage temp, °C	$[\alpha]^{24}$ _D , deg	L/L (HPLC)
0		+18.2	100
1	-30	+17.9	99/1
9	-30	+17.4	99/1
9	+24	+6.9	7/3

workers, ¹¹ who found that the reduction–oxidation procedure (BH₃·THF–CrO₃/Py) generates Boc- α -amino aldehydes with complete retention of chiral integrity (>99.5%). The optical lability of the crude aldehydes depends on their structure. Thus, as expected from previous studies, ¹⁰ Boc-L-phenylalaninal appeared to be much less stable than Boc-L-leucinal. Very illustrative results of optical stability investigations of Boc-L-leucinal during storage at various temperatures are shown in Table 2. These studies led to the conclusion that even Boc-L-leucinal subjected to any prolonged treatment regimen, including drying, could no longer be regarded as optically pure unless otherwise verified as such. ¹¹

Recently, two important reports on configurational stability of N-protected α -amino aldehydes have appeared. The first one by Lubell and Rapoport¹² describes the synthesis of N-(9-(9-phenylfluorenyl))-Lalaninal. Exposure to silica gel or to a nonnucleophilic base caused no detectable racemization. The PhFl N-protecting group also maintains the configurational integrity of L-alaninal during C–C bond-forming reactions, affording enantiomerically pure products from Wittig reactions, aldol condensations, and Grignard additions. ¹²

The second report by Garner and Park¹³ describes the synthesis of N,O-diprotected L-serinal 8 and L-threoninal 9. These differentially protected β -hydroxy α -

amino aldehydes were shown to be produced in a 93–95% enantiomeric excess. The configurational stability of compounds 8 and 9 during their purification either by vacuum distillation or by flash chromatography was also demonstrated. ¹³

III. Preparation Methods of N-Protected α -Amino Aldehydes

A. Reductive Methods

 α -Amino aldehydes are mainly obtained from α -amino acids and only on occasion are they obtained from

TABLE 3. Preparation of N-Protected α-Amino Aldehydes by Reductive Methods

X	Y	temp, °C	time	R′	R"	yield, %	ref
OMe, OEt	DIBAL	-50	50 min	Н	Cbz	33-68	10
	DIBAL	-78	6 min	H	Boc	85-97	14, 15
	DIBAL	-65	45 min	H	Boc	80	9
-N N	LiAlH₄	-20	45 min	H	Cbz	50	16
` <u></u> `	DIBAL	-40	30 min	H	Cbz, Boc	52-83	17
-N-	LiAlH ₄	-20	2 h	Н	Cbz, Boc	85–95	18
-N(OMe)Me	LiAlH4	0 ·	20 min	н	Boc	86-96	19, 20
-N_0	LiAlH ₄	0	20 min		PhFl	90	12
-O(O)CoAlk	$H_2/Pd-C$	5	4-11 h	Н	Ac	12-81	21
-Cl	$H_2/Pd-C$	100	3 h		Pht	60	22

TABLE 4. Preparation of N-Protected α-Amino Aldehydes by Oxidative Methods

(O)	temp, $^{\circ}\mathrm{C}$	time	\mathbf{R}'	R"	yield, %	ref
CrO ₃ /Py	-10	30 min	Н	Boc	56-67	11, 26
	-10	30 min	P	ht	35	25
DMSO/SO ₃ ·Py	20	10 min	H	Boc	70-95	25, 27
DMSO/(COCl) ₂	-63	30 min	Ĥ	Boc	90	25, 28
DMSO/TFAA	-78	1 h	H	Boc	88	29
DMSO/DCC	20	48 h	P	ht	43	21
,	а	а	H	Ts	а	30
PCC	25	4 h	Bzl	Ts	75	31
PDC	20	24 h	H	Boc	75-95	11, 32, 33

other chiral precursors. Usually the synthetic route proceeds via esters or active amides of α -amino acids. which are finally reduced. A second approach is based on α -amino alcohols obtained from α -amino acids, which are oxidized to afford the desired α -amino aldehvdes.

Procedures based on reduction of esters and/or active amides of N-protected α -amino acids are listed in Table

Diisobutylaluminum hydride (DIBAL) reduction of methyl or ethyl esters is often accompanied by some overreduction to the respective alcohols. 9,10,14,15 The same remarks apply to lithium aluminum hydride reduction of imidazolides. 16,17 However, the reduction of 3,5-dimethylpyrazolides is apparently free from overreduction.18

Recently, an efficient method without racemization and overreduction was reported by Fehrentz and Castro. ^{19a} Thus, the preparation of Boc- α -amino aldehydes 11 is based on reduction of N-methoxy-N-methyl carboxamides 10 with lithium aluminum hydride, 20 which proceeds through a stable lithium-chelated intermediate; further reduction of the lithium salt is precluded by intramolecular complexation, and the aldehyde 11 is obtained upon hydrolysis (Scheme 3). 19a Similar results were found by Lubell and Rapoport¹² during reduction of the respective isoxazolides. The Fehrentz and Castro method was also successfully applied to the synthesis of tripeptide C-terminal aldehydes. 19b

Other reductive methods involving catalytic hydrogenation of mixed carbonic-carboxylic anhydrides21 and acid chlorides²² are of limited applicability.

B. Oxidative Methods

Procedures based on oxidation of N-protected α -amino alcohols are listed in Table 4. The N-protected α-amino alcohols are best obtained by borane-tetrahydrofuran reduction of N-protected α -amino acids²³ or by sodium borohydride-lithium chloride²⁴ or sodium borohydride-calcium chloride²⁵ reduction of the corresponding methyl ester.

Collin's reagent was the first oxidizing reagent shown to be efficient, offering a racemization-free procedure for Boc-L-leucinal synthesis. 11 Various activated dimethyl sulfoxide oxidations are of special interest owing to their general nature. These methods are free from racemization and can be applied for oxidation of relatively unstable derivatives of such α -amino acids as tryptophan and methionine.7 Among these oxidation methods the Parikh-Doering procedure is the best documented^{25,27} and is effective not only in the synthesis

$$H_2N-CH-CH_2-OH$$
 R
 NAD^{\odot}
 $PH 8.7$
 $NADH$
 R
 $NADH$
 R
 $NADH$
 R
 $PHNH_2$
 $PHNH_2$

SCHEME 5°

R=Me, Et, Prn, CH=CH2, CH2CH=CH2

^a (a) See ref 4b; (b) PhtNH, Ph₃P, EtO₂CN=NCO₂Et; (c) TFA; (d) Pb(OAc)₄.

of simple N-protected α -amino aldehydes but also in the preparation of peptide C-terminal aldehydes. Oxidation of pentapeptide C-terminal alcohols produced enkephalin aldehydes in 60-70% yield.²⁷

Pyridinium dichromate (PDC) oxidation³² is suspected to cause racemization to various extents, depending on the type of α -amino aldehyde,¹¹ whereas pyridinium chlorochromate (PCC) was found to be convenient for oxidation of N-benzyl-N-tosyl α -amino alcohols without racemization.³¹

 α -Amino alcohols with nonpolar side chains, such as L-leucinol, L-phenylalaninol, and L-valinol, were found to be good substrates for alcohol dehydrogenase from horse liver, affording the respective α -amino aldehydes (Scheme 4).³⁴

This mild enzymatic method enables preparation of nonprotected α -amino aldehyde semicarbazones, which can be directly used for coupling to certain peptides via the free amino group. To favor complete oxidation of the alcohol, the reaction was carried out in the presence of semicarbazide to trap the aldehyde product, and flavine mononucleotide was used to regenerate the oxidized coenzyme. The resulting α -amino aldehyde semicarbazone was subsequently hydrolyzed at pH 1 in the presence of formaldehyde, regenerating the aldehyde function in good yield. 34

C. Miscellaneous Methods

A convenient and elegant synthesis of five Pht-L- α -amino aldehydes from 2,3-O-isopropylidene-D-glyceraldehyde (12) was recently described by Mulzer et al. ³⁵ (Scheme 5).

