

# Intracellular $\text{Ca}^{2+}$ -calmodulin system involved in the palytoxin-induced $\text{K}^+$ release from rabbit erythrocytes

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Palytoxin (PTX) caused  $\text{K}^+$  release from rabbit erythrocytes which was dependent on the concentrations of extracellular  $\text{Ca}^{2+}$  and PTX. In a  $\text{Ca}^{2+}$ -free solution, PTX still caused a slow  $\text{K}^+$  release. An intracellular  $\text{Ca}^{2+}$  antagonist, TMB-8, an intracellular  $\text{Ca}^{2+}$  chelator, quin 2, and calmodulin inhibitors, prenylamine, W-7 and W-5, inhibited the PTX-induced  $\text{K}^+$  release in a  $\text{Ca}^{2+}$ -free solution. These results suggest that the PTX-induced  $\text{K}^+$  release is dependent on the process including intracellular  $\text{Ca}^{2+}$  and calmodulin.

*Palytoxin       $\text{K}^+$  release       $\text{Ca}^{2+}$       Calmodulin*

## 1. INTRODUCTION

Palytoxin (PTX), isolated from marine coelenterates of the genus *Palythoa*, is one of the most poisonous animal toxins [1]. It has been reported that PTX increases  $\text{K}^+$  efflux and  $\text{Na}^+$  influx in erythrocytes, smooth muscle and pheochromocytoma cells [2–5]. In the preceding paper, we suggested that the PTX-induced  $\text{K}^+$  release from erythrocytes is mediated by a  $\text{Ca}^{2+}$ -calmodulin system [6]. Since Chhatwal et al. [7] suggested that external  $\text{Ca}^{2+}$  is not essential for the PTX-induced  $\text{K}^+$  release, we examined the role of intracellular  $\text{Ca}^{2+}$  on the PTX-induced  $\text{K}^+$  release from rabbit erythrocytes using an intracellular Ca antagonist, TMB 8 [8], an intracellular Ca chelator, quin 2 [9], and calmodulin inhibitors, prenylamine [10], W-7 and W-5 [11].

## 2. MATERIALS AND METHODS

Rabbit erythrocytes were prepared as reported in [5]. Loss of  $\text{K}^+$  from erythrocytes was continuously determined at 37°C using a  $\text{K}^+$ -selective electrode (Philips, IS 561 K). Since calmodulin inhibitors changed the sensitivity of the  $\text{K}^+$ -selective

electrode, erythrocytes tested with these compounds were incubated with PTX for 18 min at 37°C, the reaction was stopped by a 2 min centrifugation at  $650 \times g$  at 25°C and the  $\text{K}^+$  content of the supernatant was measured with a Hitachi type 208 flame photometer. The amount of maximum releasable  $\text{K}^+$  was determined by adding saponin ( $10^{-5}$  g/ml) at the end of each experiment. Physiological salt solution (PSS) contained (mM): 136.9 NaCl, 1.0  $\text{CaCl}_2$ , 1.0  $\text{MgCl}_2$ , 5.5 glucose and 10 Hepes at pH 7.4.  $\text{Ca}^{2+}$ -free solution (below  $5 \times 10^{-7}$  M  $\text{Ca}^{2+}$ ) was made by adding EGTA.

PTX isolated from *P. tuberculosa* (kindly donated by Dr Y. Hirata, Meijo University, Nagoya) was dissolved in a solution containing 0.1% bovine serum albumin and 2 mM Hepes (pH 7.0) at  $10^{-4}$  M, stored at  $-20^\circ\text{C}$  and diluted just before use. 2-[(2-Bis[carboxymethyl]amino-5-methylphenoxy)methyl]-6-methoxy-8-bis[carboxymethyl]aminoquinolinetetrakis[acetoxymethyl] ester (quin 2-AM, Sigma) and prenylamine (Hoechst) were dissolved in dimethyl sulfoxide (DMSO). Nigericin (Calbiochem) was dissolved in ethanol. 8-(*N,N*-Diethylamino)octyl 3,4,5-trimethoxybenzoate (TMB-8, generous gifts from Tanabe), *N*-(6-aminohexyl)-5-chloro-1-naphthalenesulfon-

