

## EQUILIBRATION OF CHLORIDE AND PYRUVATE DISTRIBUTIONS BETWEEN LIVER MITOCHONDRIA AND MEDIUM MEDIATED BY ORGANO-TIN SALTS

E.J. HARRIS, J.A. BANGHAM and B. ZUKOVIC

*Department of Biophysics, University College London, London WC1E 6BT, England*

Received 15 November 1972

### 1. Introduction

Rat liver mitochondria are normally impermeable to chloride [1] so when separated from suspensions they carry about the same volume of  $\text{Cl}^-$ -accessible water as sucrose-accessible water [2]. In contrast to  $\text{Cl}^-$ , most organic anions are internally accumulated to concentrations exceeding those outside, and such penetrant anions intercompete for occupation of the interior [3, 4].

Manger [5] observed that triethyl tin sulphate, added to a suspension in a 150 mM KCl based medium lessened the contents of organic anions. Studies of the swelling of liposomes, erythrocytes and mitochondria in response to organo-tin compounds led Selwyn et al. [6] to conclude that a  $\text{Cl}^-$ -for- $\text{OH}^-$  exchange was mediated by the compounds. Taken together the two findings point to the organo-tin mediated  $\text{Cl}^-$  entry providing another permeant anion to compete with the organic anions accumulated in the mitochondrial interior. This displacement of substrate anions is one possible explanation of the inhibition of respiration obtained by additions of organo-tin salts in  $\text{Cl}^-$ -containing media [7]. The object of this work was to obtain direct evidence for the  $\text{Cl}^-$  entry and to compare its extent with that of a penetrant substrate (pyruvate). It has been observed [8] that the organo-tins inhibit phosphorylation in  $\text{Cl}^-$ -free media; another action unrelated to  $\text{Cl}^-$  movement was sought, and this is also described. A model system was employed to confirm that the  $\text{Cl}^-$ -for- $\text{OH}^-$  exchange will take place across an oil layer.

### 2. Methods

Rat liver mitochondria were prepared as described before [9], and finally suspended in 0.25 M sucrose. The anion distribution studies were made at 20° with 3–5 mg protein/ml suspended in 150 mM potassium methane sulphonate, 20 mM Tris methane sulphonate and 50 mM sucrose at pH 6.8. Trace amounts of tritiated water were added, along with about 0.5 mM  $^{36}\text{Cl}$  and 0.5 mM pyruvate as Tris salts. In a few experiments,  $^{14}\text{C}$  dextran,  $^{14}\text{C}$  sucrose, or  $^{14}\text{C}$ -ADP were substituted for the  $^{36}\text{Cl}$ . The distribution of solutes between the medium and the mitochondrial pellet was found after separation of the suspension by centrifugation in Coleman Microfuges in tubes pre-loaded with 1.5 M perchloric acid beneath a layer of silicone oil (General Electric Versilube F.50) as described before [5]. Pyruvate was measured by enzymatic assay, phosphate by extraction of phosphomolybdate in isopropyl acetate [10]. ADP entry was measured by incubating the suspension with  $^{14}\text{C}$  ADP in presence of hexokinase (2 units),  $\text{MgCl}_2$  (0.5 mM) and glucose (5 mM) in the medium. The movement was stopped by adding atractylate to 10  $\mu\text{M}$  [11]. The mitochondria were separated as described to find their content of  $^{14}\text{C}$ -labelled nucleotide.

Model experiments to investigate the counter movements of  $\text{Cl}^-$  and  $\text{OH}^-$  were made in the apparatus sketched in fig. 1. The aqueous media in the two U-tubes were in communication only via the oil layer at their common top. The contents of the U's and the floating oil layer were stirred. One U was charged with 3 M KCl buffered with 0.1 M Tris chloride to pH 7.4.

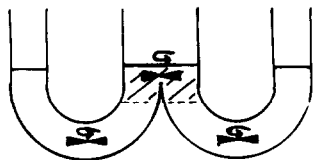


Fig. 1. Sketch of model system used to show the counter transfer of  $\text{Cl}^-$  in one sense and  $\text{OH}^-$  in the other through an interposed 'oil' layer between two aqueous media. One aqueous solution was buffered 3 M KCl, the other was 10 mM Cl-free buffer into which a Radiometer 'pH-Stat' fed Tris base to hold the pH at 7.4. The tin compound was added to the oil layer.

The other U was charged with 10 mM Tris methane sulphonate at pH 7.4; a Radiometer pH stat was set to hold the pH of this latter side constant by addition of 0.1 M Tris base. With an oil made up with 2 parts toluene and 1 part n-butanol, a slow titration rate was observed. This rate and the effects on it of additions of triorgano-tin salts to the oil layer are given in sect. 3.