The key steps in the sequence are anti-selective addition of metalloorganic reagent to 12, Mitsunobu inversion substituting N-phthalimide for hydroxyl in compound 13, and acetonide hydrolysis and glycol

SCHEME 6°

^a (a) LiAlH₄; (b) NaIO₄; (c) o-dichlorobenzene, AcONa, 170 °C; (d) (i) O₃, (ii) DMS; (e) ClCO₂Bzl; (f) NaBH₄.

SCHEME 7ª

^a (a) See ref 39; (b) TMSN₃; (c) BH₃·SMe₂, NaBH₄ (cat.); (d) Me₂C(OMe)₂, Me₂CO, p-TsOH; (e) H₂/Pd-C; (f) (Boc)₂O or ClCO₂Bzl; (g) DIBAL.

cleavage affording N-Pht-D- α -amino aldehyde 16. The investigated examples testify to the general applicability of this method.

Ohfune and Kurokawa³⁶ described a practical method for the synthesis of N-protected serinal in both enantiomeric forms from L- or D-methionine (Scheme 6).

Methionine was used as starting material, since both enantiomers are available and the sulfur functional group is readily convertible to a double bond. Reduction of D-methionine derivative 17, followed by oxidation of the resulting alcohol, afforded sulfoxide 18. Thermal elimination provided optically pure unsaturated compound 19, which was then converted to N-Boc-L-serinal (20a) by ozonolysis.³⁶ Recently, a new method for preparation of N-Cbz-L-serinal (20b) was reported.³⁷ The synthesis is based on sodium periodate oxidation of suitably protected D-glucosamine 21 (Scheme 6).

Another example of the synthesis of chiral β -hydroxy α -amino aldehydes, starting from L- and D-tartaric acid, was described³⁸ (Scheme 7). This approach involves a novel oxirane ring opening with hydrozoic acid and selective reduction of α -hydroxy ester as key steps. Transformation of L-tartaric acid into epoxide 22 was performed via a three-step procedure according to Mori and Iwasawa.³⁹ The opening of the oxirane ring with

SCHEME 9

hydrazoic acid, generated in situ from trimethylsilyl azide, afforded compound 23 in a 97% yield. The next crucial step consisted of reduction of the ester group in position α to the hydroxy group. For this regioselective transformation, a reducing system⁴⁰ containing borane, dimethyl sulfide, and catalytic amounts of sodium borohydride was applied. The resulting compound 24 was then treated with dimethoxypropane to produce the stable derivative 25, which was reduced to amine 26. The final steps involved protection of the amino group and reduction of the ester group with DIBAL to afford the desired N-protected α -amino aldehvde derivatives 28.

IV. Basic Transformations of N-Protected α-Amino Aldehydes Into Useful Synthons

A. Additions of Metalloorganic Reagents

Reactions of metalloorganic reagents with chiral aldehydes are of considerable interest in the context of acyclic stereoselective synthesis.2e This type of reaction with N-protected α -amino aldehydes is often used in the synthesis of peptide isosteres.⁷

Simple addition of vinylmagnesium bromide (30) to Boc-L-phenylalaninal (29), carried out at -78 °C in tetrahydrofuran, afforded a 56:44 mixture of syn (threo) and anti (erythro) allylic alcohols 31 and 32, respectively (Scheme 8).41 Similarly, low diastereoselectivity was reported by Thaisrivongs et al.⁴² for addition of 30 to N-trityl-L-leucinal.

The diastereoselectivity was improved in favor of the chelation-controlled Cram product 31 by carrying out the addition reaction at 25 °C, which gave a 70:30 ratio of 31 to 32. At higher temperatures a greater proportion of NH protons should be removed to the transition state A prior to addition to the aldehyde carbonyl group, resulting in preferential formation of syn alcohol 31 (Scheme 8).⁴¹

Addition of the chiral Grignard reagent 34 to Boc-Lleucinal (33) was characterized by higher diastereoselectivity.43 The syn product 35 was formed as the major diastereoisomer (ratio of 35 to 36 was 4:1), presumably

SCHEME 10

NPht

AcO
$$\stackrel{\square}{\longrightarrow}$$
 H + Me₂Al $\stackrel{\square}{\longrightarrow}$ C₁₃H₂₇ $\stackrel{\square}{\longrightarrow}$

37 38

NPht

AcO $\stackrel{\square}{\longrightarrow}$ C₁₃H₂₇ + AcO $\stackrel{\square}{\longrightarrow}$ C₁₃H₂₇

OH OH

39 4 : | 40

SCHEME 11

SCHEME 12

due to double asymmetric induction (Scheme 9).

Addition of the lithium derivative of ethyl propiolate to Boc-L-leucinal (33) afforded corresponding hydroxyacetylenic esters as a mixture of diastereoisomers.44 Further transformations proved that also in this case the syn diastereoisomer was formed predominantly (syn:anti ratio was 4.5:1).

Addition of vinylalane 38 to O-Ac-N-Pht-L-serinal (37), carried out in a 2:1 mixture of benzene and ether at 5-10 °C, afforded the desired alcohols anti-39 and syn-40 in a 4:1 ratio (Scheme 10).45a

Addition of ((Z)-3-pentadecenyl)magnesium bromide to the aldehyde 37 resulted in a ca. 7:1 mixture of anti to syn products in 18.9% yield.45b

Similar additions of vinylalanes were studied by two other groups. 46,47 In both cases, the oxazolidine form of L-serinal 41 was used. Addition of metalloorganic reagent 42 to aldehyde 41 proceeded without any selectivity, affording an equimolar mixture of alcohols 43 and 44 (Scheme 11).47

Similarly, ylides of type 45 reacted with N-protected α -amino aldehydes, e.g., 29, to afford an equimolar mixture of two diastereoisomeric epoxides 46 and 47 (Scheme 12).⁴⁸

B. Aldol Condensation and Related Reactions

The renaissance of the aldol condensation observed during the past decade resulted from the development

SCHEME 14

52

of new methods of organic synthesis, enabling stereocontrol of the reaction course. A number of syntheses of natural products, starting from N-protected α -amino aldehydes, involves aldol condensation as the key step. Unfortunately, aldol condensation with N-protected α -amino aldehydes is characterized, similarly as in case of metalloorganic addition, by rather low diastereoselectivity. In only one case this reaction with N-protected α -amino aldehydes was found to attain high diastereoselectivity. Is

6.5

5(

Simple condensation of the lithiated acetic ester with Boc-L-leucinal (33) provided a 1:1 mixture of diastereoisomeric alcohols 48 and 49 (Scheme 13).^{11,14}

Significant diastereoselectivity was observed for β -substituted N-protected α -amino aldehydes. Condensation of aldehydes 50a with lithiated tert-butyl acetate afforded a 6.5:1 mixture of diastereoisomeric alcohols syn-51 and anti-52 in a 96% yield (Scheme 14).³³ The selectivity of this reaction was obviously dependent upon the configuration of the β -substituent and decreased to a 3:1 ratio of the respective alcohols when the anti aldehyde 50b was employed under the same reaction conditions. As expected, condensation of aldehyde 50c with lithiated tert-butyl acetate afforded an equimolar mixture of diastereoisomers. The authors³³ proposed for this reaction a model in which both α -chelation and the bulky group in the β -position play an important role in stereocontrol of the reaction course.

The cyclic structure of N-protected α -amino aldehydes, e.g., Boc-L-prolinal (53), seems to play a similar role; the lack of the NH proton in such structure has to be stressed. Aldol condensation of 53 with lithiated ethyl acetate produced a 4:1 mixture of diastereoisomeric alcohols syn-54 and anti-55 (Scheme 15).⁴⁹

This degree of diastereoselectivity is substantial when compared with the results of a similar aldol condensation with the acyclic aldehyde 33 (cf. Scheme 13). Noteworthy is the fact that use of vinylmagnesium bromide instead of lithiated ethyl acetate in the reaction

SCHEME 15

SCHEME 16

SCHEME 17

with aldehyde 53, gave a 1:1 mixture of diastereoisomers. Also in the case of 4-substituted N-protected L-prolinal, reaction with even chiral lithium enolate led to a mixture of diastereoisomers in an only 3:1 syn:anti ratio.²⁹

Boc-L-Cyclohexylaninal (56) reacted with the lithium enolate of acetonitrile similarly as with Boc-L-leucinal or with Boc-L-phenylalaninal, to afford a 3:2 diastereoisomeric mixture of alcohols syn-57 and anti-58 (Scheme 16).⁵⁰ The low diastereoselectivity found for the above aldol condensation can be explained by competition between chelation and steric factor control.

