

Synthesis of New Benzoimidazole Derivatives

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Abstract: The reactions of cryptotanshinone and tanshinone IIA with cadaverine and putrescine were investigated. Six new compounds, four with imidazole functional groups and two with oxazole groups, were obtained. The possible reaction mechanism was proposed.

Keywords: Cryptotanshinone, tanshinone IIA, putrescine, cadaverine, biogenic bisamines.

Tanshinones are naturally occurring diterpenoids isolated from *Salvia miltiorrhiza* Bunge. This type of compounds has shown comprehensive pharmacological activities such as enzyme inhibitory activity and cytotoxicity^{1,2}. In this paper, the reactions of cryptotanshinone **1** and tanshinone IIA **2** with biogenic bisamines were investigated. Biogenic bisamines such as cadaverine (1, 5-diamino pentane) and putrescine (1, 4-diamino butane) were considered to be involved in the process of cell multiplication and regulation^{3,4}, we believe that the current work may provide some evidence for the mechanisms on enzymatic inhibitory activities and cytotoxicity of tanshinones.

The reaction of **1** with putrescine or cadaverine gave major products of **3** or **4**, respectively (**Scheme 1**). FAB-MS and elemental analysis of **4** suggested a formula of C₂₄H₂₈N₂O. ¹³C NMR and IR data indicated that **4** had no *O*-quinone carbon, because there was no absorption signal of quinone function group in the UV spectrum, indicating that the *O*-quinone moiety was changed during reaction. NMR data showed that the product contained three methyl, eight methylene groups and three methenyl. An additional aromatic quaternary carbon signal (δ 152.0) appeared at lower field that could be assigned to C-1'. Two protons with an AMX type splitting at δ 4.38 and δ 4.87 could be assigned to CH₂-16, which was made diastereotopic by the chirality of CH-15. Signals of aromatic protons (CH-6 and CH-7) of **4** appeared at δ 7.46 and δ 7.91 as a sharp AB-quartet. H-H COSY spectrum showed that **4** contained a -CH₂CH₂CH₂CH₂- moiety which was not found in **1**, indicating that the fragment was probably derived from cadaverine. In NOE spectrum, irradiating CH₃-17 (δ 1.53) resulted in the enhancement of CH-15 and CH₂-16, and irradiating one proton of CH₂-1 (δ 3.20) resulted in the enhancement of CH-5' (δ 4.41), indicating that CH₂-1 and CH₂-5' were spatially

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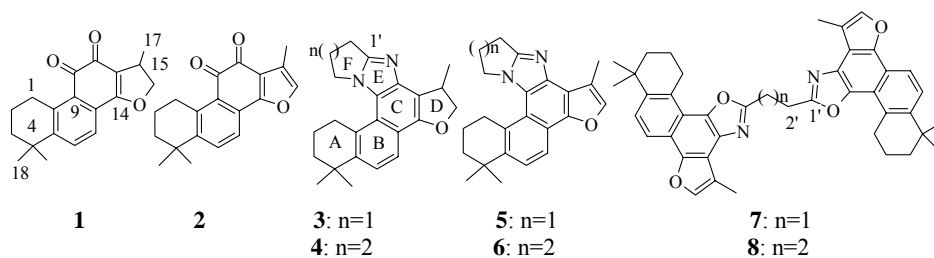
neighboring protons. The CH₂-1 protons were anisochronic and their ¹H NMR signals appeared at δ 3.18-3.23 and δ 3.27-3.32 as multiple peaks. The signals differ from those of **1**, which were triplet peaks (δ 3.22, *J* = 7 Hz)⁵. Similarly, the two proton signals of CH₂-5' appeared at δ 4.30-4.34 and δ 4.39-4.44, splitting as multiple peaks, respectively. According to all of the spectral data⁶, product **4** could be assigned as 2, 7, 7-trimethyl-2, 3, 7, 8, 9, 10, 11, 12, 13, 14-decahydro [furo[2, 3-*d*]phenanthro][1, 2-*d*]pyrido[1, 2-*a*]imidazole.