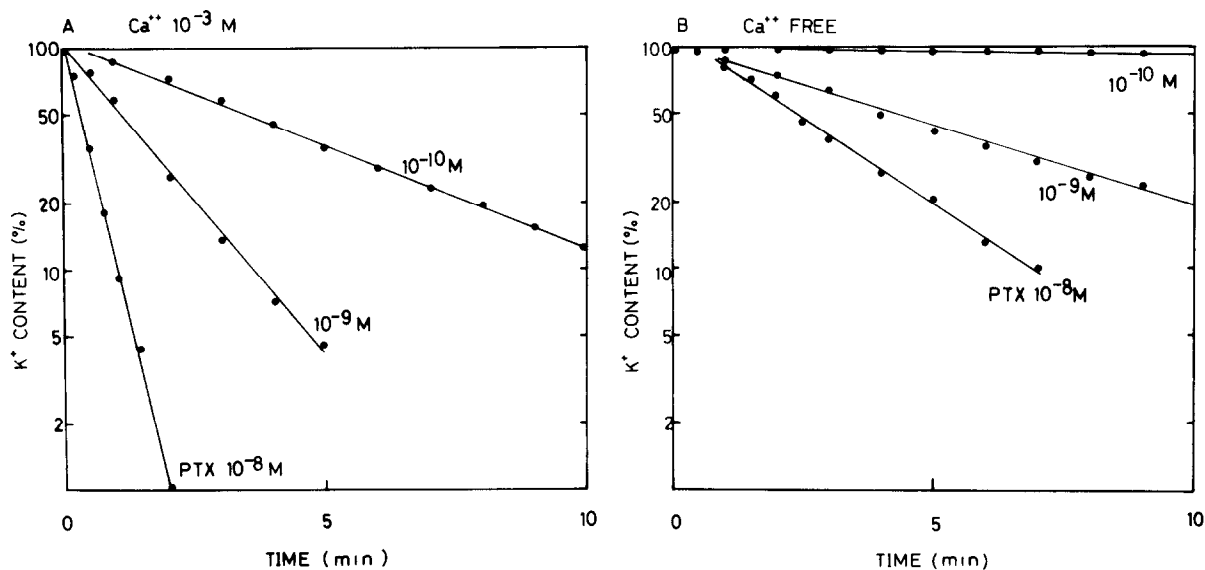


Fig.1. PTX-induced K<sup>+</sup> release in the presence or absence of extracellular Ca<sup>2+</sup>. PTX was added after a 10 min preincubation period at 37°C in the presence of 10<sup>-3</sup> M Ca<sup>2+</sup> (A) or in the absence of Ca<sup>2+</sup> (B). K<sup>+</sup> release was measured with a K<sup>+</sup>-selective electrode. Abscissa, time (min); ordinate, K<sup>+</sup> content in erythrocytes (%). 100% represents the amount of total releasable K<sup>+</sup> measured by the addition of 10<sup>-5</sup> g/ml saponin.

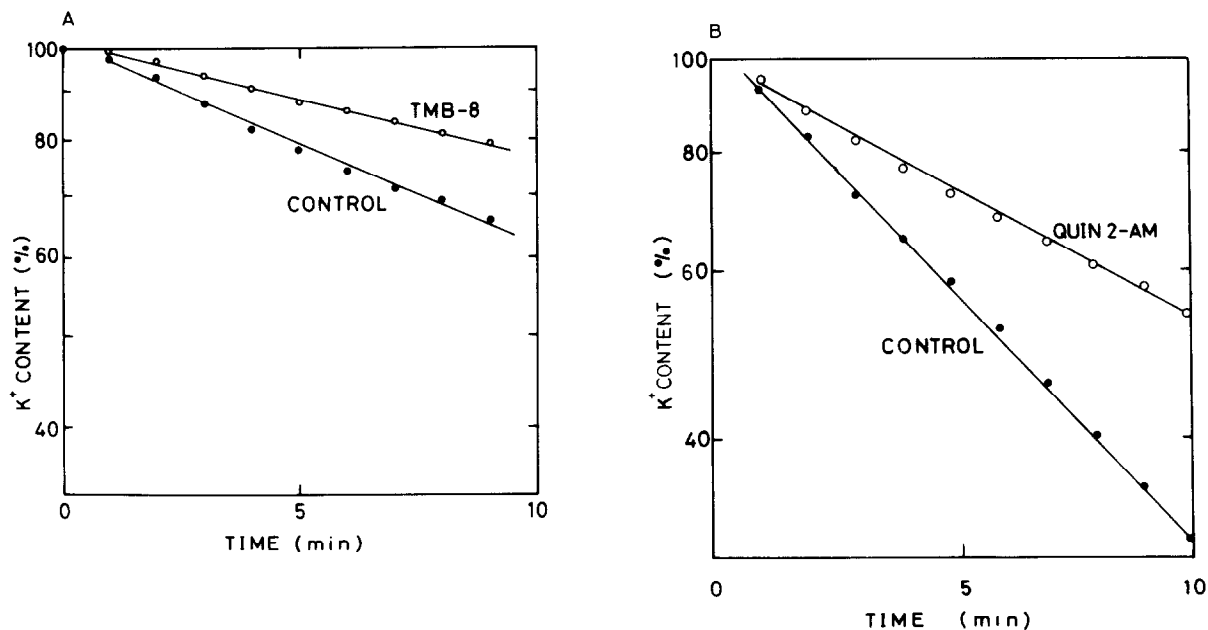


Fig.2. Effects of TMB-8 and quin 2-AM on the PTX-induced K<sup>+</sup> release in a Ca<sup>2+</sup>-free solution. (A) 10<sup>-4</sup> M TMB-8 was added 10 min before the application of 10<sup>-9</sup> M PTX. (B) Erythrocytes were preincubated with a Ca<sup>2+</sup>-free solution containing 10<sup>-4</sup> M quin 2-AM at 37°C for 1 h, washed 3 times with a Ca<sup>2+</sup>-free solution and then 10<sup>-9</sup> M PTX was applied. In the control experiment, erythrocytes were treated with 1% DMSO alone. Abscissa, time (min); ordinate, K<sup>+</sup> content in erythrocytes (%).

amide (W-7), *N*-(6-aminohexyl)-1-naphthalenesulfonamide (W-5) (both from Rikaken) and saponin (ICN) were dissolved in PSS.