Aldol condensation of N-trityl-L-phenylalaninal (59) with the lithium enolate of dimethyl phosphonate (60) afforded a 3:1 anti:syn ratio of alcohols 61 and 62 (Scheme 17).⁵¹ The same reaction with Boc-L-phenylalaninal gave a 1:1 diastereoisomeric mixture. The above results are consistent with the Cram model for asymmetric induction when the benzyl moiety is assumed to be a large substituent. The trityl protecting group is very bulky and prevents chelating interaction between lithium and the NH proton, whereas the Boc group is not sufficiently bulky.

Titanium-mediated condensation of dilithiated N-alkylmethacrylamides with N-protected α -amino aldehydes also resulted in moderate diastereoselectivity. For example, Boc-L-phenylalaninal (29) reacted with

SCHEME 19

TABLE 5. Aldol Condensation of N-Protected L-Alaninal Derivatives 66a-c with (Vinyloxy)boranes 67a,b

major product	temp, °C	time, min	overall yield, %	diaster ratio
68a	0	30	70	8:1
68b	0	30	60	35:1
68c	0	30	62	20:1
68d	0	30	60	10:1
68e	0	60	64	8:1

dilithiated N-methylmethacrylamide (63) in the presence of chlorotitanium triisopropoxide to afford a 3:2 mixture of diastereoisomeric hydroxy amides 64 and 65 (Scheme 18).⁵² The same reaction with Boc-L-cyclohexylalaninal or Boc-L-leucinal led to a ca. 1:1 mixture of diastereoisomers.

In light of the above-mentioned examples, the results obtained by Umezawa et al. 18a seem to be very promising. Aldol condensation of variedly N-protected Lalaninals (66a-c) with (vinyloxy)borane derivatives (67a,b) was examined under different reaction conditions (Scheme 19). The results are summarized in Table 5.

The overall yield of the diastereoisomeric aldol products was generally good. The high diastereoselectivity of this reaction was rationalized by using the commonly accepted six-membered cyclic transition state.18a

Only one example of the Reformatsky reaction with N-protected α -amino aldehydes is known.⁵³ Thus, in the reaction of Pht-L-leucinal (70) with the organozinc derivative of isopropyl acetate, a 55:45 mixture of syn-71 and anti-72 alcohols is obtained (Scheme 20). This result is in good agreement with those obtained for simple aldol condensation (cf. Schemes 13 and 16).

SCHEME 20

SCHEME 21

SCHEME 22a

^a (a) ZnCl₂; (b) HMPT-THF, -78 °C; (c) TFA; (d) PPTS.

C. Diels-Alder Cycloaddition and Related Reactions

Since the early observations on the ability of activated carbonyl compounds to undergo the Diels-Alder reaction with 1,3-dienes, this field of organic chemistry has been greatly explored.⁵⁴

In basic studies of the cyclocondensation reaction of activated 1,3-dienes with various aldehydes, Danishefsky et al. 55 tested Boc-L-leucinal (33) as a dienophile component. The reaction of diene 73 with aldehyde 33 catalyzed by zinc chloride led to a 9:1 diastereoisomeric mixture of pyrones 74 and 75 (Scheme 21). This high syn-anti selectivity can be explained by a strong chelating interaction forming the quasi-cyclic structure of N-protected α -amino aldehyde as well as by the high steric demand of the approaching planar diene conformer.

High diastereoselectivity was also observed for the reaction of the same diene 73 with L-serine derivative 8 (Scheme 22).⁵⁶

These results are also consistent with chelation-controlled cycloaddition of diene 73 to aldehyde 8 in conformation B from the less hindered si face, to afford a 60:1 mixture of syn-76 and syn-77 pyrones (Scheme 23).

SCHEME 24

Obviously, obtainment of the opposite (anti) diaster-eoselectivity in this reaction creates some difficulties. In view of the possibility of two competitive routes for the cyclocondensation reaction of diene 73 with aldehydes, ^{6a} Garner and Ramakanth⁵⁶ proposed an ingenious method for overcoming these difficulties. Addition of lithium enolate 78 to aldehyde 8 afforded epimeric aldol products 79, which were readily converted to a 1:7 mixture of syn-76 and anti-77 (Scheme 22). This was due to the predominance of the Felkin-Anh⁵⁷ conformation C of aldehyde 8 and to the attack of diene 73 from the less hindered re face (Scheme 23).

In long-term studies by Jurczak and Gol/ębiowski⁵⁸ on N-protected α -amino aldehydes, a search was conducted for a more general anti-selective strategy. Initial investigations involved the reaction of diene 80 with two O-protected N-Cbz-L-threoninals 81a and 81b in the presence of zinc bromide as catalyst (Scheme 24).

The results pointed out a strong preference of α -chelation interaction leading in both cases to formation of syn-82 pyrones as the major diastereoisomers. For this reason the high-pressure techniques⁵⁹ was applied, enabling milder reaction conditions and no need for the strong Lewis acid catalysis used in the previous case. Thus, the high-pressure reactions of diene 84 with variedly N-protected L-alaninal (66a-c) and D-alaninal (87a-c) catalyzed by 1–5% Eu(fod)₃⁶⁰ were extensively studied^{15,58} (Scheme 25).

The results testified to substantial reversion of stereoselectivity for Pht-L- (66a) or -D-alaninal (87a) when compared with the Cbz- or Boc-protected analogues (66b,c and 87b,c). This was probably caused by the removal of both NH protons from the amino group paralleled by nitrogen atom deactivation due to the presence of two carbonyl groups in the phthalimide moiety. The necessity of the occurrence of a free NH proton for chelation control was also confirmed by two experiments under very high pressure (25 kbar). The

SCHEME 25

SCHEME 26

noncatalyzed reaction of Cbz-L-alaninal (66b) with diene 84 led to a 2:1 mixture of syn-85b and anti-86b adducts (Scheme 25). The same reaction was carried out in the presence of catalytic amounts of Eu(fod)₃ to afford a higher yield of the cycloadducts 85b and 86b, but the diastereoselectivity was the same as in the previous case. These facts can be interpreted from the three possible conformations of aldehyde 66b: the Felkin-Anh form D, chelating form E, and hydrogenbonded form F (Scheme 26). It can be expected that in the catalyzed reaction conformation E will predominate over conformation D. On the other hand, in the noncatalyzed reaction conformation F is preferred over conformation D.⁵⁸

High-pressure [4 + 2] cycloadditions of diene 84 to variedly O-protected N-Cbz-D-threoninal (81a,b) or N-Cbz-D-allo-threoninal (92a,b) catalyzed by 2-3% Eu(fod)₃ were also studied;⁵⁸ the results are shown in Scheme 25. Explanation of these results is also based on an analysis of the transition-state stereochemical models G, H, and J (Scheme 26). For aldehydes 81b and 92b (R = TBDPS) conformation H should pre-

SCHEME 28

dominate over conformation G, leading to diastereoisomers syn-91b and syn-94b, respectively, as major products. When the bulky tert-butyldiphenylsilyl group was replaced by the strongly chelating (benzyloxy)methyl group (BOM), the population of conformation J substantially increased, giving rise to a higher yield of diastereoisomers anti-90a and anti-93a when aldehydes 81a and 92a, respectively, were used (Scheme 25).58

D. Wittig-Type Reactions

In N-protected α -amino aldehydes, the carbonyl group not only participates in reactions leading to formation of a new chiral center but also may be transformed into other functionalities. This section presents a discussion of the Wittig reaction, 61 permitting introduction of a C=C bond to replace the formyl group of N-protected α -amino aldehydes. The mechanism of the Wittig reaction still remains unclear, and thus the factors governing Z-E selectivity are not yet well-known.⁶² Nevertheless, some conclusions concerning the effect of the type of Wittig reagent and of the reaction conditions on Z-E selectivity were reached. For example, it is known that application of a stabilized organophosphorus compound in nonpolar solvents yields products predominantly with the E configuration, whereas in alcohol-type solvents isomer Z predominates. In the case of a nonstabilized Wittig reagent, there is usually a predominance of the Z isomer.