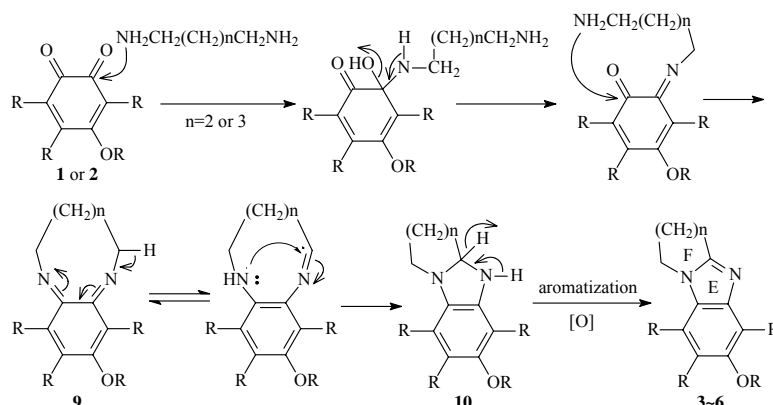
Similar to **4**, the product **3** could be assigned as 2, 7, 7-trimethyl-2, 3, 7, 8, 9, 10, 11, 12, 13-enneahydro[furo[2, 3-*d*]phenanthro][1, 2-*d*]pyrrolo[1, 2-*a*]imidazole according to its spectral data⁶. The difference between the structure of compound **3** and **4** was the F ring. The pyrrolidine moiety was formed from the reaction of **1** with putrescine, while the reaction of cadaverine with **1** gave F ring as piperidine moiety. As can be seen from its structure, the spatial distance between the CH₂-1 and CH₂-4' in **4** was longer than that in **3**, this in turn resulted that **3** has less sterical hindrance comparing to **4**. Consequently, the ¹H NMR signals of CH₂-1 appeared at δ 3.31 as multiple peaks, and those of CH₂-4' appeared at δ 4.66.

The reaction of **2** with cadaverine gave two major products of **6** and **8** as shown in **Scheme 1**. FAB-MS and elemental analysis showed that the molecular formula of product **6** was C₂₄H₂₆N₂O. According to the spectral data⁶, similar to compound **4**, product **6** was determined to be 2, 7, 7-trimethyl-7, 8, 9, 10, 11, 12, 13, 14-octahydro [furo[2, 3-*d*]phenanthro][1, 2-*d*]pyrido[1, 2-*a*]imidazole.

FAB-MS and elemental analysis indicated that the molecular formula of **8** was C₄₃H₄₂N₂O₄. ¹H NMR showed that the structure of **8**¹⁰ was probably symmetric. The signal area of the CH₂-2' methylene groups appeared as four protons, while the CH₂-3' signal at δ 2.71 appeared as two protons and was quintuplet splitting peaks (*J* = 7 Hz). In ¹H NMR spectrum, product **8** had AX₃ style long-range coupling, δ 2.35 (d, *J*=1 Hz) and δ 7.40 (q, *J*=1 Hz), character of the signals of β-methyl furan moiety. The singlet peak at δ 1.37 could be assigned as the signal of the geminal dimethyl groups in **8**. Comparing its NMR data with that of compound **2**, it could be confirmed that compound **8** contained the same molecular framework with compound **2**. According to all of spectral data, compound **8** could be assigned as 1, 3-di(4, 9, 9-trimethyl-9, 10, 11, 12-tetrahydro-1, 6-dioxo-3-azadicyclopenta[*a, c*]phenanthren-2-yl)-propane⁷.

Scheme 1



Scheme 2 Possible mechanism of the formation of imidazole derivatives

Two major products **5** and **7**¹⁰ were obtained from the reaction of **2** with putrescine. Their structures were similar to that of products **6** and **8**. According to the spectral data, compounds **5** and **7** could be assigned as 2, 7, 7-trimethyl-7, 8, 9, 10, 11, 12, 13-hepta-hydro[furo[2, 3-*d*]phenanthro][1, 2-*d*]pyrrolo[1, 2-*a*]imidazole and 1, 2-di(4, 9, 9-trimethyl-9, 10, 11, 12-tetrahydro-1, 6-dioxo-3-azadicyclopenta[*a, c*]phenanthren-2-yl)-ethane, respectively.

