

## Two Phaeophytin Type Analogues from Marine Sponge *Dysidea sp.*

Peng Fei JIN<sup>1,3</sup>, Zhi Wei DENG<sup>2</sup>, Yue Hu PEI<sup>3</sup>, Wen Han LIN<sup>1\*</sup>

<sup>1</sup>State Key Laboratory of Natural and Biomimetic Drugs, Peking University, Beijing 100083

<sup>2</sup>The Chemical and Test Center, Beijing Normal University, Beijing 100073

<sup>3</sup>College of Traditional Chinese Medicine, Shenyang Pharmaceutical University, Shenyang 110016

**Abstract:** A new compound named 13b (*S*)-hydroxy-17c-ethoxyphaeophorbide a (**2**) together with a known compound 17c-ethoxyphaeophorbide a (**1**) were isolated from marine sponge *Dysidea sp.* collected in South China sea. The structures were elucidated by spectroscopic analysis as well as comparison with those reported in literatures.

**Keywords:** Marine sponge, *Dysidea sp.*, 17c-ethoxyphaeophorbide a, 13b(*S*)-hydroxy-17c-ethoxyphaeophorbide a.

Phaeophytins and related compounds widely distributed in green plants including marine alga, silkworm, photosynthetic bacteria<sup>1-4</sup>. They play an important role in the transmission and primary light conversion events in photosynthesis. Hitherto a few phaeophytins have been discovered in marine sponges, only *Corallistes sp.* and *Darwinella oxeata* have been reported to contain phaeorphyrin type compounds<sup>5-7</sup>. Previous works revealed that phaeophytins possess potent cytotoxic activities against several solid tumor cell lines<sup>8</sup> and antioxidant activities<sup>9-10</sup>. In the continuous investigation of the bioactive natural products from marine organisms, the marine sponge *Dysidea sp.* was collected from Hainan island, South China sea. The MeOH extract of sponge (560 g) was concentrated in vacuum and then partitioned between CH<sub>2</sub>Cl<sub>2</sub> and H<sub>2</sub>O. The CH<sub>2</sub>Cl<sub>2</sub> fraction was subjected to silica gel column chromatography, eluting with a gradient (petroleum ether-acetone) to yield five fractions. The fractions were tested on tumor cell lines (HL60, PC-3MIE8, BGC-823, Bel-7402, Hela, MDA-MB-435), of which one fraction showed significant cytotoxicities against the selected cell lines. Subsequent separation of the active fraction by repeated silica gel column chromatography and followed by semi-preparative HPLC led to yield compound **1** (3.5 mg) and compound **2** (1.2 mg).

Compound **1** was identified as 17c-ethoxyphaeophorbide a by comparison of its spectral data and physical properties with those reported in literature<sup>10</sup>.

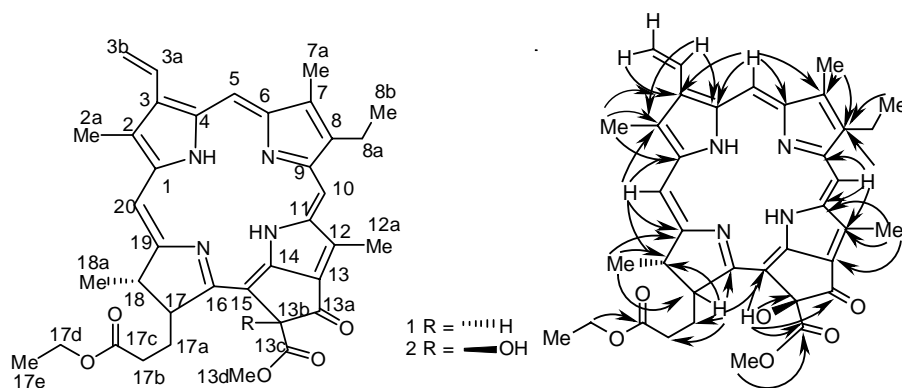
Compound **2**, [ $\alpha$ ]<sub>D</sub><sup>25</sup>-6.51 (*c* 0.2, CHCl<sub>3</sub>), was isolated as a dark brown amorphous solid, and its molecular formula C<sub>37</sub>H<sub>41</sub>N<sub>4</sub>O<sub>6</sub> was established by HRFABMS (*m/z* 637.3026 [M+H]<sup>+</sup>, calcd. for C<sub>37</sub>H<sub>40</sub>N<sub>4</sub>O<sub>6</sub>, 637.3026). IR absorptions at 3429, 1737, 1706 and 1616

\* E-mail: whlin@bjmu.edu.cn

$\text{cm}^{-1}$  suggested the presence of hydroxyl, carbonyl and vinyl groups. Its  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (**Table 1**) closely resembled those of compound **1**. In  $^1\text{H}$  NMR spectrum, there are seven methyls at  $\delta$ 3.45 (s, H-2a), 3.30 (s, H-7a), 1.71 (t,  $J=7.5$  Hz, H-8b), 3.66 (s, H-12a), 3.76 (s, OMe), 1.14 (t,  $J=7.0$  Hz, H-17e) and 1.63 (d,  $J=7.0$  Hz, H-18a); three olefinic singlets at  $\delta$ 9.51 (s, H-5), 9.65 (s, H-10), and 8.66 (s, H-20); one mono-substituted vinyl group at  $\delta$ 8.06 (dd,  $J=17.5, 11.5$  Hz, H-3a), 6.31 (brd,  $J=17.5$  Hz, H-3b1) and 6.20 (brd,