In N-protected α -amino aldehydes, the chiral center seems to play no part in the stereochemistry of the reaction. The E/Z ratio mainly depends on the nature of the ylide and on the reaction conditions. Recently, Luly et al. 25 took up the problem of racemization during this reaction. In studies of the reaction of ylide 95 with Boc-L-leucinal (33), the resulting olefin 96 and starting aldehyde 33 exhibited a similar degree of optical purity (Scheme 27). This result can be rationalized by a racemization mechanism involving enol formation, which is generally accepted for N-protected α -amino aldehydes. 10

The Wittig reaction of Cbz-L-threoninal derivative 97 with acetonylidenetriphenylphosphorane (98) afforded enone 99 without any perceptible epimerization (Scheme 28), 63

Likewise, the reaction of Boc-L-alaninal (66c) with 2-(triphenylphosphoranylidene)propionate (100) afforded E-olefin 101 as the predominant product (E/Z)ratio was 95:5) (Scheme 29).64

In the case of the reaction of Cbz-L-alaninal (66b) with chiral ylide 103, Z-olefin 104 was exclusively formed (Scheme 30).65

SCHEME 29

SCHEME 30

SCHEME 31

TABLE 6. Olefination of N-Benzyl-N-tosyl-L-phenylalaninal (105)

phosphonate	products	temp, °C	base	yield, %	Z:E ratio 107:108
106a	107a + 108a	0	NaH	84	1:2
106a	107a + 108a	-78	KH	93	1:8.5
106 b	107a + 108a	0	NaH	100	1:1
106 b	107a + 108a	-78	KH	80	1:9
106c	107b + 108b	0	NaH		1:2
106c	107b + 108b	-78	NaH		2:3
106c	107b + 108b	-78	KH	41	2:3
106 d	107c + 108c	0	NaH	98	5:1
106 d	107c + 108c	-78	NaH	8i	3,3:1
106 d	107c + 108c	-78	KH	56	3,3:1
106e	107c + 108c	0	NaH	91	7:1
106 f	107b + 108b	-78	KH	79	25:1
106 f	107b + 108b	0	KH	45	20:1
106g	107a + 108a	-78	KH	79	70:1

Very interesting studies on the selectivity of the reaction between N-benzyl-N-tosyl-L-phenylalaninal (105) and a variety of phosphonates and phosphononitriles were recently reported by Hensel and Fuchs⁶⁶ (Scheme

31). The results are summarized in Table 6.

The best Z selectivity was obtained with highly electrophilic β -trifluoroethyl phosphonates 106f and 106g. More bulky phosphonate esters (e.g., 106b) permit equilibration of the β -oxido phosphonate intermediates and a shift of the product distribution toward the thermodynamically preferred E-olefin isomers.

Kogen and Nishi⁶⁷ carried out similar investigations concerning several Cbz-protected α -amino aldehydes of the type of 109 (Scheme 32). They also obtained a very good Z selectivity for the highly electrophilic β -trifluoroethyl phosphonate 110.

V. Total Synthesis of Natural Products

A. Dipeptide Isosteres

A new branch of synthetic chemistry in the peptide field appeared when chemists started to modify the peptide backbone itself, while maintaining the original side chains. In this way an increased stability toward enzymatic degradation can be obtained, leading to a prolonged or a more selective activity of peptide hormones. Alternatively, nonhydrolyzable peptide analogues can be designed to inhibit a variety of enzymes. A number of such peptide backbone modifications are known, e.g., $-CH_2$ -, -NH-, or -O- intercalations, α -aza, α -alkyl, and N-alkyl peptides, depsipeptides, phosphonamidates, etc. 7

Dipeptide isosteres, compounds in which the peptide bond is replaced by some approximately isosteric functional groups, e.g., hydroxylmethylene, ketomethylene, thiomethylene, and the carbon-carbon E double bond, proved to be valuable for producing enzyme inhibitors and proteolytically stable peptides.⁷

N-Protected α -amino aldehydes are very suitable starting materials for modified peptide synthesis. Moreover, some naturally occurring peptide enzyme inhibitors, such as leupeptin, antipain, chymostatin, and elastinal, ⁶⁹ have an amino aldehyde function at the C-terminal part.

Evans et al. 48 described the stereocontrolled synthesis of the isosteric hydroxymethylene dipeptide 113. The method is based on the reaction between ylide 45 and Boc-L-phenylalaninal (29), leading to a mixture of epoxides 46 and 47, which was then used for the synthesis of several dipeptide analogues of the type of 113 (Scheme 33).

SCHEME 33

SCHEME 34°

64

a (a) Xylene, reflux; (b) +"R".

SCHEME 35

A general method for the synthesis of isosteric hydroxymethylene dipeptides was recently reported by Kempf.⁵² Pyrolytic cyclization of the compound 64, obtained from Boc-L-phenylalaninal (29) according to Scheme 18, afforded derivative 114. Addition of nucleophiles to the double bond of 114 led to protected 115 (Scheme 34).

Hanson and Lindberg⁴¹ achieved the synthesis of dipeptide analogues through the reaction of vinylmagnesium bromide (30) with Boc-L-phenylalaninal (29), affording a mixture of allylic alcohols 31 and 32 (cf. Scheme 8). After chromatographic resolution, pure diastereoisomers 31 and 32 were separately transformed into the respective dipeptide analogues 116 and 117 (Scheme 35). A similar approach to the synthesis of dipeptide analogues was recently reported by Plattner et al.⁷⁰

SCHEME 36°

^a(a) BuLi, Ph₃P(Br)CH₂C≡CTMS; (b) (Sia)₂BH; (c) H₂O₂, OH⁻.

SCHEME 37°

^a(a) NaBH₃CN, pH 6-8; (b) (Boc)₂O; (c) (i) O₃, (ii) DMS; (d) L-Asp-OBzl; (e) H₂/Pd-C; (f) PDC.

SCHEME 38

An interesting strategy for the synthesis of an olefinic isostere 120 was applied by Sammes et al. 9,71a and later by Johnson^{71b} (Scheme 36). The Wittig reaction of aldehyde 118 with an ylide generated from Ph₃P(Br)- $CH_2C \equiv C(TMS)$ afforded acetylene derivative (E)-119. which was then oxidized by using the Zweifel-Backlund procedure⁷² to produce acid 120. Compound 120 was used as a substrate in several isosteric peptide syntheses.

Poly(amino acids) in which the amino acid moieties are linked by the N-alkyl bond attracted considerable attention since they possess interesting biological activities.⁷³ Ohfune and Kurokawa³⁶ recently reported the synthesis of a typical representative of this class of compounds—aspergillomarasmine A acid (Scheme 37). Amine 121 and N-Boc-L-serinal (20a), both obtained from D-methionine according to the procedure shown in Scheme 6, were converted to diol 122 by reductive coupling. Protection of the free amino group, followed by ozonolysis and coupling with L-aspartic acid benzyl ester, afforded the acid 123. Deprotection of the ester group and oxidation of the free hydroxy group should give the required acid 124, but because of steric hindrance, the central hydroxy group remained unchanged.36

Reductive coupling was also used as the key step in the synthesis of simple peptide isostere 126 (Scheme $38).^{28,74}$

B. Unusual Amino Acids

The synthesis of unusual amino acids is of growing interest. 14b,42,75 L-Statine (127)76 is one of the most important members in this class of compounds because of its presence in a number of recently discovered protease inhibitors, such as pepstatine, renin, etc. A simple and efficient synthesis of L-statine was reported by Rich et al. 14a (Scheme 39).

SCHEME 39°

^a(a) LDA; (b) OH⁻; (c) H⁺.

SCHEME 40

SCHEME 41°

^a(a) DIBAL; (b) m-CPBA; (c) Red-Al; (d) O₂, Pt.

SCHEME 429

^a (a) NaHSO₃; (b) KCN; (c) HCl; (d) crystallization.

A similar approach was subsequently used by Evans et al. 11 Aldol condensation of Boc-L-leucinal (33) with lithiated ethyl acetate afforded an equimolar syn-anti mixture of the respective esters in 80% yield (cf. Scheme 13). After chromatographic separation, the pure diastereoisomer 48 was hydrolyzed and deprotected to yield optically pure L-statine (127).