### 3. Results

#### 3.1. Solute volumes associated with the mitochondrial pellets

Measurements of the amounts of alternative solutes carried down with the pellets into the acid beneath the silicone oil are shown in table 1. When tributyl tin acetate was present at 4 nmole/mg protein and  $\text{Cl}^-$  at 0.35 mM, there was no change of tritiated water, sucrose or dextran spaces, but the calculated Cl 'space' increased to double or more. With more tributyl-tin salt in presence of 0.77 mM Cl the tritiated water space increased slightly (by less than 10%) while the Cl 'space' doubled. The movement of  $\text{Cl}^-$  in these conditions is not accompanied by gross movement of water so it must be inferred that the  $\text{Cl}^-$  is being accumulated in the matrix. It was noted in these experiments that if tributyl-tin salt in excess of 10 nmole/mg were added to suspensions, the particles swelled, even in Cl-free media. This effect was observed both by optical density and packed volume measurements.

Table 1

Solute-accessible volumes carried by the mitochondrial pellet into the acid beneath the oil calculated assuming that there is no change of concentration.

#### Expt. 1 (KCl present at 0.35 mM)

	Volumes occupied by solutes at their external conc. in the pellet (ml/g protein).			
	$\text{T}_2\text{O}$	$\text{Cl}^-$	Dextran	Sucrose
Before tin salt	3.72	3.63	2.76	3.22
After tributyl tin at 4 $\mu\text{mole/g}$	3.71	7 to 10 over 5 min	2.80	3.22

#### Expt. 2 (KCl present at 0.77 mM)

Before tin salt	3.09	3.14	Not meas.	2.68
After tributyl tin at 5.7 $\mu\text{mole/g}$	3.29	6.3 at 6 min	Not meas.	2.62

Medium: As in Methods with KCl additions noted.

Values are means of 3 individual measurements which agree to within  $\pm 0.15$  (S.D.).

#### 3.2. Induced $\text{Cl}^-$ uptakes and their relation to pyruvate accumulation

When the triorgano-tin salts were added at 3–7  $\mu\text{mole/g}$  protein, the Cl content rose over some minutes and the pyruvate content fell (fig. 2). With tributyl tin, which was most used, the uptake was complete within 5 min, and the contents of both  $\text{Cl}^-$  and pyruvate would then remain static or diminish slightly over 10–15 min. The result was obtained whether or not metabolism was inhibited (fig. 2). With triphenyl tin as additive the  $\text{Cl}^-$  uptake more obviously passed through a maximum and then fell, along with the pyruvate. In the results in table 2, a note is made of the times of measurement, and a remark about the constancy or otherwise of the pyruvate and  $\text{Cl}^-$  contents. Table 2 is arranged to show that after the tin salt addition the ratios internal pyruvate/external pyruvate and internal chloride/external chloride become equal. The result is obtained with each of the 6 different tin compounds tested, although the ratios applying to each anion were high in some conditions and low in others.

When chloride ions enter the mitochondria there is not an exactly equivalent displacement of pyruvate, since other anions will also redistribute. When pyruvate is being metabolised the citrate formed tends to accu-

Table 2

Changes in Cl and pyruvate contents of mitochondria in response to additions of triorgano-tin salts to suspensions.

Ref.	Extl. Cl (mM)	Contents ( $\mu$ mole/g) before			Addition and time after taking of samples	Extl. Pyr. (mM)	Contents ( $\mu$ mole/g) after				Ratios	
		Totals Pyr.	Cl	Net Pyr.			Totals Pyr.	Cl	Net Pyr.	Cl	Pyr.	Cl
1	0.35	10.3	0.99	7.06	Trimethyl tin chloride 9 $\mu$ mole/g at 10 and 15 min	1.03	9.1	3.20	6.2	2.21	6.0	6.3
2	0.095	10.0	0.31	6.9	Triethyl tin sulphate 2.5 $\mu$ mole/g at 3.3 and 4.3 min	0.87	8.6	0.94	5.7	0.63	6.5	6.6
3	0.245	13.9	0.68	11.8	Tripropyl tin acetate 7 $\mu$ mole/g at 5 and 6 min	0.68	8.0	2.39	6.1	1.71	9.0	7.0
4	0.295	13.9	0.68	11.8	Tributyl tin acetate 7 $\mu$ mole/g at 6 min	0.72	12.2	3.90	10.2	3.22	14.2	13.1
5	1.34	9.52	5.21	7.17	Tributyl tin acetate 5 $\mu$ mole/g at 6 min	0.51	8.1	20.7	5.87	14.7	11.3	10.9
6	0.245	13.9	0.68	11.8	Triphenyl tin acetate 7 $\mu$ mole/g at 3.6 min (falls later)	0.72	6.7	1.99	4.7	1.31	6.5	5.3
7	0.245	13.9	0.68	11.8	Tricyclohexyl tin methane sulphonate 7 $\mu$ mole/g at 4.5 and 6 min	0.72	9.5	3.40	7.52	2.72	10.7	11.1