It was noticeable that every reaction of compound **1** or **2** with bisamines gave imidazole derivative products. The possible reaction mechanism was proposed in **Scheme 2**. A nucleophilic reaction took place between tanshinone and amine. One of the amino groups in bisamine attacked one *O*-quinone carbonyl groups, and a quinone-imine intermediate was formed and one equivalent H₂O was eliminated. Then, another amino group of the bisamine attacked another carbonyl group of the tanshinone. The rearrangement of intermediate **9** to the aromatic Schiff base might be spontaneous because of the stability of the latter⁸. Intermolecular additional reactions took place in the Schiff base intermediate, and resulted in intermediate **10**, which, in the presence of an oxidant such as tanshinone or air⁵, underwent aromatization to give the imidazole product⁹.

Two major products were isolated from the reaction of tanshinone IIA with biogenic bisamines. One of the two products was oxazole derivative. The formation of oxazole product might be through a previously proposed pathway^{5,7}.

In conclusion, the reaction of tanshinones with biogenic bisamines revealed a possible mechanism of the enzymatic inhibitory activities and cytotoxicity of tanshinones. The reaction offers a potential strategy for the preparation of quinine derivatives bearing imidazole functional groups. Modification of the reaction conditions to get higher yields of functionalized quinines is currently underway.