**Table 1**  $^1\text{H}$  and  $^{13}\text{C}$  NMR of compounds **1** and **2** (500 MHz, in  $\text{CDCl}_3$ ,  $\delta$  ppm)

No.	<b>1</b>		<b>2</b>	
	$\delta_{\text{H}}$	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{C}}$
1		142.1		144.8
2		133.0		131.9
2a	3.41, s	12.13	3.45, s	12.5
3		136.3		137.3
3a	8.01, dd (17.5, 11.5)	129.1	8.06, dd (17.5, 11.5)	129.4
3b1	6.30, brd (17.5)	122.8	6.31, brd (17.5)	123.8
3b2	6.19, brd (11.5)		6.20, brd (11.5)	
4		136.2		137.2
5	9.41, s	97.6	9.51, s	98.4
6		155.0		155.6
7		136.5		137.2
7a	3.24, s	11.27	3.30, s	11.7
8		145.2		145.4
8a	3.69, q (7.5)	19.5	3.72, q (7.5)	20.0
8b	1.72, t (7.5)	17.4	1.71, t (7.5)	17.7
9		151.7		151.9
10	9.55, s	104.5	9.65, s	104.8
11		137.9		138.4
12		129.1		129.4
12a	3.70, s	12.1	3.66, s	12.8
13		129.1		127.1
13a		189.6		192.2
13b	6.27, s	64.7		89.5
13c		169.6		173.9
13d	3.88, s	52.9	3.76, s	53.9
14		149.7		150.0
15		105.2		104.9
16		161.3		163.1
17	4.22, m	51.1	4.16, m	52.5
17a	2.62, 2.34, m	29.8	2.29, m	31.6
17b	2.48, 2.18, m	31.2	2.92, 2.52, m	32.0
17c		172.9		173.9
17d	4.02, q (7.0)	60.5	4.10, q (7.0)	63.3
17e	1.12, t (7.0)	14.1	1.14, t (7.0)	14.5
18	4.48, m	50.1	4.50, m	50.9
18a	1.82, d (7.0)	23.1	1.63, d (7.0)	23.1
19		172.2		173.3
20	8.58, s	93.2	8.66, s	95.1
13b-OH			5.55, brs	

**Figure 1** The structures of **1** and **2** and main HMBC correlation of **2**

$J=11.5$  Hz, H-3b2), as well as one  $D_2O$  exchangeable signal at  $\delta$  5.55 (brs.). Compound **2** differed from **1** in the position C-13b, where a proton H-13b ( $\delta$  6.27, s) in **1** was replaced by a hydroxyl group in **2**, which was supported by a quaternary carbon at  $\delta$  89.5 (s) in the  $^{13}C$  NMR spectrum of **2** instead of a methine carbon at  $\delta$  64.7 (d, C-13b) of **1**. In the HMBC spectrum of **2** (see **Figure 1**), the long range correlations of the exchangeable proton ( $\delta$  5.55, brs) with C-13a ( $\delta$  192.2, s), C-13b ( $\delta$  89.5, s), C-13c ( $\delta$  173.9, s) and C-15 (104.9, s) further confirmed the location of hydroxyl group. The configuration at chiral center C-13b in **2** was determined as *S* due to the up-field chemical shift of H-17 ( $\delta$  4.16, m)<sup>11,12</sup>. Accordingly, the structure of compound **2** was identified as 13b(*S*)-hydroxy-17c-ethoxyphaeophorbide a.

### Acknowledgments

The work was supported by grants from National High Technology Development Project (863 project) (No. 2001AA620403 and 2002AA217081), and NNSFC (30171106, 40176038).

### References

1. M. S. Buchanan, H. Toshihiro, Y. Asakawa, *Phytochemistry*, **1996**, 41(5), 1373.
2. K. M. Smith, J. F. Unsworth, *Tetrahedron*, **1975**, 31, 367.
3. A. Matsuo, K. Ono, K. Hamasaki, *et al.*, *Phytochemistry*, **1996**, 42(2), 427.
4. K. M. Smith, D. A. Goff, *Tetrahedron Lett.*, **1981**, 22(48), 4873.
5. M. D. Ambrosio, A. Guerriero, C. Debitus, *et al.*, *Helv. Chim. Acta*, **1989**, 72, 1451.
6. P. Karuso, P. R. Bergquist, J. S. Buckleton, *et al.*, *Tetrahedron Lett.*, **1986**, 27(19), 2177.
7. M. D. Ambrosio, A. Guerriero, C. Debitus, *et al.*, *Helv. Chim. Acta*, **1993**, 76, 1489.
8. K. Sakata, K. Yamamoto, H. Ishikawa, *et al.*, *Tetrahedron Lett.*, **1990**, 31(8), 1165.
9. H. H. Cheng, H. K. Wang, J. Ito, *et al.*, *J. Nat. Prod.*, **2001**, 64, 915.
10. S. L. Schwikkard, D. A. Mulholland, A. Hutchings, *Phytochemistry*, **1998**, 49(8), 2391.
11. Y. Nakatani, G. Ourisson, J. P. Beck, *Chem. Pharm. Bull.*, **1981**, 29(8), 2261.
12. H. Worf, H. J. Brockmann, H. Biere, *et al.*, *Liebigs Ann. Chem.*, **1967**, 704, 208.

Received 19 December, 2003