Medium:  $\text{Cl}^-$ -free as Methods, plus Tris-pyruvate as noted, Tris-malate about 0.5 mM, Tris-chloride as noted.

Net internal contents in the matrix have been calculated assuming that the initial  $\text{Cl}^-$  content is all carried outside the matrix. Contents are referred to biuret protein. After the tin salts have induced  $\text{Cl}^-$  uptake, the ratios between the net contents and the respective concentrations of  $\text{Cl}^-$  and pyruvate become similar (last two columns).

multate internally and compete out both pyruvate and chloride. As an example the internal citrate rose from 0.6 to 7.0  $\mu$ mole/g protein in a 5 min incubation. The competitive effect of citrate was shown directly by adding it to the incubation medium and measuring the citrate, pyruvate and chloride contents (table 3). Again these anion ratios tended to the same value. Other experiments (not detailed) showed that  $\text{Cl}^-$  and pyruvate accumulation ratios could be diminished by adding more malate, pyruvate or chloride. These data indicate that the accumulated chloride penetrates the same mitochondrial compartments as pyruvate, malate and citrate [see 12, 13].

### 3.3. Expulsion of DNP by $\text{Cl}^-$ when tributyl tin is present

It has been observed [7] that triorgano tin salts will inhibit 2, 4-DNP-stimulated respiration in a chloride-containing medium. One explanation could be that the internal substrate anions are displaced. A second could be that the internal 2, 4-DNP level has been reduced (table 4). Most of the 2, 4-DNP is likely to be undissociated or in another compartment (membrane lipids, for example) which explains the high accumulation ratios. Only the dissociated acid ratio should be compared with other anion ratios.

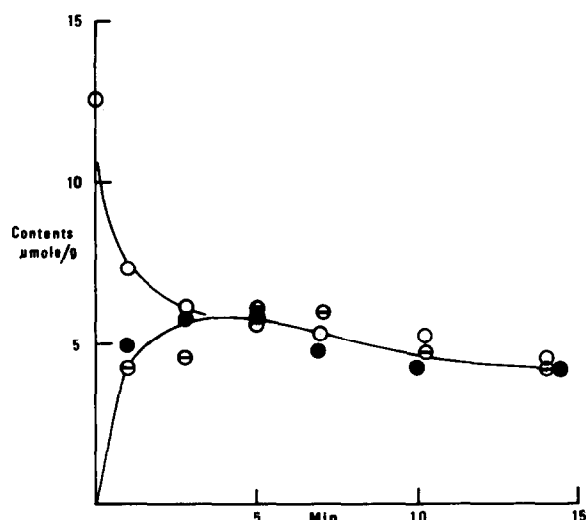


Fig. 2. Changes of mitochondrial pyruvate ( $\circ$ ) and chloride ( $\circ$  and  $\bullet$ ) contents following an addition of tributyl tin acetate to the suspension at  $t = 0$ . The contents are corrected for the amounts carried in the non-specific volume associated with the pellet. The Cl points are from two parallel experiments, one with energy supplied by addition of hydroxy-butyrate and the other with energy supply inhibited. Media: as in Methods, with 0.5 mM pyruvate, 0.5 mM Cl and: (points  $\circ$  and  $\circ$ ) 0.5 mM Na arsenite, 5 mM Tris-hydroxy-butyrate; (points  $\bullet$ ) rotenone 1  $\mu$ g/ml, oligomycin 5  $\mu$ g/ml. Protein 4.5 mg/ml, pH 6.8, 20°.

Table 3

Equilibration between  $\text{Cl}^-$ , pyruvate and added citrate with trimethyl tin acetate present in the system at 8.7 nmole/mg.