A different approach to the L-statine synthesis was reported by Danishefsky et al.⁵⁵ Pyrone 74 obtained from diene 73 and Boc-L-leucinal (33) according to the procedure shown in Scheme 21 was subjected to ozonolysis followed by oxidative fragmentation to afford the ethyl ester of N-Boc-L-statine (128) (Scheme 40).

An efficient synthesis of N-Cbz-L-statine (131) was recently reported by Kogen and Nishi.⁶⁷ Unsaturated ester 111b, obtained from Cbz-L-leucinal via the Wittig reaction according to the procedure shown in Scheme 32, was reduced and then epoxidized to afford compound 129 as a major product (ratio = 28:1). The epoxide syn-129 was reduced, giving with very high selectivity the desired α -amino alcohol syn-130. Selective oxidation of the primary hydroxy group provided N-Cbz-L-statine (131) in 95% yield (Scheme 41).

L-Statine and its analogues were also obtained by Dellaria and Maki⁵¹ as well as by Rich et al.⁷⁷

A nonselective approach to the synthesis of another important unusual amino acid, bestatine, 78 was reported by Umezawa et al. 18b,79 Addition of sodium hydrogen sulfite to Cbz-D-phenylalaninal (132) afforded an

SCHEME 43a

 a (a) (i) ZnCl2, (ii) HCl; (b) (i) NaIO4, RuO2·H2O (cat.), (ii) HCl, (iii) CH2N2; (c) (i) Et2NTMS, (ii) flash chromatography; (d) MeOH, p-TsOH (cat.); (e) (i) KMnO4, NaOH(aq), (ii) HCl.

equimolar mixture of epimers 133, which was transformed into a mixture of cyanohydrins 134. Hydrolysis of 134, followed by fractional crystallization, provided diastereoisomerically pure N-Cbz-D-bestatine (135) in 25% overall yield (Scheme 42).

The synthesis of syn- β -hydroxy-L-glutamic acid (140) was described by Garner. ⁸⁰ Cyclocondensation of diene 73 with aldehyde 136 led to a >9:1 mixture of pyrones 137. Subsequent oxidative degradation followed by esterification and protection of the hydroxy group led to compound 138, which was converted to diol 139 and finally oxidized to afford the desired amino acid 140 (Scheme 43).

Detoxinine (145) is an amino acid component of detoxine (141), a selective antagonist of the antibiotic blasticidin S.⁸¹ Recently, Ohfune and Nishio³³ presented a strategy for the synthesis of detoxinine (145). The starting material was Boc-L-allylglycine (142), which after allylic oxidation, followed by protection of the resulting hydroxy group afforded, after chromatographic separation, diastereoisomerically pure compound 143 in ca. 40% overall yield. Transformation of 143 into aldehyde 50a, followed by its condensation with the lithium enolate of tert-butyl acetate, yielded, after chromatographic separation, the pure aldol product 51; ozonolysis, Wittig homologation, and subsequent cyclization afforded compound 144, which was finally deprotected to give the required detoxinine (145) (Scheme 44).

Another approach to the synthesis of detoxinine (145) was recently reported by Joullie et al.²⁹ The key step in this synthesis is a stereoselective aldol condensation of N,O-protected 3-hydroxy-L-prolinal with the chiral enolate derived from L-mandelic acid to afford the respective alcohol, which was transformed into detoxinine (145).

C. Amino Sugars

The modern approach to the synthesis of sugars via [4+2] cycloaddition^{6,82} offers some new applications of N-protected α -amino aldehydes. The simplest nonchiral α -amino aldehyde, Boc-glycinal, was reacted with diene 84 under high-pressure conditions to afford after acidic isomerization the racemic adduct 146 in good yield.⁸³ Adduct 146 was the starting compound in the synthesis of D,L-purpurosamine C, which is one of the two sugar components of the aminoglycoside antibiotic gentamicin G_{1a} .⁸⁴ Hydroboration of adduct 146, followed by oxidative workup, afforded alcohol 147, which after oxidation yielded ketone 148. Transformation of

SCHEME 44a

 a (a) (i) SeO₂, Bu^tOH, (ii) TBDMSCl, imidazole; (b) (i) LiAlH₄, (ii) PDC; (c) LDA, AcOBu^t; (d) (i) O₃, (ii) DMS, (iii) Am^tONa, Ph₃P(Cl)CH₂OMe, (iv) Hg(OAc)₂, (v) KI, (vi) NaBH₄, (vii) Ph₃P, NBS, (viii) NaH; (e) (i) CSA, (ii) TFA, (iii) 1 N NH₃.

SCHEME 45°

^a(a) (i) ThxBH₂·DMS, (ii) H₂O₂, OH⁻; (b) PCC, 4A molecular sieves; (c) NH₂OH·HCl; (d) Ac₂O, Et₃N; (e) BH₃·THF; (f) TFA.

ketone 148 into an oxime, followed by acetylation, provided derivative 149, which was readily converted to methyl N,N-diacetyl-D,L-purpurosaminide C (150) in 25% overall yield (Scheme 45).⁸⁵

A similar approach was applied to the total synthesis of 6-epi-D-purpurosamine B $(151)^{15}$ and D-purpurosamine B $(152)^{58}$ the components of the aminoglycoside antibiotics fortimicin A and gentamicin C_2 . Starting from either N-protected L-alaninal (66b,c) or D-alaninal (87a) and controlling diastereoselectivity of the high-pressure [4+2] cycloaddition reaction with diene 84, adducts 85b,c and 88a were obtained, respectively (cf. Scheme 25). These adducts were transformed via the above-presented route⁸⁵ into the desired derivatives of purpurosamines 6-epi B and B (Scheme 46). 15,58

Another stereocontrolled synthesis of 6-epi-D-purpurosamine B derivative 155 was based on iodo-cyclocarbamation of olefin 104, obtained from Cbz-L-alaninal (66b) via Wittig reaction (cf. Schemes 30 and 47).⁶⁵ The C-5, C-6, and C-7 carbon atoms derived from N-protected L-alaninal whereas the remaining part of the carbon skeleton originated from L-malic acid.

Retrosynthetic analysis of lincosamine (156), the sugar component of the antibiotic lincomycin⁸⁷ (Scheme

SCHEME 47°

 $^a(a)$ BzlBr, NaH; (b) I₂; (c) Bu₃SnH; (d) 1 N HCl; (e) PhCOCl, Et₃N; (f) MsCl, Et₃N; (g) NaN₃; (h) Ba(OH)₂; (i) H₂/Pd(OH)₂; (j) ClCO₂Bzl; (k) RuCl₂·(PPh₃)₃; (l) Ac₂O, Py.

SCHEME 48

48), offers the possibility of application of a [4 + 2] cycloaddition reaction between dienes 80 or 84 and N,O-protected D-allo-threoninal 159 as the key step in the synthetic sequence. Moreover, the use of any one of the four threonine diastereoisomers, combined with stereocontrol of cycloaddition, permits an easy access to each diastereoisomer of lincosamine modified in the C-5, C-6, or C-7 positions.⁵⁸ These diastereoisomers of lincosamine, important in biological activity studies, are

SCHEME 49

° (a) $ZnCl_2$; (b) TFA; (c) $NaBH_4$, $CeCl_3$; (d) NH_3 , MeOH; (e) Ac_2O , Et_3N ; (f) m-CPBA; (g) H_2/Pd -C; (h) Bu_4NF .

SCHEME 50

difficult to obtain by traditional methods.

