

Optimized Separation of Nine Xanthones by Microemulsion Electrokinetic Chromatography

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Abstract: A microemulsion electrokinetic chromatography method has been firstly used for the separation of the therapeutically important xanthones from *Securidaca inappendiculata*. The separation of the nine xanthones was systematically optimized with respect to pH, composition of microemulsion, addition of cyclodextrins, applied voltage and column temperature. Baseline separation was successfully achieved for the nine xanthones, which was also compared with that by micellar electrokinetic chromatography.

Keywords: Separation, xanthones, *Securidaca inappendiculata*, microemulsion electrokinetic chromatography, micellar electrokinetic chromatography.

Securidaca inappendiculata Hassk. is a traditional Chinese herbal medicine. Pharmacological investigations have shown that the xanthones, as main components accumulated in *S. inappendiculata* have many bioactivities, such as monoamine oxidase inhibition, antitumor activity, cytotoxicity, antibacterial activity, antifungal activity, anti-inflammatory properties, antioxidant activity and tuberculoatatic activity, *etc.*¹. In our previous work, the xanthones have been separated successfully by capillary zone electrophoresis (CZE)², and the separation mechanism by CZE has been described³.

Microemulsion electrokinetic chromatography (MEEKC) is a special variant of electrokinetic chromatography (EKC), whose separation principle is based on the different partition of solutes between the pseudostationary phase and buffers, and MEEKC shows a high solubilization capacity and separation efficiency^{4,5}. In this work, nine xanthones were firstly resolved by MEEKC, which gave very satisfactory results. The MEEKC separation was compared with that by MEKC.

Experimental

All separations were performed on an Agilent 3D CE system with air-cooling and a diode array detector (Agilent Technologies, Waldbronn, Germany). A 40 cm×50 μm I.D. fused silica capillary (Ruifeng Inc., Hebei, China) was used. The UV detection wavelength was set at 265 nm. Samples were injected into the capillary at 5000 Pa for 10 seconds.

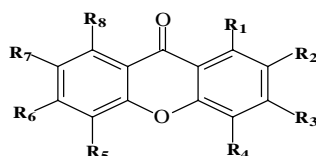
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The capillary was conditioned daily by washing first with 0.5 mol/L sodium hydroxide (10 minutes), then with water (10 minutes) and finally with the running buffer (15 minutes). Between consecutive analysis, the capillary was flushed with 0.5 mol/L sodium hydroxide (1 minute), then with water (2 minutes) and finally with the running buffer (3 minutes) in order to improve the migration time and peak-shape reproducibility.

The xanthenes (for their structural formula, refer to **Table 1**) were provided by Institute of Medicinal Plant Development (Beijing, China). Pure water prepared by Milli-Q system (Millipore, Bedford, MA, USA) was used for all buffer solutions. Sulfated β -CD and β -CD were kindly presented by Bioanalytical System Inc (West Lafayette, USA). All other chemicals were of analytical-reagent grade.

The running buffers for MEEKC analysis were adjusted to the desired pH with 1 mol/L NaOH. All the buffers were filtered through a 0.45 μ m membrane filter and degassed by ultrasonication for approximately 10 minutes before use. A standard solution of *ca.* 20 ppm of each xanthone was prepared in methanol, degassed in an ultrasonic bath and filtered through a 0.45 μ m membrane filter.

Table 1 Structural formula of the xanthenes



Compd.	Substituted groups							
	R ₁	R ₂	R ₃	R ₄	R ₅	R ₆	R ₇	R ₈
1	HO	H	HO	MeO	H	H	HO	H
2	HO	MeO	HO	H	H	H	HO	H
3	HO	MeO	MeO	H	H	HO	HO	H
4	HO	H	HO	H	H	H	HO	H
5	MeO	HO	H	H	H	H	HO	H
6	HO	MeO	MeO	H	H	H	HO	H
7	MeO	H	H	MeO	H	H	HO	H
8	HO	H	H	MeO	H	H	HO	H
9	H	H	MeO	HO	H	H	MeO	H

Results and Discussion

The pH, borate concentration and composition of microemulsion in the MEEKC analysis were optimized, and the optimum pH and borate concentration were 9.5 and 50 mmol/L, respectively, under 20 kV applied voltage and 35°C temperature. The composition of microemulsion was 80 mmol/L heptane, 120 mmol/L sodium dodecyl sulfate (SDS) and 10% (v/v) *n*-butanol, respectively. The separation is indicated in **Figure 1**.

Optimized Separation of Nine Xanthenes by Microemulsion Electrokinetic Chromatography 879

Five millimoles per liter β -CD and 6 mmol/L sulfated β -CD were evaluated as additives for optimizing separation, the effect of which on CE selectivity was studied at the conditions described above. The experiment shows that sulfated β -CD has better selectivity than β -CD for the MEEKC separation, as indicated in **Figures 2** and **3**. Sulfated β -CD is a counter-migrating complexing agent and combines the properties of CD and a surfactant by providing a hydrophobic cavity and a negative charge. Analytes will form inclusion complexes with sulfated β -CD whose electrophoretic mobilities are in direction opposite to the electro-osmotic flow. What is more, surfactant can result in electric and hydrogen bond effects with the phenolic hydroxyl groups of the xanthenes. The inclusive effects are different because of the different amount and position of the phenolic hydroxyl groups of the xanthenes, which induces different electric and hydrogen bond effect with sulfated β -CD. So adding sulfated β -CD results in different mobilities of the xanthenes.