Pyruvate			Chloride			Citrate			(Ratio) <sup>1/3</sup>
Exter- nal (mM)	Con- tent ( $\mu$ mole/g)	Ratio	Exter- nal (mM)	Con- tent ( $\mu$ mole/g)	Ratio	Exter- nal (mM)	Con- tent (mM)	Ratio	
1.03	6.18	6.3	0.35	2.21	6.5	None added		—	
1.00	2.9	2.9	0.35	1.34	2.85	0.41	12.4	3.1	
0.97	2.6	2.7	0.35	0.91	2.5	1.30	12.9	2.2	

Medium: As in Methods, pH 6.5 plus 0.1 mM Tris-malate, Tris-pyruvate, citrate and chloride as noted.

Mitochondrial contents have been corrected for the amounts carried down in a volume equal to that occupied by the  $\text{Cl}^-$  before the tin addition. The cube root of the citrate ratio is noted because it would equal the ratio for the singly charged anions when they are equilibrated, provided the solvent volume is 1 ml per g protein.

Table 4

Changes of mitochondrial DNP content in response to addition of tributyl tin acetate to a suspension in a chloride-containing medium.

Experiment	DNP conc. (mM)	Ratio : content ( $\mu$ mole/g)/conc. (mM)		
		Before;	after;	tin salt
		at:		
		(nmole/mg)		
1	0.10	40.3	25.0	: 9
2	0.025	34.3	23.5	: 4.3
3	0.008	53.7	23.0	: 9.7

Medium: 150 mM KCl, Tris-chloride 20 mM, 30 mM sucrose, pH 7.2

The ratio between the content of DNP and the external concentration is noted.

### 3.4. Inhibition of phosphorylation and adenine nucleotide exchange by triethyl and tributyl tin salts

The triorgano-tin salts inhibit phosphorylation even in the absence of chloride [8]. It was confirmed that in a chloride-free medium, addition of tributyl or triethyl-tin salts did not cause a loss of internal substrate; indeed, there was sometimes an increase of 2 to 4  $\mu$ moles of substrate anions and phosphate per g protein. Thus it was possible that access of the adenine nucleotide to the enzymic site or the site itself was being restricted. It has been suggested that the organo-tin salts have an effect similar to oligomycin; however, it seemed possible there was an alternative effect upon the adenine nucleotide translocator. The rate of  $^{14}\text{C}$ -labelled ADP entry was measured in a  $\text{Cl}^-$ -free medium using a high concentration of ADP maintained with hexokinase and glucose. The results for the ratio of internal to external  $^{14}\text{C}$ -labelled nucleotide as a function of time show (fig. 3) that at 6  $\mu$ mole of tributyl or triethyl tin added per g of protein some minutes before the ADP, the organo-tin salts strongly inhibit the uptake. There was little or no effect if the organo-tins were added at 2  $\mu$ mole/g before the ADP or at 6  $\mu$ mole/g at the same time as the ADP. Thus it is likely that these triorgano-tin salts react with the nucleotide exchanger groups.

### 3.5. Model system experiments

The model system was used to find the rate at which alkali had to be added to the weak buffer solution to maintain its pH when 3 M KCl was in the other

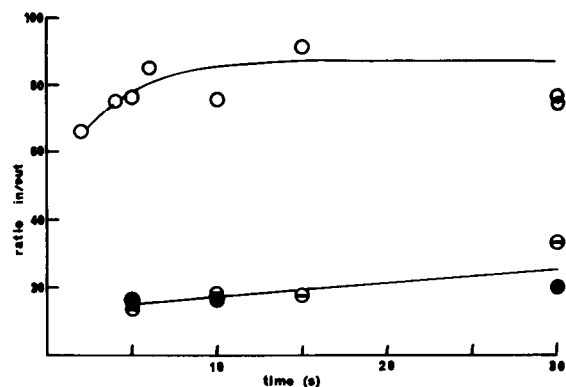


Fig. 3. The effect of triethyl and tributyl tin salts on the labeling of the internal adenine nucleotide pool from  $^{14}\text{C}$ -labelled ADP at  $0.2\ \mu\text{M}$ . The exchange was stopped by atractylate ( $10\ \mu\text{M}$ ) at the different times. Pre-treatment for 10 min at  $0^\circ$  with the tin compounds at  $6\ \text{nmole/mg}$  protein give the results marked  $\ominus$  (triethyl) and  $\bullet$  (tributyl). Untreated controls are marked  $\circ$ . Medium:  $\text{Cl}^-$ -free mixture as in Methods, pH 6.8,  $20^\circ$ .

Table 5

Titration rates measuring  $\text{OH}^-$  disappearance from the low  $\text{Cl}^-$  side of the model apparatus (fig. 1) when  $\text{Cl}^-$  moves into it across the oil layer.