Preliminary studies of the above-mentioned systems showed the predominance of the α -chelating interaction in the cyclocondensation step, resulting in high syn selectivity (cf. Scheme 26). Further transformations based on the Danishefsky et al. 88 approach afforded a diastereoisomer 164 of peracetylated lincosamine (Scheme 49). 58

Recently Garner and Ramakanth⁵⁶ reported the results of preliminary studies on the total synthesis of the amino sugar related antibiotic amipurimycin (165).⁸⁹ Retrosynthetic analysis of the sugar part 166 of this antibiotic (Scheme 50) suggested the possibility of applying the cyclocondensation reaction between diene 73 and N,O-protected L-serinal 8 (cf. Scheme 22) as a way for construction of the sugar skeleton. The branching at the C-3 position in 166 can be introduced via a hydroxyl-directed Wittig reaction.⁹⁰

D. Miscellaneous

As shown above, N-protected α -amino aldehydes have been used mainly for the synthesis of peptide analogues, unusual amino acids, and amino sugars. Cerebroside $(167)^{91}$ is an example of a natural compound not belonging to those classes that is obtainable from an N-protected α -amino aldehyde. Retrosynthetic analysis of 167 (Scheme 51)⁴⁵⁻⁴⁷ showed that amino diol 169, a crucial intermediate for the synthesis of cerebroside, can be readily prepared by metallorganic addition to aldehyde 41 according to the procedure presented in Scheme 11.

Another application of chiral α -amino aldehydes in the total synthesis of natural products was recently reported by Kozikowski et al.⁶³ The synthesis of a broad-spectrum antibiotic, actinobolin (170),⁹² was based on the [4 + 2] cycloaddition reaction of diene 172 with ketone 171 (Scheme 52). Compound 99, a pre-

SCHEME 52

SCHEME 53

cursor of diene 172, was obtained from N,O-protected L-threoninal 97 via the Wittig reaction as shown in Scheme 28.

Recently, Fujii et al.⁶⁴ described the total synthesis of 1'-methylzeatin (173), which is a new representative of cytokinins.⁹³ Retrosynthetic analysis (Scheme 53) showed that compound 101, a key intermediate in this synthesis, can be easily obtained from Boc-D-alaninal (87c) by the procedure presented in Scheme 29.

(S)-4-((Methoxycarbonyl)methyl)-2-azetidinone (180), a crucial intermediate in the synthesis of (+)-thienamycin, ⁹⁴ was obtained by Yamada et al. ⁹⁵ from 2,3-O-isopropylidene-D-glyceraldehyde (12) (Scheme 54). In a sequence of five steps, compound 12 was transformed into α -amino aldehyde 177, which via the Wittig reac-

SCHEME 54°

 $^{\rm o}$ (a) BuLi, Ph₃P(Br)CH₂CO₂Me; (b) BzlNH₂; (c) ClCO₂Bzl; (d) 75% AcOH; (e) NaIO₄; (f) BuLi, Ph₃P(Br)CH₂OMe; (g) Jones reagent; (h) H₂/Pd–C; (i) Ph₃P–(PyS)₂, MeCN.

tion, followed by oxidation, was converted to the chiral monoester of the dicarboxylic acid (179). After deprotection of the amino group, compound 179 was cyclized to the desired azetidinone derivative 180.

Apart from the above examples, some other natural products were synthesized by using N-protected α -amino aldehydes. Bleomycin, an antitumor antibiotic, 96 was obtained by Umezawa et al. 97 using as the key synthon (2S,3S,4S)-4-amino-3-hydroxy-2-methylpentanoic acid, readily obtainable from N-protected D-alaninal via aldol condensation. 18a N-benzyl-N-to-syl-L-phenylalaninal (105)66 was used for preparation of chiral diene, which was then applied in the total synthesis of cytochalasin C.98

VI. Conclusions

As can be seen from the above-presented literature data, N-protected α -amino aldehydes are versatile chirons, widely recognized, inexpensive, and easily accessible from natural sources. However, the degree of stereoselectivity obtained in some reactions shown is not high enough to meet the present requirements, and thus more work has to be done to elucidate the nature of all factors responsible for asymmetric induction. Higher stereoselectivities will surely extend the utility of these valuable chiral synthons. Finally, we believe that the near future will bring many more examples of synthetic sequences starting from N-protected α -amino aldehydes. The very recent papers published during the past few weeks seem to testify to our optimism. ^{99,100}

Acknowledgments. We thank all colleagues whose assistance was very helpful in our work in this review. Financial support by the Polish Academy of Sciences (Grant CPBP 01.13) is acknowledged.

VII. References

 Recent reviews: (a) Whitesell, J. K. Acc. Chem. Res. 1985, 18, 280. (b) ApSimon, J. W.; Collier, T. L. Tetrahedron 1986, 42, 5157. (c) Morrison, J. D., Ed. Asymmetric Synthesis, Academic Press: New York, 1983, 1984; 5 vols.

(2) (a) Vasella, A. In Modern Synthetic Methods 1980; Scheffold, R., Ed.; Salle & Sauerländer: Frankfurt, 1980; p 173. (b) Hanessian, S. Total Synthesis of Natural Products: The "Chiron" Approach, Pergamon Press: Oxford, 1983. (c)

- Scott, J. W. In Asymmetric Synthesis; Morrison, J. D., Ed.; Academic Press: New York, 1984; p 1. (d) Inch, T. D. Tetrahedron 1984, 40, 3161. (e) Jurczak, J.; Pikul, S.; Bauer, T.

- rahedron 1984, 40, 3161. (e) Jurczak, J.; Pikul, S.; Bauer, T. Ibid. 1986, 42, 447.
 (3) Martens, J. Top. Curr. Chem. 1984, 125, 165.
 (4) (a) Yamamoto, Y.; Yatagi, H.; Ishihara, Y.; Maeda, N.; Maruyama, K. Tetrahedron 1984, 40, 2239. (b) Maed, K.; Macdonald, T. L. J. Org. Chem. 1985, 50, 422. (c) Reetz, M. T.; Kessler, K. Ibid. 1985, 50, 5436. (d) Pikul, S.; Raczko, J.; Ankner, K.; Jurczak, J. J. Am. Chem. Soc. 1987, 109, 3981.
 (5) (a) Evans, D. A.; Nelson, J. V.; Taber, T. R. Top. Stereochem. 1983, 13, 1. (b) Reetz, M. T.; Jung, A. J. Am. Chem. Soc. 1983, 105, 4833. (c) Heathcock, C. H. In Asymmetric Synthesis; Morrison, J. D., Ed.; Academic Press: New York, 1984; Vol. 3B, p 111. (d) Wuts, P. G. M.; Walters, M. A. J. Org. Chem. 1984, 49, 4573.
 (6) (a) Danishefsky, S.; Larson, E.; Askin, D.; Kato, N. J. Am. Chem. Soc. 1985, 107, 1246. (b) Jurczak, J.; Bauer, T. Tetrahedron 1986, 42, 5045.
- rahedron 1986, 42, 5045.
- (7) Tourvé, D. Janssen Chim. Acta 1985, 3, 3.
 (8) Still, W. C.; Kahn, M.; Mitra, A. J. Org. Chem. 1978, 43, 2923.
 (9) Hann, M. M.; Sammes, P. G.; Kennewell, P. D.; Taylor, J. B. J. Chem. Soc., Perkin Trans. 1 1982, 307.
- (10) Ito, A.; Takahashi, R.; Baba, Y. Chem. Pharm. Bull. 1975, 23,
- (11) Rittle, K. E.; Homnick, C. F.; Ponticello, G. S.; Evans, B. E. J. Org. Chem. 1982, 47, 3016.
 (12) Lubell, W. D.; Rapoport, H. J. Am. Chem. Soc. 1987, 109,