### 3. RESULTS

PTX caused  $K^+$  release from erythrocytes in the presence of  $10^{-3}$  M  $Ca^{2+}$  (fig.1A). The rate constants of  $K^+$  release due to  $10^{-10}$ ,  $10^{-9}$  and  $10^{-8}$  M PTX were 0.21, 0.66 and  $2.34 \text{ min}^{-1}$ , respectively. The rate of  $K^+$  release was markedly retarded in a  $Ca^{2+}$ -free solution (fig.1B); the rate constants due to  $10^{-10}$ ,  $10^{-9}$  and  $10^{-8}$  M PTX were 0.0075, 0.17 and  $0.36 \text{ min}^{-1}$ , respectively. In contrast, the rates of  $K^+$  release induced by saponin ( $10^{-5}$  g/ml) and nigericin ( $10^{-9}$  M) were independent of extracellular  $Ca^{2+}$  concentrations (not shown).

Fig.2 shows the effects of TMB-8 ( $10^{-4}$  M) and quin 2-AM ( $10^{-4}$  M) on the rate of  $K^+$  release induced by PTX ( $10^{-9}$  M) in a  $Ca^{2+}$ -free solution. Although the control rate constants were slightly different, both TMB-8 and quin 2-AM decreased them to 56.8 and 51.7%, respectively.

Fig.3 shows the effects of prenylamine, W-7 and W-5 in a  $Ca^{2+}$ -free solution. The  $K^+$  release due to

PTX ( $10^{-9}$  M) was inhibited to 10.6% by prenylamine, 13.3% by W-7 and 66.6% by W-5.

### 4. DISCUSSION

In the present experiment, we confirmed the result by Chhatwal et al. [7] that the PTX-induced  $K^+$  release is not completely inhibited in the absence of extracellular  $Ca^{2+}$ . However, this result does not necessarily indicate that  $Ca^{2+}$  is not involved in the effect of PTX since cellular bound  $Ca^{2+}$  in erythrocytes is resistant to the incubation with a  $Ca^{2+}$ -free solution [12]. In fact, the PTX-induced  $K^+$  release in a  $Ca^{2+}$ -free solution was inhibited by an intracellular Ca antagonist, TMB-8, and an intracellular  $Ca^{2+}$  chelator, quin 2. Further, the calmodulin inhibitors, prenylamine and W-7, inhibited the PTX-induced  $K^+$  release in the absence of extracellular  $Ca^{2+}$  whereas W-5, an analog of W-7 with lower affinity to calmodulin [11], showed smaller inhibition. Similar inhibitory potencies of these calmodulin inhibitors were reported in the presence of extracellular  $Ca^{2+}$  [6]. These results suggest that PTX mobilizes cellular bound  $Ca^{2+}$  and activates a  $Ca^{2+}$ -calmodulin system to induce  $K^+$  release from rabbit erythrocytes.

### ACKNOWLEDGEMENTS

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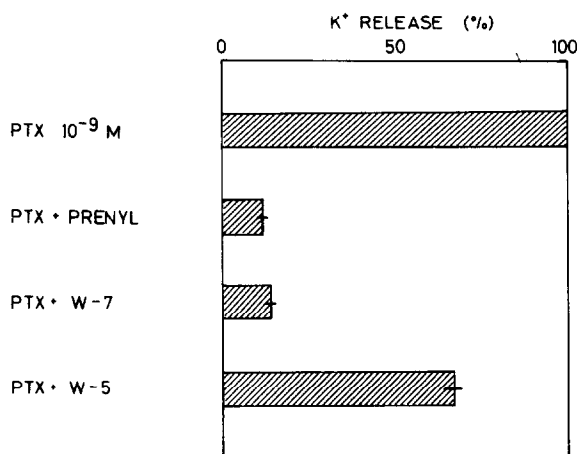


Fig.3. Inhibitory effects of prenylamine, W-7 and W-5 on the PTX-induced  $K^+$  release in the absence of extracellular  $Ca^{2+}$ . Erythrocytes were preincubated for 10 min with a  $Ca^{2+}$ -free solution with or without  $3 \times 10^{-5}$  M prenylamine,  $10^{-4}$  M W-7 or  $10^{-4}$  M W-5. The amount of  $K^+$  released during a 20 min PTX incubation period was measured by flame photometry. Mean  $\pm$  SE of 4 experiments is shown.

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