Compound	Amount added in $\mu\text{atoms Sn}$ to 5 ml oil	Rate of titration ( $\mu\text{mole/min}$ )
None		0.10
$(\text{CH}_3)_3\text{SnCl}$	8.5	0.41
	58.5	1.00
$(\text{C}_2\text{H}_5)_3\text{Sn})_2\text{SO}_4$	9.0	0.23
	40.0	0.43
$(\text{C}_3\text{H}_7)_3\text{SnAc}$	10.0	0.62
$(\text{C}_4\text{H}_9)_3\text{SnAc}$	9.0	0.60
	45.0	0.90
$(\text{C}_6\text{H}_5)_3\text{SnAc}$	7.1	0.19
	28.4	0.75
$(\text{C}_6\text{H}_{11})_3\text{SnAc}$	10.0	0.29
	41.0	1.07

arm (table 5). Without any additions to the butanol-toluene there was a slow titration rate. Adding any of the tin compounds listed in table 2 caused an opaque layer to appear at the oil water interface. No absolute significance is attached to the observed rates because inadequate stirring and the plating phenomenon will

have affected them. The point is that the compounds have comparable abilities to mediate the  $\text{Cl}^-$ -for- $\text{OH}^-$  exchange through a non-aqueous solvent.

#### 4. Discussion

Our results show that triorgano tin salts tested in table 2 have more than one action on anion permeability. The mediation of  $\text{Cl}^-$  permeation, whatever its mechanism, allows the  $\text{Cl}^-$  to equilibrate with the pyruvate. Whether or not the two ions are at electrochemical equilibrium cannot be decided but they are equilibrated between themselves. Earlier results [10, 12, 13] suggest that pyruvate, malate and citrate are similarly equilibrated. If we accept that a free exchange of  $\text{OH}^-$ -for- $\text{Cl}^-$  is provided by the tin compounds when they dissolve in the membrane, then it would follow that the equilibrium holds between the anions and  $\text{OH}^-$  and implicitly  $\text{H}^+$  also. As the accumulation occurs after respiration is inhibited (fig. 2), it is necessary to regard the inside as having been 'charged up' during previous metabolic activity [13, 14]. Our finding that the adenine nucleotide exchanger is inhibited extends the known toxic effects of the triorgano tin compounds. It does not preclude an additional oligomycin-like effect [15, 16]. It may be useful in providing an additional agent to atractylate for titrating the adenine exchanger groups.

#### Acknowledgements

We have to thank Dr. J.E. Cremer for gifts of the organo-tin salts and  $^{14}\text{C}$ -labelled DNP, and Dr. M. Rose for communicating to us results with DNP similar to table 4. We are indebted to the Wellcome Trust for a studentship for J.A.B. and for support from the M.R.C.

#### References

- [1] J.L. Gamble, J. Biol. Chem. 240 (1965) 2668.
- [2] E.J. Harris and K. van Dam, Biochem. J. 106 (1968) 759.
- [3] E.J. Harris and J.R. Manger, Biochem. J. 109 (1969) 239.
- [4] K. van Dam and C.S. Tsou, in: Energy level and metabolic control in mitochondria, eds. S. Papa, J. Tager, E. Quagliariello and E.C. Slater (Adriatica Ed., Bari, 1969) p. 21.

- [5] J.R. Manger, FEBS Letters 5 (1969) 331.
- [6] M.J. Selwyn, A.P. Dawson, M. Stockdale and N. Gains, European J. Biochem. 14 (1970) 120.
- [7] W.N. Aldridge, Biochem. J. 69 (1958) 367.
- [8] M.S. Rose and W.N. Aldridge, Biochem. J. 127 (1972) 51.
- [9] E.J. Harris, C. Tate, J.A. Bangham and J.R. Manger, J. Bioenergetics 5 (1971) 221.
- [10] E.J. Harris, FEBS Letters 11 (1971) 225.
- [11] H.W. Heldt, in: Mitochondrial Structure and Compartmentation, Eds. E. Quagliariello, S. Papa, E.C. Slater and J.M. Tager, (Adriatica Ed., Bari, 1967) p. 260.
- [12] E.J. Harris and C. Berent, FEBS Letters 10 (1970) 7.
- [13] E.J. Harris and J.A. Bangham, J. Mem. Biol. 9 (1972) 141.
- [14] F. Palmieri, E. Quagliariello and M. Klingenberg. European J. Biochem. 17 (1969) 230.
- [15] M.J. Selwyn, S.J. Dunnnett, R.D. Philo and A.P. Dawson, Biochem. J. 127 (1972) 66 p.
- [16] Y. Kagawa and E. Racker, J. Biol. Chem. 241 (1966) 2461.