- (13) Garner, P.; Park, J. M. J. Org. Chem. 1987, 52, 2361.
 (14) (a) Rich, D. H.; Sun, E. T.; Boparai, A. S. Ibid. 1978, 43, 3624.
 (b) Rich, D. H.; Sun, E. T.; Ulm, E. J. Med. Chem. 1980, 23,
- (15) Gol'ebiowski, A.; Jacobsson, U.; Jurczak, J. Tetrahedron 1987, 43, 3063.
- (16) Zemlicka, H.; Murata, M. J. Org. Chem. 1976, 41, 3317.
 (17) Khatri, H.; Stammer, C. H. J. Chem. Soc., Chem. Commun. **1979**, 79.
- (18) (a) Narita, M.; Otsuka, S.; Kobayashi, S.; Ohno, M.; Umezawa, Y.; Morishima, H.; Saito, S.; Takita, T.; Umezawa, H. Tetrahedron Lett. 1982, 23, 525. (b) Nishizawa, R.; Saino, T.; Takita, T.; Suda, H.; Aoyagi, T.; Umezawa, H. J. Med. Chem. 1977, 20, 510.
- (19) (a) Fehrentz, J. A.; Castro, B. Synthesis 1983, 676. (b) Fehrentz, J. A.; Heitz, A.; Castro, B.; Cazaubon, C.; Nisato, D. *FEBS Lett.* 1984, 167, 273.
 Nahm, S.; Weireb, S. *Tetrahedron Lett.* 1981, 22, 3815.
- (21) Seki, H.; Koga, K.; Yamada, S. Chem. Pharm. Bull. 1972, 20,
- (22) (a) Balenović, K.; Bregant, N.; Cerar, D.; Fles, D.; Jambresić,
 I. J. Org. Chem. 1953, 18, 297. (b) Balenivić, K.; Bregant, N.;
 Galijan, T.; Stefanac, Z.; Skarić, V. Ibid. 1956, 21, 115.
- Stanfield, C. F.; Parker, J. E.; Kanellis, P. Ibid. 1981, 46, 4799.
- (24) Hamada, Y.; Shioiri, T. Tetrahedron Lett. 1982, 23, 1193.
 (25) Luly, J. R.; Dellaria, J. F.; Plattner, J. J.; Soderquist, J. L.; Yi, N. J. Org. Chem. 1987, 52, 1487.
 (26) Evans, D. A.; Takacs, J. M.; Hurst, K. M. J. Am. Chem. Soc. 1027, 101, 101.
- **1979**, 101, 371.
- Hamada, Y.; Shioiri, T. Chem. Pharm. Bull. 1982, 30, 1921. Natarajan, S.; Condon, M. E.; Nakane, M.; Reid, J.; Gordon, E. M.; Cushman, D. W.; Ondetti, M. A. In Peptides, Synthesis-Structure-Function; Rich, D. H.; Gross, E., Eds.; Pierce Chemical Co.: Rockford, IL, 1981; p 429.
- (29) Ewing, W. R.; Harris, B. D.; Bhat, K. L.; Joullie, M. M. Tetrahedron 1986, 42, 2421.
- Szelke, M.; Hudson, D.; Sharpe, R.; MacIntyre, I.; Pickering, A. M. C. Molecular Endocrynology; Elsevier/North Holland Biomedical Press: Amsterdam, 1977; p 57.
- (31) Pyne, S. G.; Hensel, M. J.; Fuchs, P. L. J. Am. Chem. Soc. 1982, 104, 5719.
- (32) Stanfield, C. F.; Parker, J. E.; Kanellis, P. J. Org. Chem. 1981,
- (33) Ohfune, Y.; Nishio, H. Tetrahedron Lett. 1984, 25, 4133.
 (34) Andersson, L.; Wolfenden, R. Anal. Biochem. 1982, 124, 150. (35) Mulzer, J.; Angermann, A.; Schubert, B.; Seilz, C. J. Org.
- Chem. 1986, 51, 5294.
- (36) Ohfune, Y.; Kurokawa, N. Tetrahedron Lett. 1984, 25, 1071.
 (37) Münster, P.; Giannis, A.; Steglich, W.; Sandhoff, K. In Abstracts of the 4th European Carbohydrate Symposium; Lichtenthaler, F. W., Neff, K. H., Eds.; Gesellschaft Deutscher Chemiker: Frankfurt, 1987; p A-54.
 (38) Saito, S.; Bunya, N.; Inaba, M.; Moriwake, T.; Torii, S. Tetrahedron Lett. 1985, 26, 5309.

- Saito, S.; Bunya, N.; Inaba, M.; Moriwake, T.; Torii, S. Tetrahedron Lett. 1985, 26, 5309.

 Mori, K.; Iwasawa, H. Tetrahedron 1980, 36, 87.
 Saito, S.; Hasegawa, T.; Inaba, M.; Nishida, R.; Fujii, T.; Nomizu, S.; Moriwake, T. Chem. Lett. 1984, 1389.

 Hanson, G. J.; Lindberg, T. J. Org. Chem. 1985, 50, 5399.

 Thaisrivongs, S.; Pals, D. T.; Kroll, L. T.; Turner, S. R.; Han, F. S. J. Med. Chem. 1987, 30, 976.

- (43) Holladay, M. W.; Rich, D. H. Tetrahedron Lett. 1983, 24,
- (44) Fray, A. H.; Kaye, R. L.; Kleinman, E. F. J. Org. Chem. 1986, 51, 4828.
- (45) (a) Newman, H. J. Am. Chem. Soc. 1973, 95, 4098. (b) Saitoh, Y.; Moriyama, Y.; Takahashi, T.; Khuong-Huu, Q. Tetrahedron Lett. 1980, 21, 75.
- Tkaczuk, P.; Thornton, E. R. J. Org. Chem. 1981, 46, 4393.
- Mori, K.; Funaki, Y. Tetrahedron 1985, 41, 2379.
- (48) Evans, B. E.; Rittle, K. E.; Homnick, C. F.; Springer, J. P.; Hirshfield, J.; Veber, D. F. J. Org. Chem. 1985, 50, 4615.
- (49) Hanson, G. J.; Baran, J. S.; Lindberg, T. Tetrahedron Lett. 1986, 27, 3577.
- Rosenberg, S. H.; Plattner, J. J.; Woods, K. W.; Stein, H. H.; Marcotte, P. A.; Cohen, J.; Perun, T. J. J. Med. Chem. 1987,
- (51) Dellaria, J. F., Jr.; Maki, R. G. Tetrahedron Lett. 1986, 27,
- (52) (a) Kempf, D. J. J. Org. Chem. 1986, 51, 3921. (b) Kempf,
 D. J.; de Lara, E.; Stein, H. H.; Cohen, J.; Plattner, J. J. J. Med. Chem. 1987, 30, 1978.

- Med. Chem. 1987, 30, 1978.
 (53) Liu, W. S.; Glover, G. I. J. Org. Chem. 1978, 43, 754.
 (54) Weinreb, S. M.; Staib, R. R. Tetrahedron 1982, 38, 3087.
 (55) Danishefsky, S.; Kobayashi, S.; Kerwin, J. F., Jr. J. Org. Chem. 1982, 47, 1981.
 (56) Garner, P.; Ramakanth, S. Ibid. 1986, 51, 2609.
 (57) Anh, N. T.; Eisenstein, O. Nouv. J. Chim. 1977, 1, 61.
 (58) (a) Golyebiowski, A. Ph.D. Thesis, Institute of Organic Chemistry, Polish Academy of Sciences, Warszawa, 1987. (b) Golyebiowski. A.: Jurczak. J. Unnublished results.
- Gol'ebiowski, A.; Jurczak, J. Unpublished results.
 (59) Recent reviews: (a) Matsumoto, K; Sera, A.; Uchida, T. Synthesis 1985, 1. (b) Matsumoto, K.; Sera, A. Ibid. 1985, 999. (c) Jurcak, J. Physica 1986, 139B; 140B, 709.

- (60) Jurczak, J.; Gol/ebiowski, A.; Bauer, T. Synthesis 1985, 928.
 (61) Bestmann, H. J. Pure Appl. Chem. 1979, 51, 515.
 (62) (a) Vedejs, E.; Meier, G. P.; Snoble, K. A. J. Am. Chem. Soc. 1981, 103, 2823. (b) Schlosser, M.; Schaub, B. Ibid. 1982, 104, 5201 5821
- (63) (a) Kozikowski, A. P.; Nieduzak, T. R.; Springer, J. P. Tetrahedron Lett. 1986, 27, 819. (b) Kozikowski, A. P.; Konoike, T.; Nieduzak, T. R. J. Chem. Soc., Chem. Commun. 1986,
- 1350. (c) Kozikowski, A. P.; Nieduzak, T. R.; Konoike, T.; Springer, J. P. J. Am. Chem. Soc. 1987, 109, 5167. (a) Itaya, T.; Fujii, T.; Evidente, A.; Randazzo, G.; Surico, G.; Iacobellis, N. S. Tetrahedron Lett. 1986, 27, 6349. (b) Sodano, G.; Spinella, A. *Ibid*. 1986, 27, 2505.
- Komiyama, K.; Urano, Y.; Kobayashi, S.; Ohno, M. Ibid.
- 1987, 28, 3123. Hensel, M. J.; Fuchs, P. L. Synth. Commun. 1986, 16, 1285. (67) Kogen, H.; Nishi, T. J. Chem. Soc., Chem. Commun. 1987,
- (68) Spatola, A. F. In Chemistry and Biochemistry of Amino Acids; Weinstein, B., Ed.; M. Dekker: New York, 1983; Vol.
- Umezawa, H. Enzyme Eng. 1982, 6, 11.
- (70) Bolis, G.; Fung, A. K. L.; Greer, J.; Kleinert, H. D.; Marcotte, P. A.; Perun, T. J.; Plattner, J. J.; Stein, H. H. J. Med. Chem. 1987, 30, 1729.
- (a) Hann, M. M.; Sammes, P. G.; Kennewell, P. D.; Taylor, J. B. J. Chem. Soc., Chem. Commun. 1980, 234. (b) Johnson, R. L. J. Med. Chem. 1984, 27, 1351. (72) Zweifel, G.; Backlund, S. J. J. Am. Chem. Soc. 1977, 99, 3184. (73) Nomoto, K.; Ohfune, Y. J. Syn. Org. Chem. Jpn. 1982, 40,