Figure 1 Chromatogram of nine xanthenes by MEEKC without the addition of CD

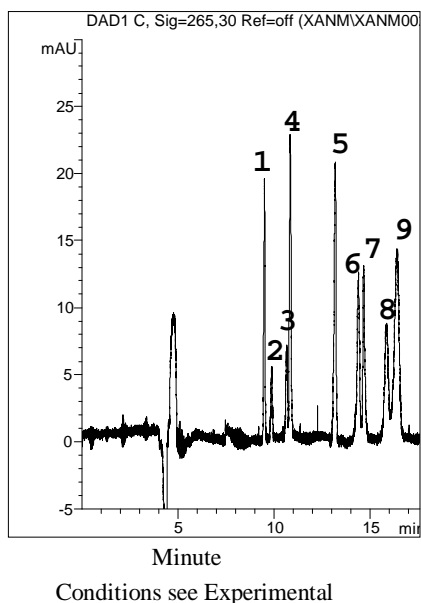
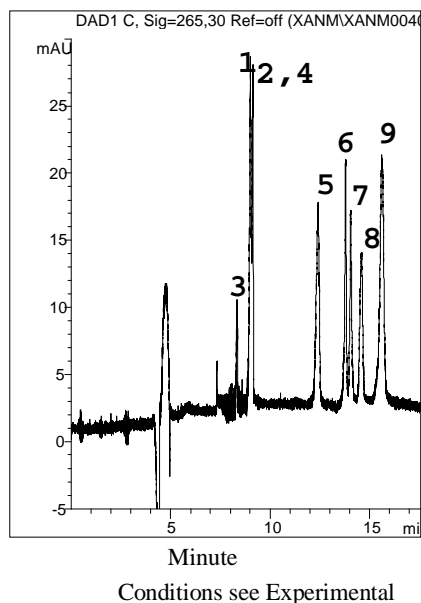


Figure 2 Chromatogram of nine xanthenes by β -CD modified MEEKC

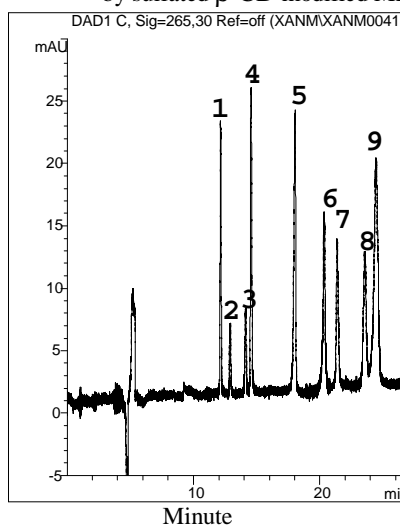


Finally, the effects of applied voltage (15-30 kV) and temperature (30-45°C) on the separation were also studied in the optimum running buffer above. The optimum voltage and temperature were found to be 20 kV and 35°C, respectively, which combined sufficient resolution with a moderate analysis time.

The repeatability of the migration time and the peak area of each xanthone under the optimum condition was very good with relatively standard deviation of less than 5%, and the theoretical plate number is in the range of 90120-252108 m^{-1} . The experiment indicated that the MEEKC analysis has better selectivity than MEKC. The MEKC

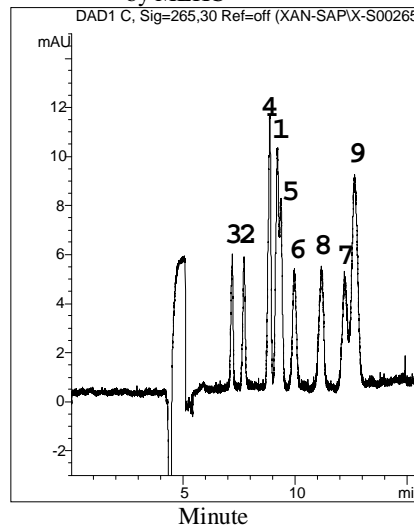
separation is shown in **Figure 4** (condition: 100 mmol/L borate, 60 mmol/L SDS, 5 mmol/L β -CD, pH 10.5, 265 nm UV detection, 20 kV applied voltage and 35°C column temperature).

Figure 3 Chromatogram of nine xanthenes by sulfated β -CD modified MEEKC



Conditions see Experimental.

Figure 4 Chromatogram of nine xanthenes by MEKC



Conditions see Experimental.

Conclusion

This work demonstrates that the nine xanthenes can be baseline separated by CE with satisfactory migration time and peak area repeatability. Compared with MEKC method, MEEKC has higher separation efficiency and selectivity.

Acknowledgments

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