Acknowledgments

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6. The data of products **3**, **4**, **5**, **6**.
Product **3**: light yellow needle crystals, yield 15%. C₂₃H₂₆N₂O, calcd: C, 79.73; H, 7.56; N, 8.09; O, 4.62; found: C, 79.69; H, 7.60; N, 8.02. m.p. 227-230 °C. FAB-MS *m/z* (rel. int.): 347 [M+1]⁺ (95), 346 (100). UV λ (EtOH): 348.0, 333.0, 258.0, 233.0 nm. IR ν (KBr): 3060, 2957, 2928, 2868, 1543, 1513, 1455, 1397, 1295 cm⁻¹. ¹H NMR (500 MHz, Acetone, δ ppm) 1.36 (s, 3H), 1.38 (s, 3H), 1.53 (d, 3H, *J* 6.5Hz), 1.81 (m, 2H), 1.83 (m, 2H), 2.65 (m, 2H), 3.01 (m, 2H), 3.31 (m, 2H), 4.02 (m, 1H), 4.30 (dd, AMX, 1H, *J* 6.3, 8.8Hz), 4.66 (m, 2H), 4.87 (t, AMX, 1H, *J* 8.8Hz), 7.45 (d, AB, 1H, *J* 9Hz), 7.85 (d, AB, 1H, *J* 8.5Hz). ¹³C NMR-DEPT (500 MHz, Acetone, δ ppm) 19.8q, 21.0t, 24.0t, 27.3t, 32.6q, 34.1t, 35.5s, 38.2d, 40.0t, 52.6t, 79.5t, 116.8s, 117.6s, 121.2d, 123.5d, 125.0s, 129.8s, 142.9s, 145.3s, 152.3s, 161.6s. The structure was confirmed by HMQC and HMBC. Product **4**: light yellow needle crystals, yield 12%, C₂₄H₂₈N₂O, calcd: C, 79.96; H, 7.83; N, 7.77; O, 4.44; found: C, 79.91; H, 7.85; N, 7.70. m.p. 103-104.5 °C. FAB-MS *m/z* (rel. int.): 361 [M+1]⁺ (100). UV λ (EtOH): 352.0, 263.0, 236.0, 203.0 nm. IR ν (KBr): 3067, 2927, 2859, 1461, 1402, 1118, 824 cm⁻¹. ¹H NMR (500 MHz, CDCl₃, δ ppm) 1.37 (s, 3H), 1.42 (s, 3H), 1.53 (d, 3H, *J* 7Hz), 1.76 (m, 2H), 1.82 (m, 2H), 1.93 (m, 2H), 2.09 (m, 2H), 3.18-3.23 (m, 1H), 3.28 (t, 2H, *J* 7Hz), 3.27-3.32 (m, 1H), 4.09 (m, 1H), 4.30-4.34 (m, 1H), 4.38 (dd, AMX, 1H, *J* 5.3, 8.3Hz), 4.39-4.44 (m, 1H), 4.87 (t, 1H, *J* 8.8Hz), 7.46 (d, AB, 1H, *J* 9Hz), 7.91 (d, AB, 1H, *J* 8.5Hz). ¹³C NMR-DEPT (500 MHz, CDCl₃, δ ppm) 20.0q, 20.2t, 21.4t, 24.7t, 25.6t, 32.7q, 34.3t, 35.2s, 36.9d, 40.1t, 51.3t, 79.3t, 115.2s, 117.4s, 120.4d, 122.9s, 123.6d, 126.6s, 129.5s, 138.9s, 142.9s, 151.9s, 152.0s. The structure was confirmed by H-H COSY, NOE, HMQC and HMBC together. Product **5**: colorless needle crystals, yield 14%, C₂₃H₂₄N₂O, calcd: C, 80.20; H, 7.02; N, 8.13; O, 4.64; found: C, 80.15; H, 7.09; N, 8.07. m.p. 282.0-284.0 °C. FAB-MS *m/z* (rel. int.): 345 [M+1]⁺ (100). UV λ (EtOH): 266.0, 203.0 nm. IR ν (KBr): 2925, 2854, 1543, 1513, 1458, 1375 cm⁻¹. ¹H NMR (500 MHz, CDCl₃, δ ppm) 1.34 (s, 6H), 1.74 (m, 2H), 1.79 (m, 2H), 2.57 (d, 3H, *J* 1Hz), 2.61 (m, 2H), 3.17 (t, 2H, *J* 7.8Hz), 3.25 (t, 2H, *J* 5.5Hz), 4.58 (t, 2H, *J* 7Hz), 7.44 (q, 1H, *J* 1Hz), 7.49 (d, AB, 1H, *J* 8.5Hz), 8.16 (d, AB, 1H, *J* 9Hz). ¹³C NMR-DEPT (500 MHz, CDCl₃, δ ppm) 9.9q, 20.6t, 23.9t, 27.1t, 32.5q, 33.8t, 35.0s, 39.3t, 52.4t, 116.2s, 116.7s, 118.0s, 119.2d, 120.4s, 124.4d, 124.7s, 127.3s, 129.3s, 140.6d, 142.3s, 149.3s, 159.4s. Product **6**: colorless solid, yield 19%, C₂₄H₂₆N₂O, calcd: C, 80.41; H, 7.31; N, 7.81; O, 4.46; found: C, 80.43; H, 7.35; N, 7.76. m.p. 126-128 °C. FAB-MS *m/z* (rel. int.): 359 [M+1]⁺ (20), 154 (100). UV λ (EtOH): 348.0, 332.0, 269.0, 205.0 nm. IR ν (KBr): 3061, 2929, 2860, 1607, 1499, 1457, 1329 cm⁻¹. ¹H NMR (500 MHz, CDCl₃, δ ppm) 1.41 (s, 6H), 1.72 (m, 2H), 1.80 (m, 2H), 1.84 (m, 2H), 2.13 (m, 2H), 2.64 (d, 3H, *J* 1.5Hz), 3.27 (t, 2H, *J* 5.8Hz), 3.35 (t, 2H, *J* 7Hz), 4.38 (t, 2H, *J* 5.5Hz), 7.50 (q, br, 1H, *J* 1.5Hz), 7.55 (d, AB, 1H, *J* 8.5Hz), 8.21 (d, AB, 1H, *J* 8Hz). ¹³C NMR-DEPT (500 MHz, CDCl₃, δ ppm) 9.9q, 20.2t, 21.5t, 24.7t, 25.4t, 32.8q, 34.1t, 35.1s, 40.2t, 51.3t, 116.2s, 116.7s, 118.5s, 119.0d, 120.7s, 124.0s, 124.4d, 127.5s, 130.2s, 140.6d, 142.4s, 149.2s, 151.4s.
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