- (74) Szelke, M.; Leckie, B.; Hallett, A.; Jones, D. M.; Sueiras, J.; Atrask, B.; Lever, A. F. Nature 1982, 299, 555.
 (75) (a) Salituro, F. G.; Agarwal, N.; Hofmann, T.; Rich, D. H. J. Med. Chem. 1987, 30, 286. (b) Luly, J. R.; Yi, N.; Soderquist, J.; Stein, H. H.; Cohen, J.; Perun, T. J.; Plattner, J. J. Ibid. 1987, 30, 1609. (c) Dellaria, J. F.; Maki, R. G.; Bopp, B. A.; Cohen, J.; Kleinert, H. D.; Luly, J. R.; Merits, I.; Plattner, J. J. Stein, H. H. Ibid. 1987, 30, 2127.
- J.; Stein, H. H. Ibid. 1987, 30, 2137.
 Umezawa, H.; Aoyagi, T.; Morishima, H.; Matsuzaki, H.; Hamada, M.; Takeuchi, T. J. Antibiot. 1970, 23, 259.
 Rich, D. H.; Sun, E. T.; Singh, J. Biochem. Biophys. Res. Commun. 1977, 74, 762.
- (78) Umezawa, H.; Aoyagi, T.; Suda, H.; Hamada, M.; Takeuchi, T. J. Antibiot. 1976, 29, 97.
- (79) Suda, H.; Takita, T.; Aoyagi, T.; Umezawa, H. Ibid. 1976, 29,
- (80) Garner, P. Tetrahedron Lett. 1984, 25, 5855
- (a) Kakinuma, K.; Otake, N.; Yonehara, H. *Ibid.* 1972, 2509. (b) Otake, N.; Ogita, T.; Seto, H.; Yonehara, H. *Experientia* 1981, 37, 926.
- (82) (a) Zamojski, A.; Banaszek, A.; Grynkiewicz, G. Adv. Carbohydr. Chem. Biochem. 1982, 40, 1. (b) Zamojski, A.; Grynkiewicz, G. In The Total Synthesis of Natural Products; ApSimon, J. W., Ed.; J. Wiley: Chichester, 1984; Vol. 6, p 141. (c) Chmielewski, M.; Jurczak, J. J. Carbohydr. Chem. 1987,

- (83) Gol'ebiowski, A.; Izdebski, J.; Jacobsson, U.; Jurczak, J. Heterocycles 1986, 24, 1205.
 (84) (a) Cooper, D. J.; Yudis, M. D.; Guthrie, R. D.; Prior, A. M. J. Chem. Soc. C 1971, 960. (b) Cooper, D. J.; Daniels, P. J. L.; Yudis, M. D.; Marigliano, H. M.; Guthrie, R. D.; Bukhari, S. T. K. Ibid. 1971, 3126. S. T. K. *Ibid.* 1971, 3126. (85) Gol/ębiowski, A.; Jacobsson, U.; Chmielewski, M.; Jurczak, J.
- (85) Gol'ębiowski, A.; Jacobsson, U.; Chmielewski, M.; Jurczak, J. Tetrahedron 1987, 43, 599.
 (86) (a) Nara, T.; Yamamoto, M.; Kawamoto, I.; Takayama, K.; Okachi, R.; Takasawa, S.; Sato, T.; Sato, S. J. Antibiot. 1977, 30, 533. (b) Okachi, R.; Takasawa, S.; Sato, T.; Sato, S.; Yamamoto, M.; Kawamoto, I.; Nara, T. Ibid. 1977, 30, 541.
 (87) (a) Mason, D. J.; DeBoer, C. Antimicrob. Agents Chemother. 1962, 544. (b) Herr, R. R.; Bergy, M. E. Ibid. 1962, 560.
 (88) Danishefsky, S.; Larson, E.; Springer, J. P. J. Am. Chem. Soc. 1985, 107, 1274.
 (89) Goto, T.; Toya, Y.; Ohgi, T.; Kondo, T. Tetrahedron Lett. 1982, 23, 1271.
 (90) Garner, P.; Ramakanth, S. J. Org. Chem. 1987, 52, 2629.
 (91) (a) Kawai, G.; Ikeda, Y. Biochim. Biophys. Acta 1983, 754, 243. (b) Karlsson, K. A.; Leffler, H.; Samuelsson, B. D. Ibid. 1979, 574, 79.

- 1979, 574, 79. (92) Haskell, T. H.; Bartz, Q. R. Antibiot. Ann. 1959, 1958/59,
- (93) Matsubara, S. Phytochem. 1980, 19, 2239.

- (94) Nagahara, T.; Kametani, T. Heterocycles 1987, 25, 729.
 (95) Matsunaga, H.; Sakamaki, T.; Nagaoka, H.; Yamada, Y. Tetrahedron. Lett. 1983, 24, 3009.
 (96) Takita, T.; Muraoka, Y.; Nakatani, T.; Fujii, A.; Umezawa, Y.; Naganawa, H.; Umezawa, H. J. Antibiot. 1978, 31, 801.
 (97) Takita, T.; Umezawa, Y.; Saito, S.; Morishima, H.; Naganawa, H.; Umezawa, H.; Tsuchiya, T.; Miyake, T.; Kageyama, S.; Umezawa, S.; Muraoka, Y.; Suzuki, M.; Otsuka, M.; Narita, M.; Kobayashi, S.; Ohno, M. Tetrahedron Lett. 1982, 23, 521.
 (98) Stork, G.; Nakamura, E. J. Am. Chem. Soc. 1983, 105, 5510.
 (99) (a) Reetz, M. T.; Drewes, M. W.; Schmitz, A. Angew. Chem., Int. Ed. Engl. 1987, 26, 1141. (b) Dondoni, A.; Fantin, G.; Fogagnolo, M.; Medici, A. J. Chem. Soc., Chem. Commun. 1988, 10.
- Fogagnolo, M.; Medici, A. J. Chem. Soc., Chem. Commun. 1988, 10.

 (100) (a) Shono, T.; Kise, N. Chem. Lett. 1987, 697. (b) Kano, S.; Yokomatsu, T.; Iwasawa, H.; Shibuya, S. Ibid. 1987, 1531. (c) Moriwake, T.; Hamano, S.; Saito, S.; Torii, S. Ibid. 1987, 2085. (d) Andrew, R. G.; Conrow, R. E.; Elliot, J. D.; Johnson, W. S.; Ramezani, S. Tetrahedron Lett. 1987, 27, 6535. (e) Bealieu, P. L.; Schiller, P. W. Tetrahedron Lett. 1988, 28, 2019. (f) Devant, R. M.; Radunz, H. E. Tetrahedron Lett. 1988, 28, 2307. (g) Beckett, R. P.; Davies, S. G. J. Chem. Soc., Chem. Commun. 1988, 160. (h) Garner, P.; Park, M. J. J. Org. Chem., in press. (i) Garner, P.; Malecki, E.; Park, M. J. Ibid., submitted for publication. Ibid., submitted for publication.